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I/85942 A

(54) Title: CYTOSKELETON-ASSOCIATED PROTEINS

(57) Abstract: The invention provides human cytoskeleton-associated proteins (CYSKP) and polynucleotides which identify and encode CYSKP. The invention also provides expression vectors, host cells, antibodies, agonists, and antagonists. The invention also provides methods for diagnosing, treating, or preventing disorders associated with aberrant expression of CYSKP.

# CYTOSKELETON-ASSOCIATED PROTEINS

# , TECHNICAL FIELD

This invention relates to nucleic acid and amino acid sequences of cytoskeleton-associated proteins and to the use of these sequences in the diagnosis, treatment, and prevention of cell proliferative, autoimmune/inflammatory, vesicle trafficking, neurological, cell motility, reproductive, and muscle disorders, and in the assessment of the effects of exogenous compounds on the expression of nucleic acid and amino acid sequences of cytoskeleton-associated proteins.

#### BACKGROUND OF THE INVENTION

The cytoskeleton, a cytoplasmic system of protein fibers, mediates cell shape, structure, and movement. The cytoskeleton supports the cell membrane and forms tracks along which organelles and other elements move in the cytosol. The cytoskeleton is a dynamic structure that allows cells to adopt various shapes and to carry out directed movements. Additionally, molecules can be sequestered to a specific cellular location through interaction with cytoskeleton associated proteins. Major cytoskeletal fibers are the microfilaments, the microtubules, and the intermediate filaments. Motor proteins, including myosin, dynein, and kinesin, drive movement of, or along, the fibers. Accessory or associated proteins modify the structure or activity of the fibers while cytoskeletal membrane anchors connect the fibers to the cell membrane. Other proteins associated with the cytoskeleton have roles in processes such as secretion and intracellular signaling. (The cytoskeleton is reviewed in Lodish, H. et al. (1995) Molecular Cell Biology Scientific American Books, New York NY.)

#### Microtubules and Associated Proteins

#### **Tubulins**

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Microtubules, cytoskeletal fibers with a diameter of 24 nm, have multiple roles in the cell. Bundles of microtubules form cilia and flagella, which are whip-like extensions of the cell membrane that are necessary for sweeping materials across an epithelium and for swimming of sperm, respectively. Marginal bands of microtubules in red blood cells and platelets are important for these cells' pliability. Organelles, membrane vesicles, and proteins are transported in the cell along tracks of microtubules. For example, microtubules run through nerve cell axons, allowing bi-directional transport of materials and membrane vesicles between the cell body and the nerve terminal. Failure to supply the nerve terminal with these vesicles blocks the transmission of neural signals. Microtubules, in the form of the spindle, are also critical to chromosomal movement during cell division. Both stable and short-lived populations of microtubules exist in the cell.

Microtubules are a polymer of GTP-binding tubulin protein subunits. Each subunit is a

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heterodimer of  $\alpha$ - and  $\beta$ - tubulin, multiple isoforms of which exist. Alpha-tubulin undergoes a number of post-translational modifications, including acetylation, polyglutamylation, truncation of two amino acids (forming  $\Delta 2$  tubulin), and tyrosination. In some cases, these modifications can affect microtubule stability. The hydrolysis of GTP is linked to the addition of tubulin subunits at the end of a microtubule. The subunits interact head to tail to form protofilaments; the protofilaments interact side to side to form a microtubule. A microtubule is polarized, one end ringed with  $\alpha$ -tubulin and the other with  $\beta$ -tubulin, and the two ends differ in their rates of assembly. Each microtubule is generally composed of 13 protofilaments although 11 or 15 protofilament-microtubules are sometimes found. Cilia and flagella contain doublet microtubules. Microtubules grow from specialized structures known as centrosomes or microtubule-organizing centers (MTOCs). MTOCs may contain one or two centrioles, which are pinwheel arrays of triplet microtubules. The basal body, the organizing center located at the base of a cilium or flagellum, contains one centriole.  $\gamma$ - tubulin present in the MTOC is important for nucleating the polymerization of  $\alpha$ - and  $\beta$ - tubulin heterodimers but does not polymerize into microtubules. The protein pericentrin is found in the MTOC and has a role in microtubule assembly.

# Microtubule-Associated Proteins

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Microtubule-associated proteins (MAPs) have roles in the assembly and stabilization of microtubules. One major family of MAPs, assembly MAPs, can be identified in neurons as well as non-neuronal cells. Assembly MAPs are responsible for cross-linking microtubules in the cytosol. These MAPs are organized into two domains: a basic microtubule-binding domain and an acidic projection domain. The projection domain is the binding site for membranes, intermediate filaments, or other microtubules. Based on sequence analysis, assembly MAPs can be further grouped into two types: Type I and Type II.

Type I MAPs, which include MAP1A and MAP1B, are large, filamentous molecules that copurify with microtubules and are abundantly expressed in brain and testis. They contain several repeats of a positively-charged amino acid sequence motif that binds and neutralizes negatively charged tubulin, leading to stabilization of microtubules. MAP1A and MAP1B are each derived from a single precursor polypeptide that is subsequently proteolytically processed to generate one heavy chain and one light chain.

Another light chain, LC3, is a 16.4 kDa molecule that binds MAP1A, MAP1B, and microtubules. It is suggested that LC3 is synthesized from a source other than the MAP1A or MAP1B transcripts, and the expression of LC3 may be important in regulating the microtubule binding activity of MAP1A and MAP1B during cell proliferation (Mann, S. S. et al. (1994) J. Biol. Chem. 269:11492-11497).

Type II MAPs, which include MAP2a, MAP2b, MAP2c, MAP4, and Tau, are characterized by three to four copies of an 18-residue sequence in the microtubule-binding domain. MAP2a, MAP2b, and MAP2c are found only in dendrites, MAP4 is found in non-neuronal cells, and Tau is found in axons and dendrites of nerve cells. Alternative splicing of the Tau mRNA leads to the existence of multiple forms of Tau protein. Tau phosphorylation is altered in neurodegenerative disorders such as Alzheimer's disease, Pick's disease, progressive supranuclear palsy, corticobasal degeneration, and familial frontotemporal dementia and Parkinsonism linked to chromosome 17. The altered Tau phosphorylation leads to a collapse of the microtubule network and the formation of intraneuronal Tau aggregates (Spillantini, M.G. and Goedert, M. (1998) Trends Neurosci. 21:428-433).

Microtubule stability may also be regulated by severing the microtubule along its length. The protein katanin, a member of the AAA adenosine triphosphatase (ATPase) superfamily, uses ATP hydrolysis energy to sever and disassemble stable microtubules. Katanin may play a role in releasing microtubules from centrosomes, regulating assembly of the mitotic spindle, and accelerating microtubule turnover during cell cycle transitions (Hartman, J.J. and Vale, R.D. (1999) Science 286:782-785).

Microtubular aggregates are associated with several disorders. An extraskeletal myxoid chondrosarcoma tumor from human contained parallel arrays of microtubules within the rough endoplasmic reticulum (Suzuki, T. et al. (1988) J. Pathol. 156:51-57). Microtubular aggregates were also found in hepatocytes from chimpanzees infected with hepatitis C virus. Monoclonal antibodies prepared to these aggregates detect a protein called p44 (or microtubular aggregates protein) (Maeda, T. et al. (1989) J. Gen. Virol. 70:1401-1407). A human homolog of p44 is inducible by interferon- $\alpha$  and interferon- $\beta$ , but not by interferon- $\gamma$ . p44 protein may be a mediator in the antiviral action of interferon (Kitamura, A. et al. (1994) Eur. J. Biochem. 224:877-883).

#### Dynein-related Motor Proteins

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Dyneins are (-) end-directed motor proteins which act on microtubules. Two classes of dyneins exist, cytosolic and axonemal. Cytosolic dyneins are responsible for translocation of materials along cytoplasmic microtubules; for example, transport from the nerve terminal to the cell body and transport of endocytic vesicles to lysosomes. Cytoplasmic dyneins are also reported to play a role in mitosis. Axonemal dyneins are responsible for the beating of flagella and cilia. Dynein on one microtubule doublet walks along the adjacent microtubule doublet. This sliding force produces bending forces that cause the flagellum or cilium to beat. Dyneins have a native mass between 1000 and 2000 kDa and contain either two or three force-producing heads driven by the hydrolysis of ATP. The heads are linked via stalks to a basal domain which is composed of a highly variable number of accessory intermediate and light chains.

#### Kinesin-related Motor Proteins

Kinesins are (+) end-directed motor proteins which act on microtubules. The prototypical kinesin molecule is involved in the transport of membrane-bound vesicles and organelles. This function is particularly important for axonal transport in neurons. Kinesin is also important in all cell types for the transport of vesicles from the Golgi complex to the endoplasmic reticulum. This role is critical for maintaining the identity and functionality of these secretory organelles.

Kinesins define a ubiquitous, conserved family of over 50 proteins that can be classified into at least 8 subfamilies based on primary amino acid sequence, domain structure, velocity of movement, and cellular function. (Reviewed in Moore, J.D. and S.A. Endow (1996) Bioessays 18:207-219; and Hoyt, A.M. (1994) Curr. Opin. Cell Biol. 6:63-68.) The prototypical kinesin molecule is a heterotetramer comprised of two heavy polypeptide chains (KHCs) and two light polypeptide chains (KLCs). The KHC subunits are typically referred to as "kinesin." KHC is about 1000 amino acids in length, and KLC is about 550 amino acids in length. Two KHCs dimerize to form a rod-shaped molecule with three distinct regions of secondary structure. At one end of the molecule is a globular motor domain that functions in ATP hydrolysis and microtubule binding. Kinesin motor domains are highly conserved and share over 70% identity. Beyond the motor domain is an  $\alpha$ -helical coiled-coil region which mediates dimerization. At the other end of the molecule is a fan-shaped tail that associates with molecular cargo. The tail is formed by the interaction of the KHC C-termini with the two KLCs.

Members of the more divergent subfamilies of kinesins are called kinesin-related proteins (KRPs), many of which function during mitosis in eukaryotes (Hoyt, <u>supra</u>). Some KRPs are required for assembly of the mitotic spindle. <u>In vivo</u> and <u>in vitro</u> analyses suggest that these KRPs exert force on microtubules that comprise the mitotic spindle, resulting in the separation of spindle poles. Phosphorylation of KRP is required for this activity. Failure to assemble the mitotic spindle results in abortive mitosis and chromosomal aneuploidy, the latter condition being characteristic of cancer cells. In addition, a unique KRP, centromere protein E, localizes to the kinetochore of human mitotic chromosomes and may play a role in their segregation to opposite spindle poles.

#### Microfilaments and Associated Proteins

#### **Actins**

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Microfilaments, cytoskeletal filaments with a diameter of 7-9 nm, are vital to cell locomotion, cell shape, cell adhesion, cell division, and muscle contraction. Assembly and disassembly of the microfilaments allow cells to change their morphology. Microfilaments are the polymerized form of actin, the most abundant intracellular protein in the eukaryotic cell. Human cells contain six isoforms of actin. The three  $\alpha$ -actins are found in different kinds of muscle, nonmuscle  $\beta$ -actin and nonmuscle  $\gamma$ -actin are found in nonmuscle cells, and another  $\gamma$ -actin is found in intestinal smooth muscle cells. G-

actin, the monomeric form of actin, polymerizes into polarized, helical F-actin filaments, accompanied by the hydrolysis of ATP to ADP. Actin filaments associate to form bundles and networks, providing a framework to support the plasma membrane and determine cell shape. These bundles and networks are connected to the cell membrane. In muscle cells, thin filaments containing actin slide past thick filaments containing the motor protein myosin during contraction. Other actin-related filaments are not part of the actin cytoskeleton, but rather associate with microtubules and dyenin.

#### **Actin-Associated Proteins**

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Actin-associated proteins have roles in cross-linking, severing, and stabilization of actin filaments and in sequestering actin monomers. Several of the actin-associated proteins have multiple functions. Bundles and networks of actin filaments are held together by actin cross-linking proteins. These proteins have two actin-binding sites, one for each filament. Short cross-linking proteins promote bundle formation while longer, more flexible cross-linking proteins promote network formation. Calmodulin-like calcium-binding domains in actin cross-linking proteins allow calcium regulation of cross-linking. Group I cross-linking proteins have unique actin-binding domains and include the 30 Kd protein, EF-1a, fascin, and scruin. Group II cross-linking proteins have a 7,000-MW actin-binding domain and include villin and dematin. Group III cross-linking proteins have pairs of a 26,000-MW actin-binding domain and include alpha-actinin, fimbrin, spectrin, dystrophin, ABP 120, and filamin.

Severing proteins regulate the length of actin filaments by breaking them into short pieces or by blocking their ends. Severing proteins include gCAP39, severin (fragmin), gelsolin, and villin. Capping proteins can cap the ends of actin filaments, but cannot break filaments. Capping proteins include CapZ, tropomodulin, and tensin.

Tensin, which is found in focal adhesions, also crosslinks actin filaments. Integrin activation by the extracellular matrix leads to the phosphorylation of tensin on tyrosine, serine, and threonine residues; this phosphorylation also occurs in cells transformed with oncogenes. Tensin has an SH2 domain and may bind to other tyrosine-phosphorylated proteins. (Lo, S.H. et al. (1997) J. Cell Biol. 136:1349-1361.) The N-terminus of tensin contains a region homologous to the catalytic domain of a putative tyrosine phosphatase (PTP) from Saccharomyces cerevisiae. This PTP domain in tensin may mediate binding interactions with phosphorylated polypeptides (Haynie, D.T. and Ponting, C.P. (1996) Protein Sci. 5:2643-2646). Mice which lack the tensin gene have kidney abnormalities, indicating that the loss of tensin leads to weakening of focal adhesions in the kidney (Lo, supra).

The proteins thymosin and profilin sequester actin monomers in the cytosol, allowing a pool of unpolymerized actin to exist. Profilin may also stimulate F-actin formation by effectively lowering the critical concentration required for actin monomer addition (Gertler, F.B. et al. (1996) Cell 87:227-239).

The actin-associated proteins tropomyosin, troponin, and caldesmon regulate muscle

contraction in response to calcium. The tropomyosin proteins, found in muscle and nonmuscle cells, are  $\alpha$ -helical and form coiled-coil dimers. Striated muscle tropomyosin mediates the interactions between the troponin complex and actin, regulating muscle contraction (PROSITE PDOC00290 Tropomyosins signature). The troponin complex is composed of troponin-T, troponin-I, and troponin-C. Troponin-T binds tropomyosin, linking troponin-I and troponin-C to tropomyosin.

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Many proteins involved in the regulation of actin assembly have characteristic protein-protein interaction domains, such as for example the calponin homology (CH) domain found in actin cross-linking proteins including alpha-actinin, spectrin, and fimbrin. Other proteins which are localized primarily in focal adhesions, macromolecular complexes which mediate the contact between extracellular matrix receptors and the cytoskeleton, contain protein-protein interaction motifs known as LIM domains. For example, zyxin is a protein that plays a role in the spatial control of actin assembly and contains three tandem LIM domains. Zyxin also interacts with alpha-actinin through its proline rich N-terminus (Beckerle, M. C. (1997) BioEssays 19:949-957).

Cytoskeletal proteins are implicated in several diseases. Pathologies such as muscular dystrophy, nephrotic syndrome, and dilated cardiomyopathy have been associated with differential expression of alpha-actinin-3 (Vainzof, M. et al. (1997) Neuropediatrics 28:223-228; Smoyer, W.E. and Mundel, P. (1998) J. Mol. Med. 76:172-183; and Sussman, M.A. et al. (1998) J. Clin. Invest. 101:51-61). Alpha-actinin and several MAPs are present in Hirano bodies, which are observed more frequently in the elderly and in patients with neurodegenerative diseases such as Alzheimer's disease (Maciver, S.K. and Harrington, C.R. (1995) Neuroreport. 6:1985-1988). Actinin-4, a novel actin-bundling protein, appears to be associated with the cell motility of metastatic cancer cells. Other disease associations include premature chromosome condensation which is frequently observed in dividing cells from tumor tissue (Murnane, J.P. (1995) Cancer Metastasis Rev. 14:17-29) and the significant roles of axonemal and assembly MAPs in viral pathogenesis (Sodeik, B. et al. (1997) J. Cell Biol. 136:1007-1021).

# Intermediate Filaments and Associated Proteins

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Intermediate filaments (IFs) are cytoskeletal fibers with a diameter of 10 nm, intermediate between that of microfilaments and microtubules. They serve structural roles in the cell, reinforcing cells and organizing cells into tissues. IFs are particularly abundant in epidermal cells and in neurons.

IFs are extremely stable, and, in contrast to microfilaments and microtubules, do not function in cell motility. IF proteins include acidic keratins, basic keratins, desmin, glial fibrillary acidic protein, vimentin, peripherin, neurofilaments, nestin, and lamins.

IFs have a central  $\alpha$ -helical rod region interrupted by short nonhelical linker segments. The rod region is bracketed, in most cases, by non-helical head and tail domains. The rod regions of intermediate filament proteins associate to form a coiled-coil dimer. A highly ordered assembly process leads from the dimers to the IFs. Neither ATP nor GTP is needed for IF assembly, unlike that of microfilaments and microtubules.

IF-associated proteins (IFAPs) mediate the interactions of IFs with one another and with other cell structures. IFAPs cross-link IFs into a bundle, into a network, or to the plasma membrane, and may cross-link IFs to the microfilament and microtubule cytoskeleton. Microtubules and IFs in particular are closely associated. IFAPs include BPAG1, plakoglobin, desmoplakin I, desmoplakin II, plectin, ankyrin, filaggrin, and lamin B receptor.

The N-terminal portion of ankyrin consists of a repeated 33-amino acid motif, the ankyrin repeat, which is involved in specific protein-protein interactions. Variable regions within the motif are responsible for specific protein binding, such that different ankyrin repeats are involved in binding to tubulin, anion exchange protein, voltage-gated sodium channel, Na<sup>+</sup>/K<sup>+</sup>-ATPase, and neurofascin. The ankyrin motif is also found in transcription factors, such as NF-κ-B, and in the yeast cell cycle proteins CDC10, SW14, and SW16. Proteins involved in tissue differentiation, such as <u>Drosophila</u> Notch and <u>C. elegans</u> LIN-12 and GLP-1, also contain ankyrin-like repeats. Lux et al. (1990; Nature 344:36-42) suggest that ankyrin-like repeats function as 'built-in' ankyrins and form binding sites for integral membrane proteins, tubulin, and other proteins.

# Cytoskeletal-Membrane Anchors

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Cytoskeletal fibers are attached to the plasma membrane by specific proteins. These attachments are important for maintaining cell shape and for muscle contraction. In erythrocytes, the spectrin-actin cytoskeleton is attached to cell membrane by three proteins, band 4.1, ankyrin, and adducin. Defects in this attachment result in abnormally shaped cells which are more rapidly degraded by the spleen, leading to anemia. In platelets, the spectrin-actin cytoskeleton is also linked to the membrane by ankyrin; a second actin network is anchored to the membrane by filamin. In muscle cells the protein dystrophin links actin filaments to the plasma membrane; mutations in the dystrophin gene lead to Duchenne muscular dystrophy. In adherens junctions and adhesion plaques the peripheral membrane proteins  $\alpha$ -actinin and vinculin attach actin filaments to the cell membrane.

IFs are also attached to membranes by cytoskeletal-membrane anchors. The nuclear lamina is attached to the inner surface of the nuclear membrane by the lamin B receptor. Vimentin IFs are

attached to the plasma membrane by ankyrin and plectin. Desmosome and hemidesmosome membrane junctions hold together epithelial cells of organs and skin. These membrane junctions allow shear forces to be distributed across the entire epithelial cell layer, thus providing strength and rigidity to the epithelium. IFs in epithelial cells are attached to the desmosome by plakoglobin and desmoplakins. The proteins that link IFs to hemidesmosomes are not known. Desmin IFs surround the sarcomere in muscle and are linked to the plasma membrane by paranemin, synemin, and ankyrin.

Proteins of the Erythrocyte Membrane Skeleton

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Distribution of oxygen throughout the vertebrate body is effected by red blood cells (erythrocytes). Oxygen diffuses from surrounding water or from the atmosphere through either gill epithelium or pulmonary epithelial type I cells. Oxygen then diffuses through the blood capillary endothelium directly to the blood circulatory system and through the erythrocyte membrane and is stored as soluble oxyhemoglobin in the cytoplasm. Oxygen is released from hemoglobin at sites throughout the organism and diffuses out from the erythrocyte to other target cells. The structure of the erythrocyte membrane as well as that of other non-erythrocyte cells must be maintained to enable efficient diffusion of oxygen to intracellular compartments.

The erythrocyte membrane is comprised of i) a cholesterol-rich phospholipid bilayer in which many trans-bilayer proteins are embedded, ii) external glycosylphosphatidylinositol-anchored proteins (GPI-proteins), and iii) the erythrocyte or membrane skeleton that laminates the inner surface of the bilayer. The trans-bilayer proteins include anion exchangers, glycophorins, glucose transporters, and a variety of cation transporters and pumps. The erythrocyte GPI-proteins include acetylcholinesterase and decay-accelerating factor (CD 55). The skeletal proteins are organized on the cortical, or cytoplasmic, face of the plasma membrane. These proteins include protein 4.1, protein p55,  $\alpha$ - and  $\beta$ spectrin, actin, and actin-binding proteins such as dematin, tropomyosin, and tropomodulin.  $\alpha$ - and  $\beta$ spectrin combine to form a heterotetramer in vivo. The spectrin heterotetramer organizes into a cortical bidimensional network with a hexagonal mesh. The network is linked to trans-bilayer proteins through a protein complex comprising β-spectrin, ankyrin, anion exchanger, and protein 4.2 and through the "triangular" interaction between protein 4.1, glycophorin C, and protein p55. Structural and functional variants of erythrocyte membrane proteins have been have been found in a variety of tissues. Variants may be transcribed from multigene families, e.g., anion exchanger, ankyrin, or spectrin, or from single gene families, e.g., protein 4.1 or protein 4.2. mRNA transcripts undergo tissue-specific alternative splicing. Many congenital hemolytic anemias result from mutations in the above-mentioned genes encoding erythrocyte membrane proteins. For example, hereditary elliptocytosis stems from an array of mutations in the spectrin genes at or near the head-to-head self-association region of the spectrin tetramer, or from mutations in the protein 4.1 gene which reduce levels of protein 4.1. In another

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example, hereditary spherocytosis is associated with mutations in the ankyrin gene, the anion exchanger gene, the protein 4.2 gene, or the  $\alpha$ - and  $\beta$ -spectrin genes. (Delaunay J. (1995) Transfus. Clin. Biol. 2:207-216.)

Protein 4.1 is an 80 kDa erythrocyte membrane protein with four functional domains. These domains include: i) a 30 kDa basic N-terminal domain, homologous to the ERM (Ezrin/Radixin/Moesin) family of actin- and transmembrane protein-binding proteins (Tsukita, S. et al. (1997) Trends Biochem. Sci. 22:53-58); ii) a 16 kDa hydrophilic domain containing a protein kinase C phosphorylation site; iii) a 10 kDa highly charged domain containing a cAMP-dependent protein kinase phosphorylation site critical for the interaction with spectrin and actin; and iv) a 22/24 kDa acidic domain. Protein 4.1 is a member of a structurally and functionally related protein 4.1 family. The protein 4.1 family is part of an evolutionarily related protein superfamily that includes many tyrosine phosphatases. (Baklouti, F. et al. (1997) Genomics 39:289-302.)

In contrast to the strictly cortical localization of protein 4.1 in mature enucleate erythrocytes, protein 4.1 epitopes have been observed throughout the cytoplasmic compartment and the nucleoskeleton in nucleated cells. In particular, protein 4.1 is present in the nucleoskeleton during interphase, in the mitotic spindle during mitosis, in perichromatin during telophase, and in the midbody during cytokinesis. (Krauss, S.W. et al. (1997) J. Cell Biol. 137:275-289.)

Differential expression of the protein 4.1 gene resulting in a number of mRNA splice variants has been observed in various human and rodent tissues. Comparison of the gene structure and mRNA splice variants revealed the extreme genomic sequence conservation of protein 4.1 between different species. The 5' UTR of both the human and rodent mRNA species has not been successfully identified and sequenced, possibly due to GC-rich regions therein which give rise to technical complications during nucleotide sequencing reactions. (Baklouti, <u>supra;</u> Conboy, J.G. (1988) Proc. Natl. Acad. Sci. 85:9062-9065,)

Analysis of proteins included in the ERM family of proteins has indicated that the N-terminal domain interacts with intracellular domains of transmembrane proteins such as CD44 and the C-terminal domain binds actin. Both interactions involve interactions with Rho-GTP protein complex, polyphosphoinositides, and serine/threonine kinase and tyrosine kinase activities. Many of the phosphorylation sites on ERM proteins are conserved. Although expression of ERM proteins in vivo is restricted to tissues such as endothelium, repression of ERM protein gene expression is released under conditions of cell culture. (Tsukita, supra.)

The cortical actin cytoskeleton participates in various membrane-based processes which necessitate a large amount of functional plasticity in the molecular components involved. A family of proteins homologous to band 4.1 is involved in the reorganization of the actin cytoskeleton in response

to various stimuli and probably plays a role in transmembrane signaling. This family includes tyrosine phosphatases, substrates of tyrosine kinases and a candidate for a tumor-suppressor gene. (Arpin M, et al. (1994) Curr. Opin. Cell Biol. 6:136-141.)

Disruptions in cytoskeletal protein interaction have been identified in a number of disease conditions or disorders. Neurofibromatosis type 2 is an autosomal dominant disease of the nervous system. Schwann cells isolated from patients with neurofibromatosis type 2 have characteristic morphology and growth parameters which differ from control Schwann cells. A gene associated with neurofibromatosis type 2 has been identified and is termed NF2. The NF2 gene product, known as schwannomin or merlin, is a member of the protein 4.1 superfamily, and mutations in the NF2 gene have been shown to be associated with the disease. (Rosenbaum, C. et al. (1998) Neurobiol. Dis. 5:55-64.) In addition, a form of psoriasis may be due to altered expression or distribution in epidermal epithelium of analogs of erythrocyte protein 4.1. (Shimizu, T. (1996) Histol. Histopathol. 11:495-501.) Erythrocytes carrying mutations in spectrin and protein 4.1 showed differing sensitivities to invasion by Plasmodium falciparum. (Facer, C.A. (1995) Parasitol Res. 81:52-57.) Furthermore, antibodies raised against erythrocyte protein 4.1 stained the majority of neurofibrillary tangles in the prefrontal cortex and hippocampus of brain tissue from patients with Alzheimer's disease. A 68 kDa protein was identified as the most likely brain analog of erythrocyte protein 4.1. (Sihag, R.K. et al. (1994) Brain Res. 656:14-26.)

The discovery of new cytoskeleton-associated proteins and the polynucleotides encoding them satisfies a need in the art by providing new compositions which are useful in the diagnosis, prevention, and treatment of cell proliferative, autoimmune/inflammatory, vesicle trafficking, neurological, cell motility, reproductive, and muscle disorders, and in the assessment of the effects of exogenous compounds on the expression of nucleic acid and amino acid sequences of cytoskeleton-associated proteins.

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# SUMMARY OF THE INVENTION

The invention features purified polypeptides, cytoskeleton-associated proteins, referred to collectively as "CYSKP" and individually as "CYSKP-1," "CYSKP-2," "CYSKP-3," "CYSKP-4," "CYSKP-5," "CYSKP-6," "CYSKP-7," "CYSKP-8," "CYSKP-9," "CYSKP-10," "CYSKP-11," "CYSKP-12," "CYSKP-13," "CYSKP-14," "CYSKP-15," "CYSKP-16," "CYSKP-17," "CYSKP-18," "CYSKP-19," "CYSKP-20," "CYSKP-21," "CYSKP-22," "CYSKP-23," "CYSKP-24," "CYSKP-25," "CYSKP-26," "CYSKP-27," "CYSKP-28," "CYSKP-29," "CYSKP-30," "CYSKP-31," "CYSKP-32," "CYSKP-33," and "CYSKP-34." In one aspect, the invention provides an isolated polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence

selected from the group consisting of SEQ ID NO:1-34, b) a naturally occurring polypeptide comprising an amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34. In one alternative, the invention provides an isolated polypeptide comprising the amino acid sequence of SEQ ID NO:1-34.

The invention further provides an isolated polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, b) a naturally occurring polypeptide comprising an amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34. In one alternative, the polynucleotide encodes a polypeptide selected from the group consisting of SEQ ID NO:1-34. In another alternative, the polynucleotide is selected from the group consisting of SEQ ID NO:35-68.

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Additionally, the invention provides a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, b) a naturally occurring polypeptide comprising an amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34. In one alternative, the invention provides a cell transformed with the recombinant polynucleotide. In another alternative, the invention provides a transgenic organism comprising the recombinant polynucleotide.

The invention also provides a method for producing a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, b) a naturally occurring polypeptide comprising an amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34. The method comprises a)

culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding the polypeptide, and b) recovering the polypeptide so expressed.

Additionally, the invention provides an isolated antibody which specifically binds to a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, b) a naturally occurring polypeptide comprising an amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34.

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The invention further provides an isolated polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:35-68, b) a naturally occurring polynucleotide comprising a polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:35-68, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). In one alternative, the polynucleotide comprises at least 60 contiguous nucleotides.

Additionally, the invention provides a method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:35-68, b) a naturally occurring polynucleotide comprising a polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:35-68, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). The method comprises a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and b) detecting the presence or absence of said hybridization complex, and optionally, if present, the amount thereof. In one alternative, the probe comprises at least 60 contiguous nucleotides.

The invention further provides a method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID

NO:35-68, b) a naturally occurring polynucleotide comprising a polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:35-68, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). The method comprises a) amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.

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The invention further provides a composition comprising an effective amount of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, b) a naturally occurring polypeptide comprising an amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, and a pharmaceutically acceptable excipient. In one embodiment, the composition comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-34. The invention additionally provides a method of treating a disease or condition associated with decreased expression of functional CYSKP, comprising administering to a patient in need of such treatment the composition.

The invention also provides a method for screening a compound for effectiveness as an agonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, b) a naturally occurring polypeptide comprising an amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34. The method comprises a) exposing a sample comprising the polypeptide to a compound, and b) detecting agonist activity in the sample. In one alternative, the invention provides a composition comprising an agonist compound identified by the method and a pharmaceutically acceptable excipient. In another alternative, the invention provides a method of treating a disease or condition associated with decreased expression of functional CYSKP, comprising administering to a patient in need of such treatment the composition.

Additionally, the invention provides a method for screening a compound for effectiveness as an antagonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, b) a naturally occurring

polypeptide comprising an amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34. The method comprises a) exposing a sample comprising the polypeptide to a compound, and b) detecting antagonist activity in the sample. In one alternative, the invention provides a composition comprising an antagonist compound identified by the method and a pharmaceutically acceptable excipient. In another alternative, the invention provides a method of treating a disease or condition associated with overexpression of functional CYSKP, comprising administering to a patient in need of such treatment the composition.

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The invention further provides a method of screening for a compound that specifically binds to a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, b) a naturally occurring polypeptide comprising an amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34. The method comprises a) combining the polypeptide with at least one test compound under suitable conditions, and b) detecting binding of the polypeptide to the test compound, thereby identifying a compound that specifically binds to the polypeptide.

The invention further provides a method of screening for a compound that modulates the activity of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, b) a naturally occurring polypeptide comprising an amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34. The method comprises a) combining the polypeptide with at least one test compound under conditions permissive for the activity of the polypeptide, b) assessing the activity of the polypeptide in the presence of the test compound, and c) comparing the activity of the polypeptide in the absence of the test compound, wherein a change in the activity of the polypeptide in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide.

The invention further provides a method for screening a compound for effectiveness in

altering expression of a target polynucleotide, wherein said target polynucleotide comprises a sequence selected from the group consisting of SEQ ID NO:35-68, the method comprising a) exposing a sample comprising the target polynucleotide to a compound, and b) detecting altered expression of the target polynucleotide.

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The invention further provides a method for assessing toxicity of a test compound, said method comprising a) treating a biological sample containing nucleic acids with the test compound; b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:35-68, ii) a naturally occurring polynucleotide comprising a polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:35-68, iii) a polynucleotide having a sequence complementary to i), iv) a polynucleotide complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv). Hybridization occurs under conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:35-68, ii) a naturally occurring polynucleotide comprising a polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:35-68, iii) a polynucleotide complementary to the polynucleotide of i), iv) a polynucleotide complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv). Alternatively, the target polynucleotide comprises a fragment of a polynucleotide sequence selected from the group consisting of i)-v) above; c) quantifying the amount of hybridization complex; and d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.

# BRIEF DESCRIPTION OF THE TABLES

Table 1 summarizes the nomenclature for the full length polynucleotide and polypeptide sequences of the present invention.

Table 2 shows the GenBank identification number and annotation of the nearest GenBank homolog for polypeptides of the invention. The probability score for the match between each polypeptide and its GenBank homolog is also shown.

Table 3 shows structural features of polypeptide sequences of the invention, including predicted motifs and domains, along with the methods, algorithms, and searchable databases used for analysis of the polypeptides.

Table 4 lists the cDNA and genomic DNA fragments which were used to assemble polynucleotide sequences of the invention, along with selected fragments of the polynucleotide sequences.

Table 5 shows the representative cDNA library for polynucleotides of the invention.

Table 6 provides an appendix which describes the tissues and vectors used for construction of the cDNA libraries shown in Table 5.

Table 7 shows the tools, programs, and algorithms used to analyze the polynucleotides and polypeptides of the invention, along with applicable descriptions, references, and threshold parameters.

#### DESCRIPTION OF THE INVENTION

Before the present proteins, nucleotide sequences, and methods are described, it is understood that this invention is not limited to the particular machines, materials and methods described, as these may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims.

It must be noted that as used herein and in the appended claims, the singular forms "a," "an," and "the" include plural reference unless the context clearly dictates otherwise. Thus, for example, a reference to "a host cell" includes a plurality of such host cells, and a reference to "an antibody" is a reference to one or more antibodies and equivalents thereof known to those skilled in the art, and so forth.

Unless defined otherwise, all technical and scientific terms used herein have the same meanings as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any machines, materials, and methods similar or equivalent to those described herein can be used to practice or test the present invention, the preferred machines, materials and methods are now described. All publications mentioned herein are cited for the purpose of describing and disclosing the cell lines, protocols, reagents and vectors which are reported in the publications and which might be used in connection with the invention. Nothing herein is to be construed as an admission that the invention is not entitled to antedate such disclosure by virtue of prior invention.

# **DEFINITIONS**

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"CYSKP" refers to the amino acid sequences of substantially purified CYSKP obtained from any species, particularly a mammalian species, including bovine, ovine, porcine, murine, equine, and human, and from any source, whether natural, synthetic, semi-synthetic, or recombinant.

The term "agonist" refers to a molecule which intensifies or mimics the biological activity of CYSKP. Agonists may include proteins, nucleic acids, carbohydrates, small molecules, or any other

compound or composition which modulates the activity of CYSKP either by directly interacting with CYSKP or by acting on components of the biological pathway in which CYSKP participates.

An "allelic variant" is an alternative form of the gene encoding CYSKP. Allelic variants may result from at least one mutation in the nucleic acid sequence and may result in altered mRNAs or in polypeptides whose structure or function may or may not be altered. A gene may have none, one, or many allelic variants of its naturally occurring form. Common mutational changes which give rise to allelic variants are generally ascribed to natural deletions, additions, or substitutions of nucleotides. Each of these types of changes may occur alone, or in combination with the others, one or more times in a given sequence.

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"Altered" nucleic acid sequences encoding CYSKP include those sequences with deletions, insertions, or substitutions of different nucleotides, resulting in a polypeptide the same as CYSKP or a polypeptide with at least one functional characteristic of CYSKP. Included within this definition are polymorphisms which may or may not be readily detectable using a particular oligonucleotide probe of the polynucleotide encoding CYSKP, and improper or unexpected hybridization to allelic variants, with a locus other than the normal chromosomal locus for the polynucleotide sequence encoding CYSKP. The encoded protein may also be "altered," and may contain deletions, insertions, or substitutions of amino acid residues which produce a silent change and result in a functionally equivalent CYSKP. Deliberate amino acid substitutions may be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues, as long as the biological or immunological activity of CYSKP is retained. For example, negatively charged amino acids may include aspartic acid and glutamic acid, and positively charged amino acids may include lysine and arginine. Amino acids with uncharged polar side chains having similar hydrophilicity values may include; asparagine and glutamine; and serine and threonine. Amino acids with uncharged side chains having similar hydrophilicity values may include: leucine, isoleucine, and valine; glycine and alanine; and phenylalanine and tyrosine.

The terms "amino acid" and "amino acid sequence" refer to an oligopeptide, peptide, polypeptide, or protein sequence, or a fragment of any of these, and to naturally occurring or synthetic molecules. Where "amino acid sequence" is recited to refer to a sequence of a naturally occurring protein molecule, "amino acid sequence" and like terms are not meant to limit the amino acid sequence to the complete native amino acid sequence associated with the recited protein molecule.

"Amplification" relates to the production of additional copies of a nucleic acid sequence.

Amplification is generally carried out using polymerase chain reaction (PCR) technologies well known in the art.

The term "antagonist" refers to a molecule which inhibits or attenuates the biological activity of

CYSKP. Antagonists may include proteins such as antibodies, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of CYSKP either by directly interacting with CYSKP or by acting on components of the biological pathway in which CYSKP participates.

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The term "antibody" refers to intact immunoglobulin molecules as well as to fragments thereof, such as Fab, F(ab')<sub>2</sub>, and Fv fragments, which are capable of binding an epitopic determinant. Antibodies that bind CYSKP polypeptides can be prepared using intact polypeptides or using fragments containing small peptides of interest as the immunizing antigen. The polypeptide or oligopeptide used to immunize an animal (e.g., a mouse, a rat, or a rabbit) can be derived from the translation of RNA, or synthesized chemically, and can be conjugated to a carrier protein if desired. Commonly used carriers that are chemically coupled to peptides include bovine serum albumin, thyroglobulin, and keyhole limpet hemocyanin (KLH). The coupled peptide is then used to immunize the animal.

The term "antigenic determinant" refers to that region of a molecule (i.e., an epitope) that makes contact with a particular antibody. When a protein or a fragment of a protein is used to immunize a host animal, numerous regions of the protein may induce the production of antibodies which bind specifically to antigenic determinants (particular regions or three-dimensional structures on the protein). An antigenic determinant may compete with the intact antigen (i.e., the immunogen used to elicit the immune response) for binding to an antibody.

The term "antisense" refers to any composition capable of base-pairing with the "sense" (coding) strand of a specific nucleic acid sequence. Antisense compositions may include DNA; RNA; peptide nucleic acid (PNA); oligonucleotides having modified backbone linkages such as phosphorothioates, methylphosphonates, or benzylphosphonates; oligonucleotides having modified sugar groups such as 2'-methoxyethyl sugars or 2'-methoxyethoxy sugars; or oligonucleotides having modified bases such as 5-methyl cytosine, 2'-deoxyuracil, or 7-deaza-2'-deoxyguanosine. Antisense molecules may be produced by any method including chemical synthesis or transcription. Once introduced into a cell, the complementary antisense molecule base-pairs with a naturally occurring nucleic acid sequence produced by the cell to form duplexes which block either transcription or translation. The designation "negative" or "minus" can refer to the antisense strand, and the designation "positive" or "plus" can refer to the sense strand of a reference DNA molecule.

The term "biologically active" refers to a protein having structural, regulatory, or biochemical functions of a naturally occurring molecule. Likewise, "immunologically active" or "immunogenic" refers to the capability of the natural, recombinant, or synthetic CYSKP, or of any oligopeptide thereof, to induce a specific immune response in appropriate animals or cells and to bind with specific antibodies.

"Complementary" describes the relationship between two single-stranded nucleic acid sequences that anneal by base-pairing. For example, 5'-AGT-3' pairs with its complement, 3'-TCA-5'.

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A "composition comprising a given polynucleotide sequence" and a "composition comprising a given amino acid sequence" refer broadly to any composition containing the given polynucleotide or amino acid sequence. The composition may comprise a dry formulation or an aqueous solution. Compositions comprising polynucleotide sequences encoding CYSKP or fragments of CYSKP may be employed as hybridization probes. The probes may be stored in freeze-dried form and may be associated with a stabilizing agent such as a carbohydrate. In hybridizations, the probe may be deployed in an aqueous solution containing salts (e.g., NaCl), detergents (e.g., sodium dodecyl sulfate; SDS), and other components (e.g., Denhardt's solution, dry milk, salmon sperm DNA, etc.).

"Consensus sequence" refers to a nucleic acid sequence which has been subjected to repeated DNA sequence analysis to resolve uncalled bases, extended using the XL-PCR kit (Applied Biosystems, Foster City CA) in the 5' and/or the 3' direction, and resequenced, or which has been assembled from one or more overlapping cDNA, EST, or genomic DNA fragments using a computer program for fragment assembly, such as the GELVIEW fragment assembly system (GCG, Madison WI) or Phrap (University of Washington, Seattle WA). Some sequences have been both extended and assembled to produce the consensus sequence.

"Conservative amino acid substitutions" are those substitutions that are predicted to least interfere with the properties of the original protein, i.e., the structure and especially the function of the protein is conserved and not significantly changed by such substitutions. The table below shows amino acids which may be substituted for an original amino acid in a protein and which are regarded as conservative amino acid substitutions.

	Original Residue	Conservative Substitution
25	Ala	Gly, Ser
	Arg	His, Lys
	Asn	Asp, Gln, His
	Asp	Asn, Glu
	Cys	Ala, Ser
30	Gln	Asn, Glu, His
	Glu	Asp, Gln, His
	Gly	Ala
	His	Asn, Arg, Gln, Glu
	Ile	Leu, Val
35	Leu	Ile, Val
	Lys	Arg, Gln, Glu
	Met	Leu, Ile
	Phe	His, Met, Leu, Trp, Tyr
	Ser	Cys, Thr

Thr	Ser, Val
Trp	Phe, Tyr
Tyr	His, Phe, Trp
Val	Ile, Leu, Thr

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Conservative amino acid substitutions generally maintain (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a beta sheet or alpha helical conformation, (b) the charge or hydrophobicity of the molecule at the site of the substitution, and/or (c) the bulk of the side chain.

A "deletion" refers to a change in the amino acid or nucleotide sequence that results in the absence of one or more amino acid residues or nucleotides.

The term "derivative" refers to a chemically modified polynucleotide or polypeptide. Chemical modifications of a polynucleotide can include, for example, replacement of hydrogen by an alkyl, acyl, hydroxyl, or amino group. A derivative polynucleotide encodes a polypeptide which retains at least one biological or immunological function of the natural molecule. A derivative polypeptide is one modified by glycosylation, pegylation, or any similar process that retains at least one biological or immunological function of the polypeptide from which it was derived.

A "detectable label" refers to a reporter molecule or enzyme that is capable of generating a measurable signal and is covalently or noncovalently joined to a polynucleotide or polypeptide.

"Differential expression" refers to increased or upregulated; or decreased, downregulated, or absent gene or protein expression, determined by comparing at least two different samples. Such comparisons may be carried out between, for example, a treated and an untreated sample, or a diseased and a normal sample.

A "fragment" is a unique portion of CYSKP or the polynucleotide encoding CYSKP which is identical in sequence to but shorter in length than the parent sequence. A fragment may comprise up to the entire length of the defined sequence, minus one nucleotide/amino acid residue. For example, a fragment may comprise from 5 to 1000 contiguous nucleotides or amino acid residues. A fragment used as a probe, primer, antigen, therapeutic molecule, or for other purposes, may be at least 5, 10, 15, 16, 20, 25, 30, 40, 50, 60, 75, 100, 150, 250 or at least 500 contiguous nucleotides or amino acid residues in length. Fragments may be preferentially selected from certain regions of a molecule. For example, a polypeptide fragment may comprise a certain length of contiguous amino acids selected from the first 250 or 500 amino acids (or first 25% or 50%) of a polypeptide as shown in a certain defined sequence. Clearly these lengths are exemplary, and any length that is supported by the specification, including the Sequence Listing, tables, and figures, may be encompassed by the present embodiments.

A fragment of SEQ ID NO:35-68 comprises a region of unique polynucleotide sequence that

specifically identifies SEQ ID NO:35-68, for example, as distinct from any other sequence in the genome from which the fragment was obtained. A fragment of SEQ ID NO:35-68 is useful, for example, in hybridization and amplification technologies and in analogous methods that distinguish SEQ ID NO:35-68 from related polynucleotide sequences. The precise length of a fragment of SEQ ID NO:35-68 and the region of SEQ ID NO:35-68 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

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A fragment of SEQ ID NO:1-34 is encoded by a fragment of SEQ ID NO:35-68. A fragment of SEQ ID NO:1-34 comprises a region of unique amino acid sequence that specifically identifies SEQ ID NO:1-34. For example, a fragment of SEQ ID NO:1-34 is useful as an immunogenic peptide for the development of antibodies that specifically recognize SEQ ID NO:1-34. The precise length of a fragment of SEQ ID NO:1-34 and the region of SEQ ID NO:1-34 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A "full length" polynucleotide sequence is one containing at least a translation initiation codon (e.g., methionine) followed by an open reading frame and a translation termination codon. A "full length" polynucleotide sequence encodes a "full length" polypeptide sequence.

"Homology" refers to sequence similarity or, interchangeably, sequence identity, between two or more polynucleotide sequences or two or more polypeptide sequences.

The terms "percent identity" and "% identity," as applied to polynucleotide sequences, refer to the percentage of residue matches between at least two polynucleotide sequences aligned using a standardized algorithm. Such an algorithm may insert, in a standardized and reproducible way, gaps in the sequences being compared in order to optimize alignment between two sequences, and therefore achieve a more meaningful comparison of the two sequences.

Percent identity between polynucleotide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program. This program is part of the LASERGENE software package, a suite of molecular biological analysis programs (DNASTAR, Madison WI). CLUSTAL V is described in Higgins, D.G. and P.M. Sharp (1989) CABIOS 5:151-153 and in Higgins, D.G. et al. (1992) CABIOS 8:189-191. For pairwise alignments of polynucleotide sequences, the default parameters are set as follows: Ktuple=2, gap penalty=5, window=4, and "diagonals saved"=4. The "weighted" residue weight table is selected as the default. Percent identity is reported by CLUSTAL V as the "percent similarity" between aligned polynucleotide sequences.

Alternatively, a suite of commonly used and freely available sequence comparison algorithms is provided by the National Center for Biotechnology Information (NCBI) Basic Local Alignment Search

Tool (BLAST) (Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410), which is available from several sources, including the NCBI, Bethesda, MD, and on the Internet at http://www.ncbi.nlm.nih.gov/BLAST/. The BLAST software suite includes various sequence analysis programs including "blastn," that is used to align a known polynucleotide sequence with other polynucleotide sequences from a variety of databases. Also available is a tool called "BLAST 2 Sequences" that is used for direct pairwise comparison of two nucleotide sequences. "BLAST 2 Sequences" can be accessed and used interactively at http://www.ncbi.nlm.nih.gov/gorf/bl2.html. The "BLAST 2 Sequences" tool can be used for both blastn and blastp (discussed below). BLAST programs are commonly used with gap and other parameters set to default settings. For example, to compare two nucleotide sequences, one may use blastn with the "BLAST 2 Sequences" tool Version 2.0.12 (April-21-2000) set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62
Reward for match: 1

Penalty for mismatch: -2

Open Gap: 5 and Extension Gap: 2 penalties

Gap x drop-off: 50

Expect: 10

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Word Size: 11

Filter: on

Percent identity may be measured over the length of an entire defined sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined sequence, for instance, a fragment of at least 20, at least 30, at least 40, at least 50, at least 70, at least 100, or at least 200 contiguous nucleotides. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures, or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

Nucleic acid sequences that do not show a high degree of identity may nevertheless encode similar amino acid sequences due to the degeneracy of the genetic code. It is understood that changes in a nucleic acid sequence can be made using this degeneracy to produce multiple nucleic acid sequences that all encode substantially the same protein.

The phrases "percent identity" and "% identity," as applied to polypeptide sequences, refer to the percentage of residue matches between at least two polypeptide sequences aligned using a standardized algorithm. Methods of polypeptide sequence alignment are well-known. Some alignment methods take into account conservative amino acid substitutions. Such conservative substitutions,

explained in more detail above, generally preserve the charge and hydrophobicity at the site of substitution, thus preserving the structure (and therefore function) of the polypeptide.

Percent identity between polypeptide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program (described and referenced above). For pairwise alignments of polypeptide sequences using CLUSTAL V, the default parameters are set as follows: Ktuple=1, gap penalty=3, window=5, and "diagonals saved"=5. The PAM250 matrix is selected as the default residue weight table. As with polynucleotide alignments, the percent identity is reported by CLUSTAL V as the "percent similarity" between aligned polypeptide sequence pairs.

Alternatively the NCBI BLAST software suite may be used. For example, for a pairwise comparison of two polypeptide sequences, one may use the "BLAST 2 Sequences" tool Version 2.0.12 (April-21-2000) with blastp set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

Open Gap: 11 and Extension Gap: 1 penalties

Gap x drop-off: 50

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Expect: 10
Word Size: 3

Filter: on

Percent identity may be measured over the length of an entire defined polypeptide sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined polypeptide sequence, for instance, a fragment of at least 15, at least 20, at least 30, at least 40, at least 50, at least 70 or at least 150 contiguous residues. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

"Human artificial chromosomes" (HACs) are linear microchromosomes which may contain DNA sequences of about 6 kb to 10 Mb in size and which contain all of the elements required for chromosome replication, segregation and maintenance.

The term "humanized antibody" refers to an antibody molecule in which the amino acid sequence in the non-antigen binding regions has been altered so that the antibody more closely resembles a human antibody, and still retains its original binding ability.

"Hybridization" refers to the process by which a polynucleotide strand anneals with a complementary strand through base pairing under defined hybridization conditions. Specific hybridization is an indication that two nucleic acid sequences share a high degree of complementarity.

Specific hybridization complexes form under permissive annealing conditions and remain hybridized after the "washing" step(s). The washing step(s) is particularly important in determining the stringency of the hybridization process, with more stringent conditions allowing less non-specific binding, i.e., binding between pairs of nucleic acid strands that are not perfectly matched. Permissive conditions for annealing of nucleic acid sequences are routinely determinable by one of ordinary skill in the art and may be consistent among hybridization experiments, whereas wash conditions may be varied among experiments to achieve the desired stringency, and therefore hybridization specificity. Permissive annealing conditions occur, for example, at 68°C in the presence of about 6 x SSC, about 1% (w/v) SDS, and about 100 μg/ml sheared, denatured salmon sperm DNA.

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Generally, stringency of hybridization is expressed, in part, with reference to the temperature under which the wash step is carried out. Such wash temperatures are typically selected to be about 5°C to 20°C lower than the thermal melting point  $(T_m)$  for the specific sequence at a defined ionic strength and pH. The  $T_m$  is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. An equation for calculating  $T_m$  and conditions for nucleic acid hybridization are well known and can be found in Sambrook, J. et al. (1989) Molecular Cloning: A Laboratory Manual,  $2^{nd}$  ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; specifically see volume 2, chapter 9.

High stringency conditions for hybridization between polynucleotides of the present invention include wash conditions of  $68^{\circ}$ C in the presence of about 0.2 x SSC and about 0.1% SDS, for 1 hour. Alternatively, temperatures of about  $65^{\circ}$ C,  $60^{\circ}$ C,  $55^{\circ}$ C, or  $42^{\circ}$ C may be used. SSC concentration may be varied from about 0.1 to 2 x SSC, with SDS being present at about 0.1%. Typically, blocking reagents are used to block non-specific hybridization. Such blocking reagents include, for instance, sheared and denatured salmon sperm DNA at about  $100\text{-}200 \,\mu\text{g/ml}$ . Organic solvent, such as formamide at a concentration of about  $35\text{-}50\% \,\text{v/v}$ , may also be used under particular circumstances, such as for RNA:DNA hybridizations. Useful variations on these wash conditions will be readily apparent to those of ordinary skill in the art. Hybridization, particularly under high stringency conditions, may be suggestive of evolutionary similarity between the nucleotides. Such similarity is strongly indicative of a similar role for the nucleotides and their encoded polypeptides.

The term "hybridization complex" refers to a complex formed between two nucleic acid sequences by virtue of the formation of hydrogen bonds between complementary bases. A hybridization complex may be formed in solution (e.g.,  $C_0 t$  or  $R_0 t$  analysis) or formed between one nucleic acid sequence present in solution and another nucleic acid sequence immobilized on a solid support (e.g., paper, membranes, filters, chips, pins or glass slides, or any other appropriate substrate to which cells or their nucleic acids have been fixed).

The words "insertion" and "addition" refer to changes in an amino acid or nucleotide sequence resulting in the addition of one or more amino acid residues or nucleotides, respectively.

"Immune response" can refer to conditions associated with inflammation, trauma, immune disorders, or infectious or genetic disease, etc. These conditions can be characterized by expression of various factors, e.g., cytokines, chemokines, and other signaling molecules, which may affect cellular and systemic defense systems.

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An "immunogenic fragment" is a polypeptide or oligopeptide fragment of CYSKP which is capable of eliciting an immune response when introduced into a living organism, for example, a mammal. The term "immunogenic fragment" also includes any polypeptide or oligopeptide fragment of CYSKP which is useful in any of the antibody production methods disclosed herein or known in the art.

The term "microarray" refers to an arrangement of a plurality of polynucleotides, polypeptides, or other chemical compounds on a substrate.

The terms "element" and "array element" refer to a polynucleotide, polypeptide, or other chemical compound having a unique and defined position on a microarray.

The term "modulate" refers to a change in the activity of CYSKP. For example, modulation may cause an increase or a decrease in protein activity, binding characteristics, or any other biological, functional, or immunological properties of CYSKP.

The phrases "nucleic acid" and "nucleic acid sequence" refer to a nucleotide, oligonucleotide, polynucleotide, or any fragment thereof. These phrases also refer to DNA or RNA of genomic or synthetic origin which may be single-stranded or double-stranded and may represent the sense or the antisense strand, to peptide nucleic acid (PNA), or to any DNA-like or RNA-like material.

"Operably linked" refers to the situation in which a first nucleic acid sequence is placed in a functional relationship with a second nucleic acid sequence. For instance, a promoter is operably linked to a coding sequence if the promoter affects the transcription or expression of the coding sequence. Operably linked DNA sequences may be in close proximity or contiguous and, where necessary to join two protein coding regions, in the same reading frame.

"Peptide nucleic acid" (PNA) refers to an antisense molecule or anti-gene agent which comprises an oligonucleotide of at least about 5 nucleotides in length linked to a peptide backbone of amino acid residues ending in lysine. The terminal lysine confers solubility to the composition. PNAs preferentially bind complementary single stranded DNA or RNA and stop transcript elongation, and may be pegylated to extend their lifespan in the cell.

"Post-translational modification" of an CYSKP may involve lipidation, glycosylation, phosphorylation, acetylation, racemization, proteolytic cleavage, and other modifications known in the art. These processes may occur synthetically or biochemically. Biochemical modifications will vary by

cell type depending on the enzymatic milieu of CYSKP.

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"Probe" refers to nucleic acid sequences encoding CYSKP, their complements, or fragments thereof, which are used to detect identical, allelic or related nucleic acid sequences. Probes are isolated oligonucleotides or polynucleotides attached to a detectable label or reporter molecule. Typical labels include radioactive isotopes, ligands, chemiluminescent agents, and enzymes. "Primers" are short nucleic acids, usually DNA oligonucleotides, which may be annealed to a target polynucleotide by complementary base-pairing. The primer may then be extended along the target DNA strand by a DNA polymerase enzyme. Primer pairs can be used for amplification (and identification) of a nucleic acid sequence, e.g., by the polymerase chain reaction (PCR).

Probes and primers as used in the present invention typically comprise at least 15 contiguous nucleotides of a known sequence. In order to enhance specificity, longer probes and primers may also be employed, such as probes and primers that comprise at least 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, or at least 150 consecutive nucleotides of the disclosed nucleic acid sequences. Probes and primers may be considerably longer than these examples, and it is understood that any length supported by the specification, including the tables, figures, and Sequence Listing, may be used.

Methods for preparing and using probes and primers are described in the references, for example Sambrook, J. et al. (1989) Molecular Cloning: A Laboratory Manual, 2<sup>nd</sup> ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; Ausubel, F.M. et al. (1987) Current Protocols in Molecular Biology, Greene Publ. Assoc. & Wiley-Intersciences, New York NY; Innis, M. et al. (1990) PCR Protocols, A Guide to Methods and Applications, Academic Press, San Diego CA. PCR primer pairs can be derived from a known sequence, for example, by using computer programs intended for that purpose such as Primer (Version 0.5, 1991, Whitehead Institute for Biomedical Research, Cambridge MA).

Oligonucleotides for use as primers are selected using software known in the art for such purpose. For example, OLIGO 4.06 software is useful for the selection of PCR primer pairs of up to 100 nucleotides each, and for the analysis of oligonucleotides and larger polynucleotides of up to 5,000 nucleotides from an input polynucleotide sequence of up to 32 kilobases. Similar primer selection programs have incorporated additional features for expanded capabilities. For example, the PrimOU primer selection program (available to the public from the Genome Center at University of Texas South West Medical Center, Dallas TX) is capable of choosing specific primers from megabase sequences and is thus useful for designing primers on a genome-wide scope. The Primer3 primer selection program (available to the public from the Whitehead Institute/MIT Center for Genome Research, Cambridge MA) allows the user to input a "mispriming library," in which sequences to avoid as primer binding sites are user-specified. Primer3 is useful, in particular, for the selection of oligonucleotides for

microarrays. (The source code for the latter two primer selection programs may also be obtained from their respective sources and modified to meet the user's specific needs.) The PrimeGen program (available to the public from the UK Human Genome Mapping Project Resource Centre, Cambridge UK) designs primers based on multiple sequence alignments, thereby allowing selection of primers that hybridize to either the most conserved or least conserved regions of aligned nucleic acid sequences. Hence, this program is useful for identification of both unique and conserved oligonucleotides and polynucleotide fragments. The oligonucleotides and polynucleotide fragments identified by any of the above selection methods are useful in hybridization technologies, for example, as PCR or sequencing primers, microarray elements, or specific probes to identify fully or partially complementary polynucleotides in a sample of nucleic acids. Methods of oligonucleotide selection are not limited to those described above.

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A "recombinant nucleic acid" is a sequence that is not naturally occurring or has a sequence that is made by an artificial combination of two or more otherwise separated segments of sequence. This artificial combination is often accomplished by chemical synthesis or, more commonly, by the artificial manipulation of isolated segments of nucleic acids, e.g., by genetic engineering techniques such as those described in Sambrook, <u>supra</u>. The term recombinant includes nucleic acids that have been altered solely by addition, substitution, or deletion of a portion of the nucleic acid. Frequently, a recombinant nucleic acid may include a nucleic acid sequence operably linked to a promoter sequence. Such a recombinant nucleic acid may be part of a vector that is used, for example, to transform a cell.

Alternatively, such recombinant nucleic acids may be part of a viral vector, e.g., based on a vaccinia virus, that could be use to vaccinate a mammal wherein the recombinant nucleic acid is expressed, inducing a protective immunological response in the mammal.

A "regulatory element" refers to a nucleic acid sequence usually derived from untranslated regions of a gene and includes enhancers, promoters, introns, and 5' and 3' untranslated regions (UTRs). Regulatory elements interact with host or viral proteins which control transcription, translation, or RNA stability.

"Reporter molecules" are chemical or biochemical moieties used for labeling a nucleic acid, amino acid, or antibody. Reporter molecules include radionuclides; enzymes; fluorescent, chemiluminescent, or chromogenic agents; substrates; cofactors; inhibitors; magnetic particles; and other moieties known in the art.

An "RNA equivalent," in reference to a DNA sequence, is composed of the same linear sequence of nucleotides as the reference DNA sequence with the exception that all occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

The term "sample" is used in its broadest sense. A sample suspected of containing CYSKP, nucleic acids encoding CYSKP, or fragments thereof may comprise a bodily fluid; an extract from a cell, chromosome, organelle, or membrane isolated from a cell; a cell; genomic DNA, RNA, or cDNA, in solution or bound to a substrate; a tissue; a tissue print; etc.

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The terms "specific binding" and "specifically binding" refer to that interaction between a protein or peptide and an agonist, an antibody, an antagonist, a small molecule, or any natural or synthetic binding composition. The interaction is dependent upon the presence of a particular structure of the protein, e.g., the antigenic determinant or epitope, recognized by the binding molecule. For example, if an antibody is specific for epitope "A," the presence of a polypeptide comprising the epitope A, or the presence of free unlabeled A, in a reaction containing free labeled A and the antibody will reduce the amount of labeled A that binds to the antibody.

The term "substantially purified" refers to nucleic acid or amino acid sequences that are removed from their natural environment and are isolated or separated, and are at least 60% free, preferably at least 75% free, and most preferably at least 90% free from other components with which they are naturally associated.

A "substitution" refers to the replacement of one or more amino acid residues or nucleotides by different amino acid residues or nucleotides, respectively.

"Substrate" refers to any suitable rigid or semi-rigid support including membranes, filters, chips, slides, wafers, fibers, magnetic or nonmagnetic beads, gels, tubing, plates, polymers, microparticles and capillaries. The substrate can have a variety of surface forms, such as wells, trenches, pins, channels and pores, to which polynucleotides or polypeptides are bound.

A "transcript image" refers to the collective pattern of gene expression by a particular cell type or tissue under given conditions at a given time.

"Transformation" describes a process by which exogenous DNA is introduced into a recipient cell. Transformation may occur under natural or artificial conditions according to various methods well known in the art, and may rely on any known method for the insertion of foreign nucleic acid sequences into a prokaryotic or eukaryotic host cell. The method for transformation is selected based on the type of host cell being transformed and may include, but is not limited to, bacteriophage or viral infection, electroporation, heat shock, lipofection, and particle bombardment. The term "transformed cells" includes stably transformed cells in which the inserted DNA is capable of replication either as an autonomously replicating plasmid or as part of the host chromosome, as well as transiently transformed cells which express the inserted DNA or RNA for limited periods of time.

A "transgenic organism," as used herein, is any organism, including but not limited to animals and plants, in which one or more of the cells of the organism contains heterologous nucleic

acid introduced by way of human intervention, such as by transgenic techniques well known in the art. The nucleic acid is introduced into the cell, directly or indirectly by introduction into a precursor of the cell, by way of deliberate genetic manipulation, such as by microinjection or by infection with a recombinant virus. The term genetic manipulation does not include classical cross-breeding, or in vitro fertilization, but rather is directed to the introduction of a recombinant DNA molecule. The transgenic organisms contemplated in accordance with the present invention include bacteria, cyanobacteria, fungi, plants and animals. The isolated DNA of the present invention can be introduced into the host by methods known in the art, for example infection, transfection, transformation or transconjugation. Techniques for transferring the DNA of the present invention into such organisms are widely known and provided in references such as Sambrook et al. (1989), supra.

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A "variant" of a particular nucleic acid sequence is defined as a nucleic acid sequence having at least 40% sequence identity to the particular nucleic acid sequence over a certain length of one of the nucleic acid sequences using blastn with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of nucleic acids may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% or greater sequence identity over a certain defined length. A variant may be described as, for example, an "allelic" (as defined above), "splice," "species," or "polymorphic" variant. A splice variant may have significant identity to a reference molecule, but will generally have a greater or lesser number of polynucleotides due to alternative splicing of exons during mRNA processing. The corresponding polypeptide may possess additional functional domains or lack domains that are present in the reference molecule. Species variants are polynucleotide sequences that vary from one species to another. The resulting polypeptides will generally have significant amino acid identity relative to each other. A polymorphic variant is a variation in the polynucleotide sequence of a particular gene between individuals of a given species. Polymorphic variants also may encompass "single nucleotide polymorphisms" (SNPs) in which the polynucleotide sequence varies by one nucleotide base. The presence of SNPs may be indicative of, for example, a certain population, a disease state, or a propensity for a disease state.

A "variant" of a particular polypeptide sequence is defined as a polypeptide sequence having at least 40% sequence identity to the particular polypeptide sequence over a certain length of one of the polypeptide sequences using blastp with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of polypeptides may show, for example, at least 50%, at least 50%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% or greater sequence identity over a

certain defined length of one of the polypeptides.

#### THE INVENTION

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The invention is based on the discovery of new human cytoskeleton-associated proteins (CYSKP), the polynucleotides encoding CYSKP, and the use of these compositions for the diagnosis, treatment, or prevention of cell proliferative, autoimmune/inflammatory, vesicle trafficking, neurological, cell motility, reproductive, and muscle disorders.

Table 1 summarizes the nomenclature for the full length polynucleotide and polypeptide sequences of the invention. Each polynucleotide and its corresponding polypeptide are correlated to a single Incyte project identification number (Incyte Project ID). Each polypeptide sequence is denoted by both a polypeptide sequence identification number (Polypeptide SEQ ID NO:) and an Incyte polypeptide sequence number (Incyte Polypeptide ID) as shown. Each polynucleotide sequence is denoted by both a polynucleotide sequence identification number (Polynucleotide SEQ ID NO:) and an Incyte polynucleotide consensus sequence number (Incyte Polynucleotide ID) as shown.

Table 2 shows sequences with homology to the polypeptides of the invention as identified by BLAST analysis against the GenBank protein (genpept) database. Columns 1 and 2 show the polypeptide sequence identification number (Polypeptide SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for polypeptides of the invention. Column 3 shows the GenBank identification number (Genbank ID NO:) of the nearest GenBank homolog. Column 4 shows the probability score for the match between each polypeptide and its GenBank homolog. Column 5 shows the annotation of the GenBank homolog along with relevant citations where applicable, all of which are expressly incorporated by reference herein.

Table 3 shows various structural features of the polypeptides of the invention. Columns 1 and 2 show the polypeptide sequence identification number (SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for each polypeptide of the invention. Column 3 shows the number of amino acid residues in each polypeptide. Column 4 shows potential phosphorylation sites, and column 5 shows potential glycosylation sites, as determined by the MOTIFS program of the GCG sequence analysis software package (Genetics Computer Group, Madison WI). Column 6 shows amino acid residues comprising signature sequences, domains, and motifs. Column 7 shows analytical methods for protein structure/function analysis and in some cases, searchable databases to which the analytical methods were applied.

Together, Tables 2 and 3 summarize the properties of polypeptides of the invention, and these properties establish that the claimed polypeptides are cytoskeleton-associated proteins. For example, SEQ ID NO:31 is 34% identical to a Caenorhabditis elegans protein similar to mouse ankyrin

(GenBank ID g3879121) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 1.1e-146, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:31 also contains Ank repeats as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. As a second example, SEQ ID NO:34 is 96% identical over 97 amino acids to human Intermediate Filament Associated Protein (GenBank ID 1333846) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 8.2e-45, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. Data from BLAST analyses using the PRODOM database provide further corroborative evidence that SEQ ID NO:34 is a cytoskeleton protein. (See Table 3.) SEQ ID NO:1-30 and SEQ ID NO:32-33 were analyzed and annotated in a similar manner. The algorithms and parameters for the analysis of SEQ ID NO:1-34 are described in Table 7.

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As shown in Table 4, the full length polynucleotide sequences of the present invention were assembled using cDNA sequences or coding (exon) sequences derived from genomic DNA, or any combination of these two types of sequences. Columns 1 and 2 list the polynucleotide sequence identification number (Polynucleotide SEQ ID NO:) and the corresponding Incyte polynucleotide consensus sequence number (Incyte Polynucleotide ID) for each polynucleotide of the invention. Column 3 shows the length of each polynucleotide sequence in basepairs. Column 4 lists fragments of the polynucleotide sequences which are useful, for example, in hybridization or amplification technologies that identify SEQ ID NO:35-68 or that distinguish between SEQ ID NO:35-68 and related polynucleotide sequences. Column 5 shows identification numbers corresponding to cDNA sequences, coding sequences (exons) predicted from genomic DNA, and/or sequence assemblages comprised of both cDNA and genomic DNA. These sequences were used to assemble the full length polynucleotide sequences of the invention. Columns 6 and 7 of Table 4 show the nucleotide start (5') and stop (3') positions of the cDNA and genomic sequences in column 5 relative to their respective full length sequences.

The identification numbers in Column 5 of Table 4 may refer specifically, for example, to Incyte cDNAs along with their corresponding cDNA libraries. For example, 3824958H1 is the identification number of an Incyte cDNA sequence, and BRAXNOT01 is the cDNA library from which it is derived. Incyte cDNAs for which cDNA libraries are not indicated were derived from pooled cDNA libraries (e.g., 71263527V1). Alternatively, the identification numbers in column 5 may refer to GenBank cDNAs or ESTs (e.g., g2276318) which contributed to the assembly of the full length polynucleotide sequences. Alternatively, the identification numbers in column 5 may refer to coding regions predicted by Genscan analysis of genomic DNA. The Genscan-predicted coding sequences may

have been edited prior to assembly. (See Example IV.) Alternatively, the identification numbers in column 5 may refer to assemblages of both cDNA and Genscan-predicted exons brought together by an "exon stitching" algorithm. (See Example V.) Alternatively, the identification numbers in column 5 may refer to assemblages of both cDNA and Genscan-predicted exons brought together by an "exon-stretching" algorithm. (See Example V.) In some cases, Incyte cDNA coverage redundant with the sequence coverage shown in column 5 was obtained to confirm the final consensus polynucleotide sequence, but the relevant Incyte cDNA identification numbers are not shown.

Table 5 shows the representative cDNA libraries for those full length polynucleotide sequences which were assembled using Incyte cDNA sequences. The representative cDNA library is the Incyte cDNA library which is most frequently represented by the Incyte cDNA sequences which were used to assemble and confirm the above polynucleotide sequences. The tissues and vectors which were used to construct the cDNA libraries shown in Table 5 are described in Table 6.

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The invention also encompasses CYSKP variants. A preferred CYSKP variant is one which has at least about 80%, or alternatively at least about 90%, or even at least about 95% amino acid sequence identity to the CYSKP amino acid sequence, and which contains at least one functional or structural characteristic of CYSKP.

The invention also encompasses polynucleotides which encode CYSKP. In a particular embodiment, the invention encompasses a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:35-68, which encodes CYSKP. The polynucleotide sequences of SEQ ID NO:35-68, as presented in the Sequence Listing, embrace the equivalent RNA sequences, wherein occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

The invention also encompasses a variant of a polynucleotide sequence encoding CYSKP. In particular, such a variant polynucleotide sequence will have at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to the polynucleotide sequence encoding CYSKP. A particular aspect of the invention encompasses a variant of a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:35-68 which has at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to a nucleic acid sequence selected from the group consisting of SEQ ID NO:35-68. Any one of the polynucleotide variants described above can encode an amino acid sequence which contains at least one functional or structural characteristic of CYSKP.

It will be appreciated by those skilled in the art that as a result of the degeneracy of the genetic code, a multitude of polynucleotide sequences encoding CYSKP, some bearing minimal similarity to the polynucleotide sequences of any known and naturally occurring gene, may be produced. Thus, the

invention contemplates each and every possible variation of polynucleotide sequence that could be made by selecting combinations based on possible codon choices. These combinations are made in accordance with the standard triplet genetic code as applied to the polynucleotide sequence of naturally occurring CYSKP, and all such variations are to be considered as being specifically disclosed.

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Although nucleotide sequences which encode CYSKP and its variants are generally capable of hybridizing to the nucleotide sequence of the naturally occurring CYSKP under appropriately selected conditions of stringency, it may be advantageous to produce nucleotide sequences encoding CYSKP or its derivatives possessing a substantially different codon usage, e.g., inclusion of non-naturally occurring codons. Codons may be selected to increase the rate at which expression of the peptide occurs in a particular prokaryotic or eukaryotic host in accordance with the frequency with which particular codons are utilized by the host. Other reasons for substantially altering the nucleotide sequence encoding CYSKP and its derivatives without altering the encoded amino acid sequences include the production of RNA transcripts having more desirable properties, such as a greater half-life, than transcripts produced from the naturally occurring sequence.

The invention also encompasses production of DNA sequences which encode CYSKP and CYSKP derivatives, or fragments thereof, entirely by synthetic chemistry. After production, the synthetic sequence may be inserted into any of the many available expression vectors and cell systems using reagents well known in the art. Moreover, synthetic chemistry may be used to introduce mutations into a sequence encoding CYSKP or any fragment thereof.

Also encompassed by the invention are polynucleotide sequences that are capable of hybridizing to the claimed polynucleotide sequences, and, in particular, to those shown in SEQ ID NO:35-68 and fragments thereof under various conditions of stringency. (See, e.g., Wahl, G.M. and S.L. Berger (1987) Methods Enzymol. 152:399-407; Kimmel, A.R. (1987) Methods Enzymol. 152:507-511.) Hybridization conditions, including annealing and wash conditions, are described in "Definitions."

Methods for DNA sequencing are well known in the art and may be used to practice any of the embodiments of the invention. The methods may employ such enzymes as the Klenow fragment of DNA polymerase I, SEQUENASE (US Biochemical, Cleveland OH), Taq polymerase (Applied Biosystems), thermostable T7 polymerase (Amersham Pharmacia Biotech, Piscataway NJ), or combinations of polymerases and proofreading exonucleases such as those found in the ELONGASE amplification system (Life Technologies, Gaithersburg MD). Preferably, sequence preparation is automated with machines such as the MICROLAB 2200 liquid transfer system (Hamilton, Reno NV), PTC200 thermal cycler (MJ Research, Watertown MA) and ABI CATALYST 800 thermal cycler (Applied Biosystems). Sequencing is then carried out using either the ABI 373 or 377 DNA sequencing

system (Applied Biosystems), the MEGABACE 1000 DNA sequencing system (Molecular Dynamics, Sunnyvale CA), or other systems known in the art. The resulting sequences are analyzed using a variety of algorithms which are well known in the art. (See, e.g., Ausubel, F.M. (1997) Short Protocols in Molecular Biology, John Wiley & Sons, New York NY, unit 7.7; Meyers, R.A. (1995) Molecular Biology and Biotechnology, Wiley VCH, New York NY, pp. 856-853.)

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The nucleic acid sequences encoding CYSKP may be extended utilizing a partial nucleotide sequence and employing various PCR-based methods known in the art to detect upstream sequences, such as promoters and regulatory elements. For example, one method which may be employed, restriction-site PCR, uses universal and nested primers to amplify unknown sequence from genomic DNA within a cloning vector. (See, e.g., Sarkar, G. (1993) PCR Methods Applic. 2:318-322.) Another method, inverse PCR, uses primers that extend in divergent directions to amplify unknown sequence from a circularized template. The template is derived from restriction fragments comprising a known genomic locus and surrounding sequences. (See, e.g., Triglia, T. et al. (1988) Nucleic Acids Res. 16:8186.) A third method, capture PCR, involves PCR amplification of DNA fragments adjacent to known sequences in human and yeast artificial chromosome DNA. (See, e.g., Lagerstrom, M. et al. (1991) PCR Methods Applic. 1:111-119.) In this method, multiple restriction enzyme digestions and ligations may be used to insert an engineered double-stranded sequence into a region of unknown sequence before performing PCR. Other methods which may be used to retrieve unknown sequences are known in the art. (See, e.g., Parker, J.D. et al. (1991) Nucleic Acids Res. 19:3055-3060). Additionally, one may use PCR, nested primers, and PROMOTERFINDER libraries (Clontech, Palo Alto CA) to walk genomic DNA. This procedure avoids the need to screen libraries and is useful in finding intron/exon junctions. For all PCR-based methods, primers may be designed using commercially available software, such as OLIGO 4.06 primer analysis software (National Biosciences, Plymouth MN) or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the template at temperatures of about 68°C to

When screening for full length cDNAs, it is preferable to use libraries that have been size-selected to include larger cDNAs. In addition, random-primed libraries, which often include sequences containing the 5' regions of genes, are preferable for situations in which an oligo d(T) library does not yield a full-length cDNA. Genomic libraries may be useful for extension of sequence into 5' non-transcribed regulatory regions.

Capillary electrophoresis systems which are commercially available may be used to analyze the size or confirm the nucleotide sequence of sequencing or PCR products. In particular, capillary sequencing may employ flowable polymers for electrophoretic separation, four different nucleotide-

specific, laser-stimulated fluorescent dyes, and a charge coupled device camera for detection of the emitted wavelengths. Output/light intensity may be converted to electrical signal using appropriate software (e.g., GENOTYPER and SEQUENCE NAVIGATOR, Applied Biosystems), and the entire process from loading of samples to computer analysis and electronic data display may be computer controlled. Capillary electrophoresis is especially preferable for sequencing small DNA fragments which may be present in limited amounts in a particular sample.

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In another embodiment of the invention, polynucleotide sequences or fragments thereof which encode CYSKP may be cloned in recombinant DNA molecules that direct expression of CYSKP, or fragments or functional equivalents thereof, in appropriate host cells. Due to the inherent degeneracy of the genetic code, other DNA sequences which encode substantially the same or a functionally equivalent amino acid sequence may be produced and used to express CYSKP.

The nucleotide sequences of the present invention can be engineered using methods generally known in the art in order to alter CYSKP-encoding sequences for a variety of purposes including, but not limited to, modification of the cloning, processing, and/or expression of the gene product. DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. For example, oligonucleotide-mediated site-directed mutagenesis may be used to introduce mutations that create new restriction sites, alter glycosylation patterns, change codon preference, produce splice variants, and so forth.

The nucleotides of the present invention may be subjected to DNA shuffling techniques such as MOLECULARBREEDING (Maxygen Inc., Santa Clara CA; described in U.S. Patent Number 5,837,458; Chang, C.-C. et al. (1999) Nat. Biotechnol. 17:793-797; Christians, F.C. et al. (1999) Nat. Biotechnol. 17:259-264; and Crameri, A. et al. (1996) Nat. Biotechnol. 14:315-319) to alter or improve the biological properties of CYSKP, such as its biological or enzymatic activity or its ability to bind to other molecules or compounds. DNA shuffling is a process by which a library of gene variants is produced using PCR-mediated recombination of gene fragments. The library is then subjected to selection or screening procedures that identify those gene variants with the desired properties. These preferred variants may then be pooled and further subjected to recursive rounds of DNA shuffling and selection/screening. Thus, genetic diversity is created through "artificial" breeding and rapid molecular evolution. For example, fragments of a single gene containing random point mutations may be recombined, screened, and then reshuffled until the desired properties are optimized. Alternatively, fragments of a given gene may be recombined with fragments of homologous genes in the same gene family, either from the same or different species, thereby maximizing the genetic diversity of multiple naturally occurring genes in a directed and controllable manner.

In another embodiment, sequences encoding CYSKP may be synthesized, in whole or in part, using chemical methods well known in the art. (See, e.g., Caruthers, M.H. et al. (1980) Nucleic Acids Symp. Ser. 7:215-223; and Horn, T. et al. (1980) Nucleic Acids Symp. Ser. 7:225-232.) Alternatively, CYSKP itself or a fragment thereof may be synthesized using chemical methods. For example, peptide synthesis can be performed using various solution-phase or solid-phase techniques. (See, e.g., Creighton, T. (1984) Proteins, Structures and Molecular Properties, WH Freeman, New York NY, pp. 55-60; and Roberge, J.Y. et al. (1995) Science 269:202-204.) Automated synthesis may be achieved using the ABI 431A peptide synthesizer (Applied Biosystems). Additionally, the amino acid sequence of CYSKP, or any part thereof, may be altered during direct synthesis and/or combined with sequences from other proteins, or any part thereof, to produce a variant polypeptide or a polypeptide having a sequence of a naturally occurring polypeptide.

The peptide may be substantially purified by preparative high performance liquid chromatography. (See, e.g., Chiez, R.M. and F.Z. Regnier (1990) Methods Enzymol. 182:392-421.) The composition of the synthetic peptides may be confirmed by amino acid analysis or by sequencing. (See, e.g., Creighton, supra, pp. 28-53.)

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In order to express a biologically active CYSKP, the nucleotide sequences encoding CYSKP or derivatives thereof may be inserted into an appropriate expression vector, i.e., a vector which contains the necessary elements for transcriptional and translational control of the inserted coding sequence in a suitable host. These elements include regulatory sequences, such as enhancers, constitutive and inducible promoters, and 5' and 3' untranslated regions in the vector and in polynucleotide sequences encoding CYSKP. Such elements may vary in their strength and specificity. Specific initiation signals may also be used to achieve more efficient translation of sequences encoding CYSKP. Such signals include the ATG initiation codon and adjacent sequences, e.g. the Kozak sequence. In cases where sequences encoding CYSKP and its initiation codon and upstream regulatory sequences are inserted into the appropriate expression vector, no additional transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a fragment thereof, is inserted, exogenous translational control signals including an in-frame ATG initiation codon should be provided by the vector. Exogenous translational elements and initiation codons may be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers appropriate for the particular host cell system used. (See, e.g., Scharf, D. et al. (1994) Results Probl. Cell Differ. 20:125-162.)

Methods which are well known to those skilled in the art may be used to construct expression vectors containing sequences encoding CYSKP and appropriate transcriptional and translational control elements. These methods include <u>in vitro</u> recombinant DNA techniques, synthetic techniques, and <u>in</u>

<u>vivo</u> genetic recombination. (See, e.g., Sambrook, J. et al. (1989) <u>Molecular Cloning</u>, A <u>Laboratory</u> <u>Manual</u>, Cold Spring Harbor Press, Plainview NY, ch. 4, 8, and 16-17; Ausubel, F.M. et al. (1995) <u>Current Protocols in Molecular Biology</u>, John Wiley & Sons, New York NY, ch. 9, 13, and 16.)

A variety of expression vector/host systems may be utilized to contain and express sequences encoding CYSKP. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; yeast transformed with yeast expression vectors; insect cell systems infected with viral expression vectors (e.g., baculovirus); plant cell systems transformed with viral expression vectors (e.g., cauliflower mosaic virus, CaMV, or tobacco mosaic virus, TMV) or with bacterial expression vectors (e.g., Ti or pBR322 plasmids); or animal cell systems. (See, e.g., Sambrook, supra; Ausubel, supra; Van Heeke, G. and S.M. Schuster (1989) J. Biol, Chem, 264:5503-5509; Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945; Takamatsu, N. (1987) EMBO J. 6:307-311; The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196; Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659; and Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355.) Expression vectors derived from retroviruses, adenoviruses, or herpes or vaccinia viruses, or from various bacterial plasmids, may be used for delivery of nucleotide sequences to the targeted organ, tissue, or cell population. (See, e.g., Di Nicola, M. et al. (1998) Cancer Gen. Ther. 5(6):350-356; Yu, M. et al. (1993) Proc. Natl. Acad. Sci. USA 90(13):6340-6344; Buller, R.M. et al. (1985) Nature 317(6040):813-815; McGregor, D.P. et al. (1994) Mol. Immunol. 31(3):219-226; and Verma, I.M. and N. Somia (1997) Nature 389:239-242.) The invention is not limited by the host cell employed.

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In bacterial systems, a number of cloning and expression vectors may be selected depending upon the use intended for polynucleotide sequences encoding CYSKP. For example, routine cloning, subcloning, and propagation of polynucleotide sequences encoding CYSKP can be achieved using a multifunctional E. coli vector such as PBLUESCRIPT (Stratagene, La Jolla CA) or PSPORT1 plasmid (Life Technologies). Ligation of sequences encoding CYSKP into the vector's multiple cloning site disrupts the *lacZ* gene, allowing a colorimetric screening procedure for identification of transformed bacteria containing recombinant molecules. In addition, these vectors may be useful for <u>in vitro</u> transcription, dideoxy sequencing, single strand rescue with helper phage, and creation of nested deletions in the cloned sequence. (See, e.g., Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509.) When large quantities of CYSKP are needed, e.g. for the production of antibodies, vectors which direct high level expression of CYSKP may be used. For example, vectors containing the strong, inducible SP6 or T7 bacteriophage promoter may be used.

Yeast expression systems may be used for production of CYSKP. A number of vectors

containing constitutive or inducible promoters, such as alpha factor, alcohol oxidase, and PGH promoters, may be used in the yeast <u>Saccharomyces cerevisiae</u> or <u>Pichia pastoris</u>. In addition, such vectors direct either the secretion or intracellular retention of expressed proteins and enable integration of foreign sequences into the host genome for stable propagation. (See, e.g., Ausubel, 1995, <u>supra;</u> Bitter, G.A. et al. (1987) Methods Enzymol. 153:516-544; and Scorer, C.A. et al. (1994) Bio/Technology 12:181-184.)

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Plant systems may also be used for expression of CYSKP. Transcription of sequences encoding CYSKP may be driven by viral promoters, e.g., the 35S and 19S promoters of CaMV used alone or in combination with the omega leader sequence from TMV (Takamatsu, N. (1987) EMBO J. 6:307-311). Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters may be used. (See, e.g., Coruzzi, G. et al. (1984) EMBO J. 3:1671-1680; Broglie, R. et al. (1984) Science 224:838-843; and Winter, J. et al. (1991) Results Probl. Cell Differ. 17:85-105.) These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection. (See, e.g., The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196.)

In mammalian cells, a number of viral-based expression systems may be utilized. In cases where an adenovirus is used as an expression vector, sequences encoding CYSKP may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to obtain infective virus which expresses CYSKP in host cells. (See, e.g., Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659.) In addition, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells. SV40 or EBV-based vectors may also be used for high-level protein expression.

Human artificial chromosomes (HACs) may also be employed to deliver larger fragments of DNA than can be contained in and expressed from a plasmid. HACs of about 6 kb to 10 Mb are constructed and delivered via conventional delivery methods (liposomes, polycationic amino polymers, or vesicles) for therapeutic purposes. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355.)

For long term production of recombinant proteins in mammalian systems, stable expression of CYSKP in cell lines is preferred. For example, sequences encoding CYSKP can be transformed into cell lines using expression vectors which may contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells may be allowed to grow for about 1 to 2 days in enriched media before being switched to selective media. The purpose of the selectable marker is to confer resistance to a selective agent, and its presence allows growth and recovery of cells which successfully express the

introduced sequences. Resistant clones of stably transformed cells may be propagated using tissue culture techniques appropriate to the cell type.

Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase and adenine phosphoribosyltransferase genes, for use in *tk* and *apr* cells, respectively. (See, e.g., Wigler, M. et al. (1977) Cell 11:223-232; Lowy, I. et al. (1980) Cell 22:817-823.) Also, antimetabolite, antibiotic, or herbicide resistance can be used as the basis for selection. For example, *dhfr* confers resistance to methotrexate; *neo* confers resistance to the aminoglycosides neomycin and G-418; and *als* and *pat* confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively. (See, e.g., Wigler, M. et al. (1980) Proc. Natl. Acad. Sci. USA 77:3567-3570; Colbere-Garapin, F. et al. (1981) J. Mol. Biol. 150:1-14.) Additional selectable genes have been described, e.g., *trpB* and *hisD*, which alter cellular requirements for metabolites. (See, e.g., Hartman, S.C. and R.C. Mulligan (1988) Proc. Natl. Acad. Sci. USA 85:8047-8051.) Visible markers, e.g., anthocyanins, green fluorescent proteins (GFP; Clontech), ß glucuronidase and its substrate ß-glucuronide, or luciferase and its substrate luciferin may be used. These markers can be used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system. (See, e.g., Rhodes, C.A. (1995) Methods Mol. Biol. 55:121-131.)

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Although the presence/absence of marker gene expression suggests that the gene of interest is also present, the presence and expression of the gene may need to be confirmed. For example, if the sequence encoding CYSKP is inserted within a marker gene sequence, transformed cells containing sequences encoding CYSKP can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a sequence encoding CYSKP under the control of a single promoter. Expression of the marker gene in response to induction or selection usually indicates expression of the tandem gene as well.

In general, host cells that contain the nucleic acid sequence encoding CYSKP and that express CYSKP may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations, PCR amplification, and protein bioassay or immunoassay techniques which include membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein sequences.

Immunological methods for detecting and measuring the expression of CYSKP using either specific polyclonal or monoclonal antibodies are known in the art. Examples of such techniques include enzyme-linked immunosorbent assays (ELISAs), radioimmunoassays (RIAs), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on CYSKP is preferred, but a competitive binding

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assay may be employed. These and other assays are well known in the art. (See, e.g., Hampton, R. et al. (1990) <u>Serological Methods, a Laboratory Manual</u>, APS Press, St. Paul MN, Sect. IV; Coligan, J.E. et al. (1997) <u>Current Protocols in Immunology</u>, Greene Pub. Associates and Wiley-Interscience, New York NY; and Pound, J.D. (1998) <u>Immunochemical Protocols</u>, Humana Press, Totowa NJ.)

A wide variety of labels and conjugation techniques are known by those skilled in the art and may be used in various nucleic acid and amino acid assays. Means for producing labeled hybridization or PCR probes for detecting sequences related to polynucleotides encoding CYSKP include oligolabeling, nick translation, end-labeling, or PCR amplification using a labeled nucleotide.

Alternatively, the sequences encoding CYSKP, or any fragments thereof, may be cloned into a vector for the production of an mRNA probe. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by addition of an appropriate RNA polymerase such as T7, T3, or SP6 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits, such as those provided by Amersham Pharmacia Biotech, Promega (Madison WI), and US Biochemical. Suitable reporter molecules or labels which may be used for ease of detection include radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents, as well as substrates, cofactors, inhibitors, magnetic particles, and the like.

Host cells transformed with nucleotide sequences encoding CYSKP may be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a transformed cell may be secreted or retained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing polynucleotides which encode CYSKP may be designed to contain signal sequences which direct secretion of CYSKP through a prokaryotic or eukaryotic cell membrane.

In addition, a host cell strain may be chosen for its ability to modulate expression of the inserted sequences or to process the expressed protein in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" or "pro" form of the protein may also be used to specify protein targeting, folding, and/or activity. Different host cells which have specific cellular machinery and characteristic mechanisms for post-translational activities (e.g., CHO, HeLa, MDCK, HEK293, and WI38) are available from the American Type Culture Collection (ATCC, Manassas VA) and may be chosen to ensure the correct modification and processing of the foreign protein.

In another embodiment of the invention, natural, modified, or recombinant nucleic acid sequences encoding CYSKP may be ligated to a heterologous sequence resulting in translation of a fusion protein in any of the aforementioned host systems. For example, a chimeric CYSKP protein

containing a heterologous moiety that can be recognized by a commercially available antibody may facilitate the screening of peptide libraries for inhibitors of CYSKP activity. Heterologous protein and peptide moieties may also facilitate purification of fusion proteins using commercially available affinity matrices. Such moieties include, but are not limited to, glutathione S-transferase (GST), maltose binding protein (MBP), thioredoxin (Trx), calmodulin binding peptide (CBP), 6-His, FLAG, *c-myc*, and hemagglutinin (HA). GST, MBP, Trx, CBP, and 6-His enable purification of their cognate fusion proteins on immobilized glutathione, maltose, phenylarsine oxide, calmodulin, and metal-chelate resins, respectively. FLAG, *c-myc*, and hemagglutinin (HA) enable immunoaffinity purification of fusion proteins using commercially available monoclonal and polyclonal antibodies that specifically recognize these epitope tags. A fusion protein may also be engineered to contain a proteolytic cleavage site located between the CYSKP encoding sequence and the heterologous protein sequence, so that CYSKP may be cleaved away from the heterologous moiety following purification. Methods for fusion protein expression and purification are discussed in Ausubel (1995, <u>supra</u>, ch. 10). A variety of commercially available kits may also be used to facilitate expression and purification of fusion proteins.

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In a further embodiment of the invention, synthesis of radiolabeled CYSKP may be achieved <u>in vitro</u> using the TNT rabbit reticulocyte lysate or wheat germ extract system (Promega). These systems couple transcription and translation of protein-coding sequences operably associated with the T7, T3, or SP6 promoters. Translation takes place in the presence of a radiolabeled amino acid precursor, for example, <sup>35</sup>S-methionine.

CYSKP of the present invention or fragments thereof may be used to screen for compounds that specifically bind to CYSKP. At least one and up to a plurality of test compounds may be screened for specific binding to CYSKP. Examples of test compounds include antibodies, oligonucleotides, proteins (e.g., receptors), or small molecules.

In one embodiment, the compound thus identified is closely related to the natural ligand of CYSKP, e.g., a ligand or fragment thereof, a natural substrate, a structural or functional mimetic, or a natural binding partner. (See, e.g., Coligan, J.E. et al. (1991) <u>Current Protocols in Immunology</u> 1(2): Chapter 5.) Similarly, the compound can be closely related to the natural receptor to which CYSKP binds, or to at least a fragment of the receptor, e.g., the ligand binding site. In either case, the compound can be rationally designed using known techniques. In one embodiment, screening for these compounds involves producing appropriate cells which express CYSKP, either as a secreted protein or on the cell membrane. Preferred cells include cells from mammals, yeast, <u>Drosophila</u>, or <u>E. coli</u>. Cells expressing CYSKP or cell membrane fractions which contain CYSKP are then contacted with a test compound and binding, stimulation, or inhibition of activity of either CYSKP or the compound is analyzed.

An assay may simply test binding of a test compound to the polypeptide, wherein binding is detected by a fluorophore, radioisotope, enzyme conjugate, or other detectable label. For example, the assay may comprise the steps of combining at least one test compound with CYSKP, either in solution or affixed to a solid support, and detecting the binding of CYSKP to the compound. Alternatively, the assay may detect or measure binding of a test compound in the presence of a labeled competitor. Additionally, the assay may be carried out using cell-free preparations, chemical libraries, or natural product mixtures, and the test compound(s) may be free in solution or affixed to a solid support.

CYSKP of the present invention or fragments thereof may be used to screen for compounds that modulate the activity of CYSKP. Such compounds may include agonists, antagonists, or partial or inverse agonists. In one embodiment, an assay is performed under conditions permissive for CYSKP activity, wherein CYSKP is combined with at least one test compound, and the activity of CYSKP in the presence of a test compound is compared with the activity of CYSKP in the absence of the test compound. A change in the activity of CYSKP in the presence of the test compound is indicative of a compound that modulates the activity of CYSKP. Alternatively, a test compound is combined with an <u>in vitro</u> or cell-free system comprising CYSKP under conditions suitable for CYSKP activity, and the assay is performed. In either of these assays, a test compound which modulates the activity of CYSKP may do so indirectly and need not come in direct contact with the test compound. At least one and up to a plurality of test compounds may be screened.

In another embodiment, polynucleotides encoding CYSKP or their mammalian homologs may be "knocked out" in an animal model system using homologous recombination in embryonic stem (ES) cells. Such techniques are well known in the art and are useful for the generation of animal models of human disease. (See, e.g., U.S. Patent Number 5,175,383 and U.S. Patent Number 5,767,337.) For example, mouse ES cells, such as the mouse 129/SvJ cell line, are derived from the early mouse embryo and grown in culture. The ES cells are transformed with a vector containing the gene of interest disrupted by a marker gene, e.g., the neomycin phosphotransferase gene (neo; Capecchi, M.R. (1989) Science 244:1288-1292). The vector integrates into the corresponding region of the host genome by homologous recombination. Alternatively, homologous recombination takes place using the Cre-loxP system to knockout a gene of interest in a tissue- or developmental stage-specific manner (Marth, J.D. (1996) Clin. Invest. 97:1999-2002; Wagner, K.U. et al. (1997) Nucleic Acids Res. 25:4323-4330). Transformed ES cells are identified and microinjected into mouse cell blastocysts such as those from the C57BL/6 mouse strain. The blastocysts are surgically transferred to pseudopregnant dams, and the resulting chimeric progeny are genotyped and bred to produce heterozygous or homozygous strains. Transgenic animals thus generated may be tested with potential therapeutic or toxic agents.

Polynucleotides encoding CYSKP may also be manipulated <u>in vitro</u> in ES cells derived from human blastocysts. Human ES cells have the potential to differentiate into at least eight separate cell lineages including endoderm, mesoderm, and ectodermal cell types. These cell lineages differentiate into, for example, neural cells, hematopoietic lineages, and cardiomyocytes (Thomson, J.A. et al. (1998) Science 282:1145-1147).

Polynucleotides encoding CYSKP can also be used to create "knockin" humanized animals (pigs) or transgenic animals (mice or rats) to model human disease. With knockin technology, a region of a polynucleotide encoding CYSKP is injected into animal ES cells, and the injected sequence integrates into the animal cell genome. Transformed cells are injected into blastulae, and the blastulae are implanted as described above. Transgenic progeny or inbred lines are studied and treated with potential pharmaceutical agents to obtain information on treatment of a human disease. Alternatively, a mammal inbred to overexpress CYSKP, e.g., by secreting CYSKP in its milk, may also serve as a convenient source of that protein (Janne, J. et al. (1998) Biotechnol. Annu. Rev. 4:55-74).

#### **THERAPEUTICS**

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Chemical and structural similarity, e.g., in the context of sequences and motifs, exists between regions of CYSKP and cytoskeleton-associated proteins. In addition, the expression of CYSKP is closely associated with lung, reproductive (including placenta), neural (including brain), adrenal, endothelial, kidney, and spleen tissue, as well as with ovarian, breast, and testicular tumor tissue. Therefore, CYSKP appears to play a role in cell proliferative, autoimmune/inflammatory, vesicle trafficking, neurological, cell motility, reproductive, and muscle disorders. In the treatment of disorders associated with increased CYSKP expression or activity, it is desirable to decrease the expression or activity of CYSKP. In the treatment of disorders associated with decreased CYSKP expression or activity, it is desirable to increase the expression or activity of CYSKP.

Therefore, in one embodiment, CYSKP or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of CYSKP. Examples of such disorders include, but are not limited to, a cell proliferative disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus; an autoimmune/inflammatory disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory

distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasisectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, 5 Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, 10 systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; a vesicle trafficking disorder such as cystic fibrosis, glucose-galactose malabsorption syndrome, hypercholesterolemia, diabetes mellitus, diabetes insipidus, hyper- and hypoglycemia, Grave's disease, goiter, Cushing's disease, and Addison's disease, 15 gastrointestinal disorders including ulcerative colitis, gastric and duodenal ulcers, other conditions associated with abnormal vesicle trafficking, including acquired immunodeficiency syndrome (AIDS), allergies including hay fever, asthma, and urticaria (hives), autoimmune hemolytic anemia, proliferative glomerulonephritis, inflammatory bowel disease, multiple sclerosis, myasthenia gravis, rheumatoid and osteoarthritis, scleroderma, Chediak-Higashi and Sjogren's syndromes, systemic lupus erythematosus, 20 toxic shock syndrome, traumatic tissue damage, and viral, bacterial, fungal, helminthic, and protozoal infections; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and 25 other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal 30 syndrome, mental retardation and other developmental disorders of the central nervous system including Down syndrome, cerebral palsy, neuroskeletal disorders, autonomic nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis, inherited, metabolic, endocrine, and toxic myopathies, myasthenia gravis, periodic paralysis, mental disorders including mood, anxiety,

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and schizophrenic disorders, seasonal affective disorder (SAD), akathesia, amnesia, catatonia, diabetic neuropathy, tardive dyskinesia, dystonias, paranoid psychoses, postherpetic neuralgia, Tourette's disorder, progressive supranuclear palsy, corticobasal degeneration, and familial frontotemporal dementia; a cell motility disorder such as ankylosing spondylitis, Chediak-Higashi syndrome, Duchenne and Becker muscular dystrophy, intrahepatic cholestasis, myocardial hyperplasia, cardiomyopathy, early onset peridontitis, cancers such as adenocarcinoma, ovarian carcinoma, and chronic myelogenous leukemia, and bacterial and helminthic infections; a reproductive disorder such as a disorder of prolactin production, infertility, including tubal disease, ovulatory defects, endometriosis, a disruption of the estrous cycle, a disruption of the menstrual cycle, polycystic ovary syndrome, ovarian hyperstimulation syndrome, an endometrial or ovarian tumor, a uterine fibroid, autoimmune disorders, ectopic pregnancy, teratogenesis, cancer of the breast, fibrocystic breast disease, galactorrhea, a disruption of spermatogenesis, abnormal sperm physiology, cancer of the testis, cancer of the prostate, benign prostatic hyperplasia, prostatitis, Peyronie's disease, impotence, carcinoma of the male breast, gynecomastia, hypergonadotropic and hypogonadotropic hypogonadism, pseudohermaphroditism, azoospermia, premature ovarian failure, acrosin deficiency, delayed puperty, retrograde ejaculation and anejaculation, haemangioblastomas, cystsphaeochromocytomas, paraganglioma, cystadenomas of the epididymis, and endolymphatic sac tumours; and a muscle disorder such as myocarditis, Duchenne's muscular dystrophy, Becker's muscular dystrophy, myotonic dystrophy, central core disease, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, infectious myositis, polymyositis, dermatomyositis, inclusion body myositis, thyrotoxic myopathy, and ethanol myopathy, angina, anaphylactic shock, arrhythmias, asthma, cardiovascular shock, Cushing's syndrome, hypertension, hypoglycemia, myocardial infarction, migraine, and pheochromocytoma, and myopathies including cardiomyopathy, encephalopathy, epilepsy, Kearns-Sayre syndrome, lactic acidosis, myoclonic disorder, and ophthalmoplegia.

In another embodiment, a vector capable of expressing CYSKP or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of CYSKP including, but not limited to, those described above.

In a further embodiment, a composition comprising a substantially purified CYSKP in conjunction with a suitable pharmaceutical carrier may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of CYSKP including, but not limited to, those provided above.

In still another embodiment, an agonist which modulates the activity of CYSKP may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of CYSKP including, but not limited to, those listed above.

In a further embodiment, an antagonist of CYSKP may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of CYSKP. Examples of such disorders include, but are not limited to, those cell proliferative, autoimmune/inflammatory, vesicle trafficking, neurological, cell motility, reproductive, and muscle disorders described above. In one aspect, an antibody which specifically binds CYSKP may be used directly as an antagonist or indirectly as a targeting or delivery mechanism for bringing a pharmaceutical agent to cells or tissues which express CYSKP.

In an additional embodiment, a vector expressing the complement of the polynucleotide encoding CYSKP may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of CYSKP including, but not limited to, those described above.

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In other embodiments, any of the proteins, antagonists, antibodies, agonists, complementary sequences, or vectors of the invention may be administered in combination with other appropriate therapeutic agents. Selection of the appropriate agents for use in combination therapy may be made by one of ordinary skill in the art, according to conventional pharmaceutical principles. The combination of therapeutic agents may act synergistically to effect the treatment or prevention of the various disorders described above. Using this approach, one may be able to achieve therapeutic efficacy with lower dosages of each agent, thus reducing the potential for adverse side effects.

An antagonist of CYSKP may be produced using methods which are generally known in the art. In particular, purified CYSKP may be used to produce antibodies or to screen libraries of pharmaceutical agents to identify those which specifically bind CYSKP. Antibodies to CYSKP may also be generated using methods that are well known in the art. Such antibodies may include, but are not limited to, polyclonal, monoclonal, chimeric, and single chain antibodies, Fab fragments, and fragments produced by a Fab expression library. Neutralizing antibodies (i.e., those which inhibit dimer formation) are generally preferred for therapeutic use.

For the production of antibodies, various hosts including goats, rabbits, rats, mice, humans, and others may be immunized by injection with CYSKP or with any fragment or oligopeptide thereof which has immunogenic properties. Depending on the host species, various adjuvants may be used to increase immunological response. Such adjuvants include, but are not limited to, Freund's, mineral gels such as aluminum hydroxide, and surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, KLH, and dinitrophenol. Among adjuvants used in humans, BCG (bacilli Calmette-Guerin) and Corynebacterium parvum are especially preferable.

It is preferred that the oligopeptides, peptides, or fragments used to induce antibodies to CYSKP have an amino acid sequence consisting of at least about 5 amino acids, and generally will consist of at least about 10 amino acids. It is also preferable that these oligopeptides, peptides, or

fragments are identical to a portion of the amino acid sequence of the natural protein. Short stretches of CYSKP amino acids may be fused with those of another protein, such as KLH, and antibodies to the chimeric molecule may be produced.

Monoclonal antibodies to CYSKP may be prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique, the human B-cell hybridoma technique, and the EBV-hybridoma technique. (See, e.g., Kohler, G. et al. (1975) Nature 256:495-497; Kozbor, D. et al. (1985) J. Immunol. Methods 81:31-42; Cote, R.J. et al. (1983) Proc. Natl. Acad. Sci. USA 80:2026-2030; and Cole, S.P. et al. (1984) Mol. Cell Biol. 62:109-120.)

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In addition, techniques developed for the production of "chimeric antibodies," such as the splicing of mouse antibody genes to human antibody genes to obtain a molecule with appropriate antigen specificity and biological activity, can be used. (See, e.g., Morrison, S.L. et al. (1984) Proc. Natl. Acad. Sci. USA 81:6851-6855; Neuberger, M.S. et al. (1984) Nature 312:604-608; and Takeda, S. et al. (1985) Nature 314:452-454.) Alternatively, techniques described for the production of single chain antibodies may be adapted, using methods known in the art, to produce CYSKP-specific single chain antibodies. Antibodies with related specificity, but of distinct idiotypic composition, may be generated by chain shuffling from random combinatorial immunoglobulin libraries. (See, e.g., Burton, D.R. (1991) Proc. Natl. Acad. Sci. USA 88:10134-10137.)

Antibodies may also be produced by inducing <u>in vivo</u> production in the lymphocyte population or by screening immunoglobulin libraries or panels of highly specific binding reagents as disclosed in the literature. (See, e.g., Orlandi, R. et al. (1989) Proc. Natl. Acad. Sci. USA 86:3833-3837; Winter, G. et al. (1991) Nature 349:293-299.)

Antibody fragments which contain specific binding sites for CYSKP may also be generated. For example, such fragments include, but are not limited to,  $F(ab')_2$  fragments produced by pepsin digestion of the antibody molecule and Fab fragments generated by reducing the disulfide bridges of the  $F(ab')_2$  fragments. Alternatively, Fab expression libraries may be constructed to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity. (See, e.g., Huse, W.D. et al. (1989) Science 246:1275-1281.)

Various immunoassays may be used for screening to identify antibodies having the desired specificity. Numerous protocols for competitive binding or immunoradiometric assays using either polyclonal or monoclonal antibodies with established specificities are well known in the art. Such immunoassays typically involve the measurement of complex formation between CYSKP and its specific antibody. A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering CYSKP epitopes is generally used, but a competitive binding assay may also be

employed (Pound, supra).

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Various methods such as Scatchard analysis in conjunction with radioimmunoassay techniques may be used to assess the affinity of antibodies for CYSKP. Affinity is expressed as an association constant,  $K_a$ , which is defined as the molar concentration of CYSKP-antibody complex divided by the molar concentrations of free antigen and free antibody under equilibrium conditions. The  $K_a$  determined for a preparation of polyclonal antibodies, which are heterogeneous in their affinities for multiple CYSKP epitopes, represents the average affinity, or avidity, of the antibodies for CYSKP. The  $K_a$  determined for a preparation of monoclonal antibodies, which are monospecific for a particular CYSKP epitope, represents a true measure of affinity. High-affinity antibody preparations with  $K_a$  ranging from about  $10^9$  to  $10^{12}$  L/mole are preferred for use in immunoassays in which the CYSKP-antibody complex must withstand rigorous manipulations. Low-affinity antibody preparations with  $K_a$  ranging from about  $10^6$  to  $10^7$  L/mole are preferred for use in immunopurification and similar procedures which ultimately require dissociation of CYSKP, preferably in active form, from the antibody (Catty, D. (1988) Antibodies, Volume I: A Practical Approach, IRL Press, Washington DC; Liddell, J.E. and A. Cryer (1991) A Practical Guide to Monoclonal Antibodies, John Wiley & Sons, New York NY).

The titer and avidity of polyclonal antibody preparations may be further evaluated to determine the quality and suitability of such preparations for certain downstream applications. For example, a polyclonal antibody preparation containing at least 1-2 mg specific antibody/ml, preferably 5-10 mg specific antibody/ml, is generally employed in procedures requiring precipitation of CYSKP-antibody complexes. Procedures for evaluating antibody specificity, titer, and avidity, and guidelines for antibody quality and usage in various applications, are generally available. (See, e.g., Catty, <u>supra</u>, and Coligan et al. <u>supra</u>.)

In another embodiment of the invention, the polynucleotides encoding CYSKP, or any fragment or complement thereof, may be used for therapeutic purposes. In one aspect, modifications of gene expression can be achieved by designing complementary sequences or antisense molecules (DNA, RNA, PNA, or modified oligonucleotides) to the coding or regulatory regions of the gene encoding CYSKP. Such technology is well known in the art, and antisense oligonucleotides or larger fragments can be designed from various locations along the coding or control regions of sequences encoding CYSKP. (See, e.g., Agrawal, S., ed. (1996) Antisense Therapeutics, Humana Press Inc., Totawa NJ.)

In therapeutic use, any gene delivery system suitable for introduction of the antisense sequences into appropriate target cells can be used. Antisense sequences can be delivered intracellularly in the form of an expression plasmid which, upon transcription, produces a sequence complementary to at least a portion of the cellular sequence encoding the target protein. (See, e.g., Slater, J.E. et al. (1998) J. Allergy Cli. Immunol. 102(3):469-475; and Scanlon, K.J. et al. (1995)

9(13):1288-1296.) Antisense sequences can also be introduced intracellularly through the use of viral vectors, such as retrovirus and adeno-associated virus vectors. (See, e.g., Miller, A.D. (1990) Blood 76:271; Ausubel, <u>supra</u>; Uckert, W. and W. Walther (1994) Pharmacol. Ther. 63(3):323-347.) Other gene delivery mechanisms include liposome-derived systems, artificial viral envelopes, and other systems known in the art. (See, e.g., Rossi, J.J. (1995) Br. Med. Bull. 51(1):217-225; Boado, R.J. et al. (1998) J. Pharm. Sci. 87(11):1308-1315; and Morris, M.C. et al. (1997) Nucleic Acids Res. 25(14):2730-2736.)

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In another embodiment of the invention, polynucleotides encoding CYSKP may be used for somatic or germline gene therapy. Gene therapy may be performed to (i) correct a genetic deficiency (e.g., in the cases of severe combined immunodeficiency (SCID)-X1 disease characterized by X-linked inheritance (Cavazzana-Calvo, M. et al. (2000) Science 288:669-672), severe combined immunodeficiency syndrome associated with an inherited adenosine deaminase (ADA) deficiency (Blaese, R.M. et al. (1995) Science 270:475-480; Bordignon, C. et al. (1995) Science 270:470-475), cystic fibrosis (Zabner, J. et al. (1993) Cell 75:207-216; Crystal, R.G. et al. (1995) Hum, Gene Therapy 6:643-666; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:667-703), thalassamias, familial hypercholesterolemia, and hemophilia resulting from Factor VIII or Factor IX deficiencies (Crystal, R.G. (1995) Science 270:404-410; Verma, I.M. and N. Somia (1997) Nature 389:239-242)), (ii) express a conditionally lethal gene product (e.g., in the case of cancers which result from unregulated cell proliferation), or (iii) express a protein which affords protection against intracellular parasites (e.g., against human retroviruses, such as human immunodeficiency virus (HIV) (Baltimore, D. (1988) Nature 335:395-396; Poeschla, E. et al. (1996) Proc. Natl. Acad. Sci. USA. 93:11395-11399), hepatitis B or C virus (HBV, HCV); fungal parasites, such as Candida albicans and Paracoccidioides brasiliensis; and protozoan parasites such as Plasmodium falciparum and Trypanosoma cruzi). In the case where a genetic deficiency in CYSKP expression or regulation causes disease, the expression of CYSKP from an appropriate population of transduced cells may alleviate the clinical manifestations caused by the genetic deficiency.

In a further embodiment of the invention, diseases or disorders caused by deficiencies in CYSKP are treated by constructing mammalian expression vectors encoding CYSKP and introducing these vectors by mechanical means into CYSKP-deficient cells. Mechanical transfer technologies for use with cells <u>in vivo</u> or <u>ex vitro</u> include (i) direct DNA microinjection into individual cells, (ii) ballistic gold particle delivery, (iii) liposome-mediated transfection, (iv) receptor-mediated gene transfer, and (v) the use of DNA transposons (Morgan, R.A. and W.F. Anderson (1993) Annu. Rev. Biochem. 62:191-217; Ivics, Z. (1997) Cell 91:501-510; Boulay, J-L. and H. Récipon (1998) Curr. Opin. Biotechnol. 9:445-450).

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Expression vectors that may be effective for the expression of CYSKP include, but are not limited to, the PCDNA 3.1, EPITAG, PRCCMV2, PREP, PVAX vectors (Invitrogen, Carlsbad CA), PCMV-SCRIPT, PCMV-TAG, PEGSH/PERV (Stratagene, La Jolla CA), and PTET-OFF, PTET-ON, PTRE2, PTRE2-LUC, PTK-HYG (Clontech, Palo Alto CA). CYSKP may be expressed using (i) a constitutively active promoter, (e.g., from cytomegalovirus (CMV), Rous sarcoma virus (RSV), SV40 virus, thymidine kinase (TK), or β-actin genes), (ii) an inducible promoter (e.g., the tetracycline-regulated promoter (Gossen, M. and H. Bujard (1992) Proc. Natl. Acad. Sci. USA 89:5547-5551; Gossen, M. et al. (1995) Science 268:1766-1769; Rossi, F.M.V. and H.M. Blau (1998) Curr. Opin. Biotechnol. 9:451-456), commercially available in the T-REX plasmid (Invitrogen)); the ecdysone-inducible promoter (available in the plasmids PVGRXR and PIND; Invitrogen); the FK506/rapamycin inducible promoter; or the RU486/mifepristone inducible promoter (Rossi, F.M.V. and Blau, H.M. supra)), or (iii) a tissue-specific promoter or the native promoter of the endogenous gene encoding CYSKP from a normal individual.

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Commercially available liposome transformation kits (e.g., the PERFECT LIPID TRANSFECTION KIT, available from Invitrogen) allow one with ordinary skill in the art to deliver polynucleotides to target cells in culture and require minimal effort to optimize experimental parameters. In the alternative, transformation is performed using the calcium phosphate method (Graham, F.L. and A.J. Eb (1973) Virology 52:456-467), or by electroporation (Neumann, E. et al. (1982) EMBO J. 1:841-845). The introduction of DNA to primary cells requires modification of these standardized mammalian transfection protocols.

In another embodiment of the invention, diseases or disorders caused by genetic defects with respect to CYSKP expression are treated by constructing a retrovirus vector consisting of (i) the polynucleotide encoding CYSKP under the control of an independent promoter or the retrovirus long terminal repeat (LTR) promoter, (ii) appropriate RNA packaging signals, and (iii) a Rev-responsive element (RRE) along with additional retrovirus *cis*-acting RNA sequences and coding sequences required for efficient vector propagation. Retrovirus vectors (e.g., PFB and PFBNEO) are commercially available (Stratagene) and are based on published data (Riviere, I. et al. (1995) Proc. Natl. Acad. Sci. USA 92:6733-6737), incorporated by reference herein. The vector is propagated in an appropriate vector producing cell line (VPCL) that expresses an envelope gene with a tropism for receptors on the target cells or a promiscuous envelope protein such as VSVg (Armentano, D. et al. (1987) J. Virol. 61:1647-1650; Bender, M.A. et al. (1987) J. Virol. 61:1639-1646; Adam, M.A. and A.D. Miller (1988) J. Virol. 62:3802-3806; Dull, T. et al. (1998) J. Virol. 72:8463-8471; Zufferey, R. et al. (1998) J. Virol. 72:9873-9880). U.S. Patent Number 5,910,434 to Rigg ("Method for obtaining retrovirus packaging cell lines producing high transducing efficiency retroviral supernatant") discloses a

method for obtaining retrovirus packaging cell lines and is hereby incorporated by reference. Propagation of retrovirus vectors, transduction of a population of cells (e.g., CD4<sup>+</sup> T-cells), and the return of transduced cells to a patient are procedures well known to persons skilled in the art of gene therapy and have been well documented (Ranga, U. et al. (1997) J. Virol. 71:7020-7029; Bauer, G. et al. (1997) Blood 89:2259-2267; Bonyhadi, M.L. (1997) J. Virol. 71:4707-4716; Ranga, U. et al. (1998) Proc. Natl. Acad. Sci. USA 95:1201-1206; Su, L. (1997) Blood 89:2283-2290).

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In the alternative, an adenovirus-based gene therapy delivery system is used to deliver polynucleotides encoding CYSKP to cells which have one or more genetic abnormalities with respect to the expression of CYSKP. The construction and packaging of adenovirus-based vectors are well known to those with ordinary skill in the art. Replication defective adenovirus vectors have proven to be versatile for importing genes encoding immunoregulatory proteins into intact islets in the pancreas (Csete, M.E. et al. (1995) Transplantation 27:263-268). Potentially useful adenoviral vectors are described in U.S. Patent Number 5,707,618 to Armentano ("Adenovirus vectors for gene therapy"), hereby incorporated by reference. For adenoviral vectors, see also Antinozzi, P.A. et al. (1999) Annu. Rev. Nutr. 19:511-544 and Verma, I.M. and N. Somia (1997) Nature 18:389:239-242, both incorporated by reference herein.

In another alternative, a herpes-based, gene therapy delivery system is used to deliver polynucleotides encoding CYSKP to target cells which have one or more genetic abnormalities with respect to the expression of CYSKP. The use of herpes simplex virus (HSV)-based vectors may be especially valuable for introducing CYSKP to cells of the central nervous system, for which HSV has a tropism. The construction and packaging of herpes-based vectors are well known to those with ordinary skill in the art. A replication-competent herpes simplex virus (HSV) type 1-based vector has been used to deliver a reporter gene to the eyes of primates (Liu, X. et al. (1999) Exp. Eye Res. 169:385-395). The construction of a HSV-1 virus vector has also been disclosed in detail in U.S. Patent Number 5,804,413 to DeLuca ("Herpes simplex virus strains for gene transfer"), which is hereby incorporated by reference. U.S. Patent Number 5,804,413 teaches the use of recombinant HSV d92 which consists of a genome containing at least one exogenous gene to be transferred to a cell under the control of the appropriate promoter for purposes including human gene therapy. Also taught by this

For HSV vectors, see also Goins, W.F. et al. (1999) J. Virol. 73:519-532 and Xu, H. et al. (1994) Dev. Biol. 163:152-161, hereby incorporated by reference. The manipulation of cloned herpesvirus sequences, the generation of recombinant virus following the transfection of multiple plasmids containing different segments of the large herpesvirus genomes, the growth and propagation of herpesvirus, and the infection of cells with herpesvirus are techniques well known to those of ordinary

patent are the construction and use of recombinant HSV strains deleted for ICP4, ICP27 and ICP22.

skill in the art.

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In another alternative, an alphavirus (positive, single-stranded RNA virus) vector is used to deliver polynucleotides encoding CYSKP to target cells. The biology of the prototypic alphavirus, Semliki Forest Virus (SFV), has been studied extensively and gene transfer vectors have been based on the SFV genome (Garoff, H. and K.-J. Li (1998) Curr. Opin, Biotechnol. 9:464-469). During alphavirus RNA replication, a subgenomic RNA is generated that normally encodes the viral capsid proteins. This subgenomic RNA replicates to higher levels than the full length genomic RNA, resulting in the overproduction of capsid proteins relative to the viral proteins with enzymatic activity (e.g., protease and polymerase). Similarly, inserting the coding sequence for CYSKP into the alphavirus genome in place of the capsid-coding region results in the production of a large number of CYSKPcoding RNAs and the synthesis of high levels of CYSKP in vector transduced cells. While alphavirus infection is typically associated with cell lysis within a few days, the ability to establish a persistent infection in hamster normal kidney cells (BHK-21) with a variant of Sindbis virus (SIN) indicates that the lytic replication of alphaviruses can be altered to suit the needs of the gene therapy application (Dryga, S.A. et al. (1997) Virology 228:74-83). The wide host range of alphaviruses will allow the introduction of CYSKP into a variety of cell types. The specific transduction of a subset of cells in a population may require the sorting of cells prior to transduction. The methods of manipulating infectious cDNA clones of alphaviruses, performing alphavirus cDNA and RNA transfections, and performing alphavirus infections, are well known to those with ordinary skill in the art.

Oligonucleotides derived from the transcription initiation site, e.g., between about positions -10 and +10 from the start site, may also be employed to inhibit gene expression. Similarly, inhibition can be achieved using triple helix base-pairing methodology. Triple helix pairing is useful because it causes inhibition of the ability of the double helix to open sufficiently for the binding of polymerases, transcription factors, or regulatory molecules. Recent therapeutic advances using triplex DNA have been described in the literature. (See, e.g., Gee, J.E. et al. (1994) in Huber, B.E. and B.I. Carr, Molecular and Immunologic Approaches, Futura Publishing, Mt. Kisco NY, pp. 163-177.) A complementary sequence or antisense molecule may also be designed to block translation of mRNA by preventing the transcript from binding to ribosomes.

Ribozymes, enzymatic RNA molecules, may also be used to catalyze the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA, followed by endonucleolytic cleavage. For example, engineered hammerhead motif ribozyme molecules may specifically and efficiently catalyze endonucleolytic cleavage of sequences encoding CYSKP.

Specific ribozyme cleavage sites within any potential RNA target are initially identified by

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scanning the target molecule for ribozyme cleavage sites, including the following sequences: GUA, GUU, and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides, corresponding to the region of the target gene containing the cleavage site, may be evaluated for secondary structural features which may render the oligonucleotide inoperable. The suitability of candidate targets may also be evaluated by testing accessibility to hybridization with complementary oligonucleotides using ribonuclease protection assays.

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Complementary ribonucleic acid molecules and ribozymes of the invention may be prepared by any method known in the art for the synthesis of nucleic acid molecules. These include techniques for chemically synthesizing oligonucleotides such as solid phase phosphoramidite chemical synthesis. Alternatively, RNA molecules may be generated by <u>in vitro</u> and <u>in vivo</u> transcription of DNA sequences encoding CYSKP. Such DNA sequences may be incorporated into a wide variety of vectors with suitable RNA polymerase promoters such as T7 or SP6. Alternatively, these cDNA constructs that synthesize complementary RNA, constitutively or inducibly, can be introduced into cell lines, cells, or tissues.

RNA molecules may be modified to increase intracellular stability and half-life. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends of the molecule, or the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages within the backbone of the molecule. This concept is inherent in the production of PNAs and can be extended in all of these molecules by the inclusion of nontraditional bases such as inosine, queosine, and wybutosine, as well as acetyl-, methyl-, thio-, and similarly modified forms of adenine, cytidine, guanine, thymine, and uridine which are not as easily recognized by endogenous endonucleases.

An additional embodiment of the invention encompasses a method for screening for a compound which is effective in altering expression of a polynucleotide encoding CYSKP.

Compounds which may be effective in altering expression of a specific polynucleotide may include, but are not limited to, oligonucleotides, antisense oligonucleotides, triple helix-forming oligonucleotides, transcription factors and other polypeptide transcriptional regulators, and non-macromolecular chemical entities which are capable of interacting with specific polynucleotide sequences. Effective compounds may alter polynucleotide expression by acting as either inhibitors or promoters of polynucleotide expression. Thus, in the treatment of disorders associated with increased CYSKP expression or activity, a compound which specifically inhibits expression of the polynucleotide encoding CYSKP expression or activity, a compound which specifically promotes expression of the polynucleotide encoding CYSKP may be therapeutically useful.

At least one, and up to a plurality, of test compounds may be screened for effectiveness in

altering expression of a specific polynucleotide. A test compound may be obtained by any method commonly known in the art, including chemical modification of a compound known to be effective in altering polynucleotide expression; selection from an existing, commercially-available or proprietary library of naturally-occurring or non-natural chemical compounds; rational design of a compound based on chemical and/or structural properties of the target polynucleotide; and selection from a library of chemical compounds created combinatorially or randomly. A sample comprising a polynucleotide encoding CYSKP is exposed to at least one test compound thus obtained. The sample may comprise, for example, an intact or permeabilized cell, or an in vitro cell-free or reconstituted biochemical system. Alterations in the expression of a polynucleotide encoding CYSKP are assayed by any method commonly known in the art. Typically, the expression of a specific nucleotide is detected by hybridization with a probe having a nucleotide sequence complementary to the sequence of the polynucleotide encoding CYSKP. The amount of hybridization may be quantified, thus forming the basis for a comparison of the expression of the polynucleotide both with and without exposure to one or more test compounds. Detection of a change in the expression of a polynucleotide exposed to a test compound indicates that the test compound is effective in altering the expression of the polynucleotide. A screen for a compound effective in altering expression of a specific polynucleotide can be carried out, for example, using a Schizosaccharomyces pombe gene expression system (Atkins, D. et al. (1999) U.S. Patent No. 5,932,435; Arndt, G.M. et al. (2000) Nucleic Acids Res. 28:E15) or a human cell line such as HeLa cell (Clarke, M.L. et al. (2000) Biochem, Biophys. Res. Commun. 268:8-13). A particular embodiment of the present invention involves screening a combinatorial library of oligonucleotides (such as deoxyribonucleotides, ribonucleotides, peptide nucleic acids, and modified oligonucleotides) for antisense activity against a specific polynucleotide sequence (Bruice, T.W. et al. (1997) U.S. Patent No. 5,686,242; Bruice, T.W. et al. (2000) U.S. Patent No. 6,022,691).

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Many methods for introducing vectors into cells or tissues are available and equally suitable for use <u>in vivo</u>, <u>in vitro</u>, and <u>ex vivo</u>. For <u>ex vivo</u> therapy, vectors may be introduced into stem cells taken from the patient and clonally propagated for autologous transplant back into that same patient. Delivery by transfection, by liposome injections, or by polycationic amino polymers may be achieved using methods which are well known in the art. (See, e.g., Goldman, C.K. et al. (1997) Nat. Biotechnol. 15:462-466.)

Any of the therapeutic methods described above may be applied to any subject in need of such therapy, including, for example, mammals such as humans, dogs, cats, cows, horses, rabbits, and monkeys.

An additional embodiment of the invention relates to the administration of a composition which generally comprises an active ingredient formulated with a pharmaceutically acceptable excipient.

Excipients may include, for example, sugars, starches, celluloses, gums, and proteins. Various formulations are commonly known and are thoroughly discussed in the latest edition of <u>Remington's Pharmaceutical Sciences</u> (Maack Publishing, Easton PA). Such compositions may consist of CYSKP, antibodies to CYSKP, and mimetics, agonists, antagonists, or inhibitors of CYSKP.

The compositions utilized in this invention may be administered by any number of routes including, but not limited to, oral, intravenous, intramuscular, intra-arterial, intramedullary, intrathecal, intraventricular, pulmonary, transdermal, subcutaneous, intraperitoneal, intranasal, enteral, topical, sublingual, or rectal means.

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Compositions for pulmonary administration may be prepared in liquid or dry powder form. These compositions are generally aerosolized immediately prior to inhalation by the patient. In the case of small molecules (e.g. traditional low molecular weight organic drugs), aerosol delivery of fast-acting formulations is well-known in the art. In the case of macromolecules (e.g. larger peptides and proteins), recent developments in the field of pulmonary delivery via the alveolar region of the lung have enabled the practical delivery of drugs such as insulin to blood circulation (see, e.g., Patton, J.S. et al., U.S. Patent No. 5,997,848). Pulmonary delivery has the advantage of administration without needle injection, and obviates the need for potentially toxic penetration enhancers.

Compositions suitable for use in the invention include compositions wherein the active ingredients are contained in an effective amount to achieve the intended purpose. The determination of an effective dose is well within the capability of those skilled in the art.

Specialized forms of compositions may be prepared for direct intracellular delivery of macromolecules comprising CYSKP or fragments thereof. For example, liposome preparations containing a cell-impermeable macromolecule may promote cell fusion and intracellular delivery of the macromolecule. Alternatively, CYSKP or a fragment thereof may be joined to a short cationic N-terminal portion from the HIV Tat-1 protein. Fusion proteins thus generated have been found to transduce into the cells of all tissues, including the brain, in a mouse model system (Schwarze, S.R. et al. (1999) Science 285:1569-1572).

For any compound, the therapeutically effective dose can be estimated initially either in cell culture assays, e.g., of neoplastic cells, or in animal models such as mice, rats, rabbits, dogs, monkeys, or pigs. An animal model may also be used to determine the appropriate concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans.

A therapeutically effective dose refers to that amount of active ingredient, for example CYSKP or fragments thereof, antibodies of CYSKP, and agonists, antagonists or inhibitors of CYSKP, which ameliorates the symptoms or condition. Therapeutic efficacy and toxicity may be determined by

standard pharmaceutical procedures in cell cultures or with experimental animals, such as by calculating the  $ED_{50}$  (the dose therapeutically effective in 50% of the population) or  $LD_{50}$  (the dose lethal to 50% of the population) statistics. The dose ratio of toxic to therapeutic effects is the therapeutic index, which can be expressed as the  $LD_{50}/ED_{50}$  ratio. Compositions which exhibit large therapeutic indices are preferred. The data obtained from cell culture assays and animal studies are used to formulate a range of dosage for human use. The dosage contained in such compositions is preferably within a range of circulating concentrations that includes the  $ED_{50}$  with little or no toxicity. The dosage varies within this range depending upon the dosage form employed, the sensitivity of the patient, and the route of administration.

The exact dosage will be determined by the practitioner, in light of factors related to the subject requiring treatment. Dosage and administration are adjusted to provide sufficient levels of the active moiety or to maintain the desired effect. Factors which may be taken into account include the severity of the disease state, the general health of the subject, the age, weight, and gender of the subject, time and frequency of administration, drug combination(s), reaction sensitivities, and response to therapy. Long-acting compositions may be administered every 3 to 4 days, every week, or biweekly depending on the half-life and clearance rate of the particular formulation.

Normal dosage amounts may vary from about  $0.1~\mu g$  to  $100,000~\mu g$ , up to a total dose of about 1 gram, depending upon the route of administration. Guidance as to particular dosages and methods of delivery is provided in the literature and generally available to practitioners in the art. Those skilled in the art will employ different formulations for nucleotides than for proteins or their inhibitors. Similarly, delivery of polynucleotides or polypeptides will be specific to particular cells, conditions, locations, etc.

## DIAGNOSTICS

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In another embodiment, antibodies which specifically bind CYSKP may be used for the diagnosis of disorders characterized by expression of CYSKP, or in assays to monitor patients being treated with CYSKP or agonists, antagonists, or inhibitors of CYSKP. Antibodies useful for diagnostic purposes may be prepared in the same manner as described above for therapeutics. Diagnostic assays for CYSKP include methods which utilize the antibody and a label to detect CYSKP in human body fluids or in extracts of cells or tissues. The antibodies may be used with or without modification, and may be labeled by covalent or non-covalent attachment of a reporter molecule. A wide variety of reporter molecules, several of which are described above, are known in the art and may be used.

A variety of protocols for measuring CYSKP, including ELISAs, RIAs, and FACS, are known in the art and provide a basis for diagnosing altered or abnormal levels of CYSKP expression. Normal or standard values for CYSKP expression are established by combining body fluids or cell extracts

taken from normal mammalian subjects, for example, human subjects, with antibodies to CYSKP under conditions suitable for complex formation. The amount of standard complex formation may be quantitated by various methods, such as photometric means. Quantities of CYSKP expressed in subject, control, and disease samples from biopsied tissues are compared with the standard values. Deviation between standard and subject values establishes the parameters for diagnosing disease.

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In another embodiment of the invention, the polynucleotides encoding CYSKP may be used for diagnostic purposes. The polynucleotides which may be used include oligonucleotide sequences, complementary RNA and DNA molecules, and PNAs. The polynucleotides may be used to detect and quantify gene expression in biopsied tissues in which expression of CYSKP may be correlated with disease. The diagnostic assay may be used to determine absence, presence, and excess expression of CYSKP, and to monitor regulation of CYSKP levels during therapeutic intervention.

In one aspect, hybridization with PCR probes which are capable of detecting polynucleotide sequences, including genomic sequences, encoding CYSKP or closely related molecules may be used to identify nucleic acid sequences which encode CYSKP. The specificity of the probe, whether it is made from a highly specific region, e.g., the 5' regulatory region, or from a less specific region, e.g., a conserved motif, and the stringency of the hybridization or amplification will determine whether the probe identifies only naturally occurring sequences encoding CYSKP, allelic variants, or related sequences.

Probes may also be used for the detection of related sequences, and may have at least 50% sequence identity to any of the CYSKP encoding sequences. The hybridization probes of the subject invention may be DNA or RNA and may be derived from the sequence of SEQ ID NO:35-68 or from genomic sequences including promoters, enhancers, and introns of the CYSKP gene.

Means for producing specific hybridization probes for DNAs encoding CYSKP include the cloning of polynucleotide sequences encoding CYSKP or CYSKP derivatives into vectors for the production of mRNA probes. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes <u>in vitro</u> by means of the addition of the appropriate RNA polymerases and the appropriate labeled nucleotides. Hybridization probes may be labeled by a variety of reporter groups, for example, by radionuclides such as <sup>32</sup>P or <sup>35</sup>S, or by enzymatic labels, such as alkaline phosphatase coupled to the probe via avidin/biotin coupling systems, and the like.

Polynucleotide sequences encoding CYSKP may be used for the diagnosis of disorders associated with expression of CYSKP. Examples of such disorders include, but are not limited to, a cell proliferative disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including

adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus; an autoimmune/inflammatory disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory 5 distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasisectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, 10 Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, 15 systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; a vesicle trafficking disorder such as cystic fibrosis, glucose-galactose malabsorption syndrome, hypercholesterolemia, diabetes mellitus, diabetes insipidus, hyper- and hypoglycemia, Grave's disease, goiter, Cushing's disease, and Addison's disease, 20 gastrointestinal disorders including ulcerative colitis, gastric and duodenal ulcers, other conditions associated with abnormal vesicle trafficking, including acquired immunodeficiency syndrome (AIDS), allergies including hay fever, asthma, and urticaria (hives), autoimmune hemolytic anemia, proliferative glomerulonephritis, inflammatory bowel disease, multiple sclerosis, myasthenia gravis, rheumatoid and osteoarthritis, scleroderma, Chediak-Higashi and Sjogren's syndromes, systemic lupus erythematosus, 25 toxic shock syndrome, traumatic tissue damage, and viral, bacterial, fungal, helminthic, and protozoal infections; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and 30 other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal

syndrome, mental retardation and other developmental disorders of the central nervous system including Down syndrome, cerebral palsy, neuroskeletal disorders, autonomic nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis, inherited, metabolic, endocrine, and toxic myopathies, myasthenia gravis, periodic paralysis, mental disorders including mood, anxiety, and schizophrenic disorders, seasonal affective disorder (SAD), akathesia, amnesia, catatonia, diabetic neuropathy, tardive dyskinesia, dystonias, paranoid psychoses, postherpetic neuralgia, Tourette's disorder, progressive supranuclear palsy, corticobasal degeneration, and familial frontotemporal dementia; a cell motility disorder such as ankylosing spondylitis, Chediak-Higashi syndrome, Duchenne and Becker muscular dystrophy, intrahepatic cholestasis, myocardial hyperplasia, cardiomyopathy, early onset peridontitis, cancers such as adenocarcinoma, ovarian carcinoma, and chronic myelogenous leukemia, and bacterial and helminthic infections; a reproductive disorder such as a disorder of prolactin production, infertility, including tubal disease, ovulatory defects, endometriosis, a disruption of the estrous cycle, a disruption of the menstrual cycle, polycystic ovary syndrome, ovarian hyperstimulation syndrome, an endometrial or ovarian tumor, a uterine fibroid, autoimmune disorders, ectopic pregnancy, teratogenesis, cancer of the breast, fibrocystic breast disease, galactorrhea, a disruption of spermatogenesis, abnormal sperm physiology, cancer of the testis, cancer of the prostate, benign prostatic hyperplasia, prostatitis, Peyronie's disease, impotence, carcinoma of the male breast, gynecomastia, hypergonadotropic and hypogonadotropic hypogonadism, pseudohermaphroditism, azoospermia, premature ovarian failure, acrosin deficiency, delayed puperty, retrograde ejaculation and anejaculation, haemangioblastomas, cystsphaeochromocytomas, paraganglioma, cystadenomas of the epididymis, and endolymphatic sac tumours; and a muscle disorder such as myocarditis, Duchenne's muscular dystrophy, Becker's muscular dystrophy, myotonic dystrophy, central core disease, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, infectious myositis, polymyositis, dermatomyositis, inclusion body myositis, thyrotoxic myopathy, and ethanol myopathy, angina, anaphylactic shock, arrhythmias, asthma, cardiovascular shock, Cushing's syndrome, hypertension, hypoglycemia, myocardial infarction, migraine, and pheochromocytoma, and myopathies including cardiomyopathy, encephalopathy, epilepsy, Kearns-Sayre syndrome, lactic acidosis, myoclonic disorder, and ophthalmoplegia. The polynucleotide sequences encoding CYSKP may be used in Southern or northern analysis, dot blot, or other membrane-based technologies; in PCR technologies; in dipstick, pin, and multiformat ELISA-like assays; and in microarrays utilizing fluids or tissues from patients to detect altered CYSKP expression. Such qualitative or quantitative methods are well known in the art.

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In a particular aspect, the nucleotide sequences encoding CYSKP may be useful in assays that

detect the presence of associated disorders, particularly those mentioned above. The nucleotide sequences encoding CYSKP may be labeled by standard methods and added to a fluid or tissue sample from a patient under conditions suitable for the formation of hybridization complexes. After a suitable incubation period, the sample is washed and the signal is quantified and compared with a standard value. If the amount of signal in the patient sample is significantly altered in comparison to a control sample then the presence of altered levels of nucleotide sequences encoding CYSKP in the sample indicates the presence of the associated disorder. Such assays may also be used to evaluate the efficacy of a particular therapeutic treatment regimen in animal studies, in clinical trials, or to monitor the treatment of an individual patient.

In order to provide a basis for the diagnosis of a disorder associated with expression of CYSKP, a normal or standard profile for expression is established. This may be accomplished by combining body fluids or cell extracts taken from normal subjects, either animal or human, with a sequence, or a fragment thereof, encoding CYSKP, under conditions suitable for hybridization or amplification. Standard hybridization may be quantified by comparing the values obtained from normal subjects with values from an experiment in which a known amount of a substantially purified polynucleotide is used. Standard values obtained in this manner may be compared with values obtained from samples from patients who are symptomatic for a disorder. Deviation from standard values is used to establish the presence of a disorder.

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Once the presence of a disorder is established and a treatment protocol is initiated, hybridization assays may be repeated on a regular basis to determine if the level of expression in the patient begins to approximate that which is observed in the normal subject. The results obtained from successive assays may be used to show the efficacy of treatment over a period ranging from several days to months.

With respect to cancer, the presence of an abnormal amount of transcript (either under- or overexpressed) in biopsied tissue from an individual may indicate a predisposition for the development of the disease, or may provide a means for detecting the disease prior to the appearance of actual clinical symptoms. A more definitive diagnosis of this type may allow health professionals to employ preventative measures or aggressive treatment earlier thereby preventing the development or further progression of the cancer.

Additional diagnostic uses for oligonucleotides designed from the sequences encoding CYSKP may involve the use of PCR. These oligomers may be chemically synthesized, generated enzymatically, or produced <u>in vitro</u>. Oligomers will preferably contain a fragment of a polynucleotide encoding CYSKP, or a fragment of a polynucleotide complementary to the polynucleotide encoding CYSKP, and will be employed under optimized conditions for identification of a specific gene or condition.

Oligomers may also be employed under less stringent conditions for detection or quantification of closely related DNA or RNA sequences.

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In a particular aspect, oligonucleotide primers derived from the polynucleotide sequences encoding CYSKP may be used to detect single nucleotide polymorphisms (SNPs). SNPs are substitutions, insertions and deletions that are a frequent cause of inherited or acquired genetic disease in humans. Methods of SNP detection include, but are not limited to, single-stranded conformation polymorphism (SSCP) and fluorescent SSCP (fSSCP) methods. In SSCP, oligonucleotide primers derived from the polynucleotide sequences encoding CYSKP are used to amplify DNA using the polymerase chain reaction (PCR). The DNA may be derived, for example, from diseased or normal tissue, biopsy samples, bodily fluids, and the like. SNPs in the DNA cause differences in the secondary and tertiary structures of PCR products in single-stranded form, and these differences are detectable using gel electrophoresis in non-denaturing gels. In fSCCP, the oligonucleotide primers are fluorescently labeled, which allows detection of the amplimers in high-throughput equipment such as DNA sequencing machines. Additionally, sequence database analysis methods, termed in silico SNP (isSNP), are capable of identifying polymorphisms by comparing the sequence of individual overlapping DNA fragments which assemble into a common consensus sequence. These computerbased methods filter out sequence variations due to laboratory preparation of DNA and sequencing errors using statistical models and automated analyses of DNA sequence chromatograms. In the alternative, SNPs may be detected and characterized by mass spectrometry using, for example, the high throughput MASSARRAY system (Sequenom, Inc., San Diego CA).

Methods which may also be used to quantify the expression of CYSKP include radiolabeling or biotinylating nucleotides, coamplification of a control nucleic acid, and interpolating results from standard curves. (See, e.g., Melby, P.C. et al. (1993) J. Immunol. Methods 159:235-244; Duplaa, C. et al. (1993) Anal. Biochem. 212:229-236.) The speed of quantitation of multiple samples may be accelerated by running the assay in a high-throughput format where the oligomer or polynucleotide of interest is presented in various dilutions and a spectrophotometric or colorimetric response gives rapid quantitation.

In further embodiments, oligonucleotides or longer fragments derived from any of the polynucleotide sequences described herein may be used as elements on a microarray. The microarray can be used in transcript imaging techniques which monitor the relative expression levels of large numbers of genes simultaneously as described below. The microarray may also be used to identify genetic variants, mutations, and polymorphisms. This information may be used to determine gene function, to understand the genetic basis of a disorder, to diagnose a disorder, to monitor progression/regression of disease as a function of gene expression, and to develop and monitor the

activities of therapeutic agents in the treatment of disease. In particular, this information may be used to develop a pharmacogenomic profile of a patient in order to select the most appropriate and effective treatment regimen for that patient. For example, therapeutic agents which are highly effective and display the fewest side effects may be selected for a patient based on his/her pharmacogenomic profile.

In another embodiment, CYSKP, fragments of CYSKP, or antibodies specific for CYSKP may be used as elements on a microarray. The microarray may be used to monitor or measure protein-protein interactions, drug-target interactions, and gene expression profiles, as described above.

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A particular embodiment relates to the use of the polynucleotides of the present invention to generate a transcript image of a tissue or cell type. A transcript image represents the global pattern of gene expression by a particular tissue or cell type. Global gene expression patterns are analyzed by quantifying the number of expressed genes and their relative abundance under given conditions and at a given time. (See Seilhamer et al., "Comparative Gene Transcript Analysis," U.S. Patent Number 5,840,484, expressly incorporated by reference herein.) Thus a transcript image may be generated by hybridizing the polynucleotides of the present invention or their complements to the totality of transcripts or reverse transcripts of a particular tissue or cell type. In one embodiment, the hybridization takes place in high-throughput format, wherein the polynucleotides of the present invention or their complements comprise a subset of a plurality of elements on a microarray. The resultant transcript image would provide a profile of gene activity.

Transcript images may be generated using transcripts isolated from tissues, cell lines, biopsies, or other biological samples. The transcript image may thus reflect gene expression <u>in vivo</u>, as in the case of a tissue or biopsy sample, or <u>in vitro</u>, as in the case of a cell line.

Transcript images which profile the expression of the polynucleotides of the present invention may also be used in conjunction with <u>in vitro</u> model systems and preclinical evaluation of pharmaceuticals, as well as toxicological testing of industrial and naturally-occurring environmental compounds. All compounds induce characteristic gene expression patterns, frequently termed molecular fingerprints or toxicant signatures, which are indicative of mechanisms of action and toxicity (Nuwaysir, E.F. et al. (1999) Mol. Carcinog. 24:153-159; Steiner, S. and N.L. Anderson (2000)

Toxicol. Lett. 112-113:467-471, expressly incorporated by reference herein). If a test compound has a signature similar to that of a compound with known toxicity, it is likely to share those toxic properties. These fingerprints or signatures are most useful and refined when they contain expression information from a large number of genes and gene families. Ideally, a genome-wide measurement of expression provides the highest quality signature. Even genes whose expression is not altered by any tested compounds are important as well, as the levels of expression of these genes are used to normalize the rest of the expression data. The normalization procedure is useful for comparison of expression data

after treatment with different compounds. While the assignment of gene function to elements of a toxicant signature aids in interpretation of toxicity mechanisms, knowledge of gene function is not necessary for the statistical matching of signatures which leads to prediction of toxicity. (See, for example, Press Release 00-02 from the National Institute of Environmental Health Sciences, released February 29, 2000, available at http://www.niehs.nih.gov/oc/news/toxchip.htm.) Therefore, it is important and desirable in toxicological screening using toxicant signatures to include all expressed gene sequences.

In one embodiment, the toxicity of a test compound is assessed by treating a biological sample containing nucleic acids with the test compound. Nucleic acids that are expressed in the treated biological sample are hybridized with one or more probes specific to the polynucleotides of the present invention, so that transcript levels corresponding to the polynucleotides of the present invention may be quantified. The transcript levels in the treated biological sample are compared with levels in an untreated biological sample. Differences in the transcript levels between the two samples are indicative of a toxic response caused by the test compound in the treated sample.

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Another particular embodiment relates to the use of the polypeptide sequences of the present invention to analyze the proteome of a tissue or cell type. The term proteome refers to the global pattern of protein expression in a particular tissue or cell type. Each protein component of a proteome can be subjected individually to further analysis. Proteome expression patterns, or profiles, are analyzed by quantifying the number of expressed proteins and their relative abundance under given conditions and at a given time. A profile of a cell's proteome may thus be generated by separating and analyzing the polypeptides of a particular tissue or cell type. In one embodiment, the separation is achieved using two-dimensional gel electrophoresis, in which proteins from a sample are separated by isoelectric focusing in the first dimension, and then according to molecular weight by sodium dodecyl sulfate slab gel electrophoresis in the second dimension (Steiner and Anderson, supra). The proteins are visualized in the gel as discrete and uniquely positioned spots, typically by staining the gel with an agent such as Coomassie Blue or silver or fluorescent stains. The optical density of each protein spot is generally proportional to the level of the protein in the sample. The optical densities of equivalently positioned protein spots from different samples, for example, from biological samples either treated or untreated with a test compound or therapeutic agent, are compared to identify any changes in protein spot density related to the treatment. The proteins in the spots are partially sequenced using, for example, standard methods employing chemical or enzymatic cleavage followed by mass spectrometry. The identity of the protein in a spot may be determined by comparing its partial sequence, preferably of at least 5 contiguous amino acid residues, to the polypeptide sequences of the present invention. In some cases, further sequence data may be obtained for definitive protein identification.

A proteomic profile may also be generated using antibodies specific for CYSKP to quantify the levels of CYSKP expression. In one embodiment, the antibodies are used as elements on a microarray, and protein expression levels are quantified by exposing the microarray to the sample and detecting the levels of protein bound to each array element (Lucking, A. et al. (1999) Anal. Biochem. 270:103-111; Mendoze, L.G. et al. (1999) Biotechniques 27:778-788). Detection may be performed by a variety of methods known in the art, for example, by reacting the proteins in the sample with a thiol- or aminoreactive fluorescent compound and detecting the amount of fluorescence bound at each array element.

Toxicant signatures at the proteome level are also useful for toxicological screening, and should be analyzed in parallel with toxicant signatures at the transcript level. There is a poor correlation between transcript and protein abundances for some proteins in some tissues (Anderson, N.L. and J. Seilhamer (1997) Electrophoresis 18:533-537), so proteome toxicant signatures may be useful in the analysis of compounds which do not significantly affect the transcript image, but which alter the proteomic profile. In addition, the analysis of transcripts in body fluids is difficult, due to rapid degradation of mRNA, so proteomic profiling may be more reliable and informative in such cases.

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In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins that are expressed in the treated biological sample are separated so that the amount of each protein can be quantified. The amount of each protein is compared to the amount of the corresponding protein in an untreated biological sample. A difference in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample. Individual proteins are identified by sequencing the amino acid residues of the individual proteins and comparing these partial sequences to the polypeptides of the present invention.

In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins from the biological sample are incubated with antibodies specific to the polypeptides of the present invention. The amount of protein recognized by the antibodies is quantified. The amount of protein in the treated biological sample is compared with the amount in an untreated biological sample. A difference in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample.

Microarrays may be prepared, used, and analyzed using methods known in the art. (See, e.g., Brennan, T.M. et al. (1995) U.S. Patent No. 5,474,796; Schena, M. et al. (1996) Proc. Natl. Acad. Sci. USA 93:10614-10619; Baldeschweiler et al. (1995) PCT application WO95/251116; Shalon, D. et al. (1995) PCT application WO95/35505; Heller, R.A. et al. (1997) Proc. Natl. Acad. Sci. USA 94:2150-2155; and Heller, M.J. et al. (1997) U.S. Patent No. 5,605,662.) Various types of microarrays are well known and thoroughly described in <u>DNA Microarrays: A Practical Approach</u>, M. Schena, ed. (1999) Oxford University Press, London, hereby expressly incorporated by reference.

In another embodiment of the invention, nucleic acid sequences encoding CYSKP may be used to generate hybridization probes useful in mapping the naturally occurring genomic sequence. Either coding or noncoding sequences may be used, and in some instances, noncoding sequences may be preferable over coding sequences. For example, conservation of a coding sequence among members of a multi-gene family may potentially cause undesired cross hybridization during chromosomal mapping. The sequences may be mapped to a particular chromosome, to a specific region of a chromosome, or to artificial chromosome constructions, e.g., human artificial chromosomes (HACs), yeast artificial chromosomes (YACs), bacterial artificial chromosomes (BACs), bacterial P1 constructions, or single chromosome cDNA libraries. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355; Price, C.M. (1993) Blood Rev. 7:127-134; and Trask, B.J. (1991) Trends Genet. 7:149-154.) Once mapped, the nucleic acid sequences of the invention may be used to develop genetic linkage maps, for example, which correlate the inheritance of a disease state with the inheritance of a particular chromosome region or restriction fragment length polymorphism (RFLP). (See, for example, Lander, E.S. and D. Botstein (1986) Proc. Natl. Acad. Sci. USA.83:7353-7357.)

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Fluorescent <u>in situ</u> hybridization (FISH) may be correlated with other physical and genetic map data. (See, e.g., Heinz-Ulrich, et al. (1995) in Meyers, <u>supra</u>, pp. 965-968.) Examples of genetic map data can be found in various scientific journals or at the Online Mendelian Inheritance in Man (OMIM) World Wide Web site. Correlation between the location of the gene encoding CYSKP on a physical map and a specific disorder, or a predisposition to a specific disorder, may help define the region of DNA associated with that disorder and thus may further positional cloning efforts.

In situ hybridization of chromosomal preparations and physical mapping techniques, such as linkage analysis using established chromosomal markers, may be used for extending genetic maps. Often the placement of a gene on the chromosome of another mammalian species, such as mouse, may reveal associated markers even if the exact chromosomal locus is not known. This information is valuable to investigators searching for disease genes using positional cloning or other gene discovery techniques. Once the gene or genes responsible for a disease or syndrome have been crudely localized by genetic linkage to a particular genomic region, e.g., ataxia-telangiectasia to 11q22-23, any sequences mapping to that area may represent associated or regulatory genes for further investigation. (See, e.g., Gatti, R.A. et al. (1988) Nature 336:577-580.) The nucleotide sequence of the instant invention may also be used to detect differences in the chromosomal location due to translocation, inversion, etc., among normal, carrier, or affected individuals.

In another embodiment of the invention, CYSKP, its catalytic or immunogenic fragments, or oligopeptides thereof can be used for screening libraries of compounds in any of a variety of drug screening techniques. The fragment employed in such screening may be free in solution, affixed to a

solid support, borne on a cell surface, or located intracellularly. The formation of binding complexes between CYSKP and the agent being tested may be measured.

Another technique for drug screening provides for high throughput screening of compounds having suitable binding affinity to the protein of interest. (See, e.g., Geysen, et al. (1984) PCT application WO84/03564.) In this method, large numbers of different small test compounds are synthesized on a solid substrate. The test compounds are reacted with CYSKP, or fragments thereof, and washed. Bound CYSKP is then detected by methods well known in the art. Purified CYSKP can also be coated directly onto plates for use in the aforementioned drug screening techniques. Alternatively, non-neutralizing antibodies can be used to capture the peptide and immobilize it on a solid support.

In another embodiment, one may use competitive drug screening assays in which neutralizing antibodies capable of binding CYSKP specifically compete with a test compound for binding CYSKP. In this manner, antibodies can be used to detect the presence of any peptide which shares one or more antigenic determinants with CYSKP.

In additional embodiments, the nucleotide sequences which encode CYSKP may be used in any molecular biology techniques that have yet to be developed, provided the new techniques rely on properties of nucleotide sequences that are currently known, including, but not limited to, such properties as the triplet genetic code and specific base pair interactions.

Without further elaboration, it is believed that one skilled in the art can, using the preceding description, utilize the present invention to its fullest extent. The following embodiments are, therefore, to be construed as merely illustrative, and not limitative of the remainder of the disclosure in any way whatsoever.

Without further elaboration, it is believed that one skilled in the art can, using the preceding description, utilize the present invention to its fullest extent. The following preferred specific embodiments are, therefore, to be construed as merely illustrative, and not limitative of the remainder of the disclosure in any way whatsoever.

The disclosures of all patents, applications, and publications mentioned above and below, in particular U.S. Ser. No. 60/201,960, U.S. Ser. No. 60/202,729, U.S. Ser. No. 60/209,705, U.S. Ser. No. 60/210,149, and U.S. Ser. No. 60/213,215, are hereby expressly incorporated by reference.

## **EXAMPLES**

## I. Construction of cDNA Libraries

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Incyte cDNAs were derived from cDNA libraries described in the LIFESEQ GOLD database (Incyte Genomics, Palo Alto CA) and shown in Table 4, column 5. Some tissues were homogenized

and lysed in guanidinium isothiocyanate, while others were homogenized and lysed in phenol or in a suitable mixture of denaturants, such as TRIZOL (Life Technologies), a monophasic solution of phenol and guanidine isothiocyanate. The resulting lysates were centrifuged over CsCl cushions or extracted with chloroform. RNA was precipitated from the lysates with either isopropanol or sodium acetate and ethanol, or by other routine methods.

Phenol extraction and precipitation of RNA were repeated as necessary to increase RNA purity. In some cases, RNA was treated with DNase. For most libraries, poly(A)+ RNA was isolated using oligo d(T)-coupled paramagnetic particles (Promega), OLIGOTEX latex particles (QIAGEN, Chatsworth CA), or an OLIGOTEX mRNA purification kit (QIAGEN). Alternatively, RNA was isolated directly from tissue lysates using other RNA isolation kits, e.g., the POLY(A)PURE mRNA purification kit (Ambion, Austin TX).

In some cases, Stratagene was provided with RNA and constructed the corresponding cDNA libraries. Otherwise, cDNA was synthesized and cDNA libraries were constructed with the UNIZAP vector system (Stratagene) or SUPERSCRIPT plasmid system (Life Technologies), using the recommended procedures or similar methods known in the art. (See, e.g., Ausubel, 1997, supra, units 5.1-6.6.) Reverse transcription was initiated using oligo d(T) or random primers. Synthetic oligonucleotide adapters were ligated to double stranded cDNA, and the cDNA was digested with the appropriate restriction enzyme or enzymes. For most libraries, the cDNA was size-selected (300-1000 bp) using SEPHACRYL S1000, SEPHAROSE CL2B, or SEPHAROSE CL4B column chromatography (Amersham Pharmacia Biotech) or preparative agarose gel electrophoresis. cDNAs were ligated into compatible restriction enzyme sites of the polylinker of a suitable plasmid, e.g., PBLUESCRIPT plasmid (Stratagene), PSPORT1 plasmid (Life Technologies), PCDNA2.1 plasmid (Invitrogen, Carlsbad CA), PBK-CMV plasmid (Stratagene), or pINCY (Incyte Genomics, Palo Alto CA), or derivatives thereof. Recombinant plasmids were transformed into competent E. coli cells including XL1-Blue, XL1-BlueMRF, or SOLR from Stratagene or DH5α, DH10B, or ElectroMAX DH10B from Life Technologies.

#### II. Isolation of cDNA Clones

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Plasmids obtained as described in Example I were recovered from host cells by <u>in vivo</u> excision using the UNIZAP vector system (Stratagene) or by cell lysis. Plasmids were purified using at least one of the following: a Magic or WIZARD Minipreps DNA purification system (Promega); an AGTC Miniprep purification kit (Edge Biosystems, Gaithersburg MD); and QIAWELL 8 Plasmid, QIAWELL 8 Plasmid, QIAWELL 8 Ultra Plasmid purification systems or the R.E.A.L. PREP 96 plasmid purification kit from QIAGEN. Following precipitation, plasmids were resuspended in 0.1 ml of distilled water and stored, with or without lyophilization, at 4°C.

Alternatively, plasmid DNA was amplified from host cell lysates using direct link PCR in a high-throughput format (Rao, V.B. (1994) Anal. Biochem. 216:1-14). Host cell lysis and thermal cycling steps were carried out in a single reaction mixture. Samples were processed and stored in 384-well plates, and the concentration of amplified plasmid DNA was quantified fluorometrically using PICOGREEN dye (Molecular Probes, Eugene OR) and a FLUOROSKAN II fluorescence scanner (Labsystems Oy, Helsinki, Finland).

#### III. Sequencing and Analysis

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Incyte cDNA recovered in plasmids as described in Example II were sequenced as follows. Sequencing reactions were processed using standard methods or high-throughput instrumentation such as the ABI CATALYST 800 (Applied Biosystems) thermal cycler or the PTC-200 thermal cycler (MJ Research) in conjunction with the HYDRA microdispenser (Robbins Scientific) or the MICROLAB 2200 (Hamilton) liquid transfer system. cDNA sequencing reactions were prepared using reagents provided by Amersham Pharmacia Biotech or supplied in ABI sequencing kits such as the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems). Electrophoretic separation of cDNA sequencing reactions and detection of labeled polynucleotides were carried out using the MEGABACE 1000 DNA sequencing system (Molecular Dynamics); the ABI PRISM 373 or 377 sequencing system (Applied Biosystems) in conjunction with standard ABI protocols and base calling software; or other sequence analysis systems known in the art. Reading frames within the cDNA sequences were identified using standard methods (reviewed in Ausubel, 1997, supra, unit 7.7). Some of the cDNA sequences were selected for extension using the techniques disclosed in Example VIII.

The polynucleotide sequences derived from Incyte cDNAs were validated by removing vector, linker, and poly(A) sequences and by masking ambiguous bases, using algorithms and programs based on BLAST, dynamic programming, and dinucleotide nearest neighbor analysis. The Incyte cDNA sequences or translations thereof were then queried against a selection of public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases, and BLOCKS, PRINTS, DOMO, PRODOM, and hidden Markov model (HMM)-based protein family databases such as PFAM. (HMM is a probabilistic approach which analyzes consensus primary structures of gene families. See, for example, Eddy, S.R. (1996) Curr. Opin. Struct. Biol. 6:361-365.) The queries were performed using programs based on BLAST, FASTA, BLIMPS, and HMMER. The Incyte cDNA sequences were assembled to produce full length polynucleotide sequences. Alternatively, GenBank cDNAs, GenBank ESTs, stitched sequences, stretched sequences, or Genscan-predicted coding sequences (see Examples IV and V) were used to extend Incyte cDNA assemblages to full length. Assembly was performed using programs based on Phred, Phrap, and Consed, and cDNA assemblages

were screened for open reading frames using programs based on GeneMark, BLAST, and FASTA. The full length polynucleotide sequences were translated to derive the corresponding full length polypeptide sequences. Alternatively, a polypeptide of the invention may begin at any of the methionine residues of the full length translated polypeptide. Full length polypeptide sequences were subsequently analyzed by querying against databases such as the GenBank protein databases (genpept), SwissProt, BLOCKS, PRINTS, DOMO, PRODOM, Prosite, and hidden Markov model (HMM)-based protein family databases such as PFAM. Full length polynucleotide sequences are also analyzed using MACDNASIS PRO software (Hitachi Software Engineering, South San Francisco CA) and LASERGENE software (DNASTAR). Polynucleotide and polypeptide sequence alignments are generated using default parameters specified by the CLUSTAL algorithm as incorporated into the MEGALIGN multisequence alignment program (DNASTAR), which also calculates the percent identity between aligned sequences.

Table 7 summarizes the tools, programs, and algorithms used for the analysis and assembly of Incyte cDNA and full length sequences and provides applicable descriptions, references, and threshold parameters. The first column of Table 7 shows the tools, programs, and algorithms used, the second column provides brief descriptions thereof, the third column presents appropriate references, all of which are incorporated by reference herein in their entirety, and the fourth column presents, where applicable, the scores, probability values, and other parameters used to evaluate the strength of a match between two sequences (the higher the score or the lower the probability value, the greater the identity between two sequences).

The programs described above for the assembly and analysis of full length polynucleotide and polypeptide sequences were also used to identify polynucleotide sequence fragments from SEQ ID NO:35-68. Fragments from about 20 to about 4000 nucleotides which are useful in hybridization and amplification technologies are described in Table 4, column 4.

#### 25 IV. Identification and Editing of Coding Sequences from Genomic DNA

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Putative cytoskeleton-associated proteins were initially identified by running the Genscan gene identification program against public genomic sequence databases (e.g., gbpri and gbhtg). Genscan is a general-purpose gene identification program which analyzes genomic DNA sequences from a variety of organisms (See Burge, C. and S. Karlin (1997) J. Mol. Biol. 268:78-94, and Burge, C. and S. Karlin (1998) Curr. Opin. Struct. Biol. 8:346-354). The program concatenates predicted exons to form an assembled cDNA sequence extending from a methionine to a stop codon. The output of Genscan is a FASTA database of polynucleotide and polypeptide sequences. The maximum range of sequence for Genscan to analyze at once was set to 30 kb. To determine which of these Genscan predicted cDNA sequences encode cytoskeleton-associated proteins, the encoded polypeptides were analyzed by querying

against PFAM models for cytoskeleton-associated proteins. Potential cytoskeleton-associated proteins were also identified by homology to Incyte cDNA sequences that had been annotated as cytoskeleton-associated proteins. These selected Genscan-predicted sequences were then compared by BLAST analysis to the genpept and gbpri public databases. Where necessary, the Genscan-predicted sequences were then edited by comparison to the top BLAST hit from genpept to correct errors in the sequence predicted by Genscan, such as extra or omitted exons. BLAST analysis was also used to find any Incyte cDNA or public cDNA coverage of the Genscan-predicted sequences, thus providing evidence for transcription. When Incyte cDNA coverage was available, this information was used to correct or confirm the Genscan predicted sequence. Full length polynucleotide sequences were obtained by assembling Genscan-predicted coding sequences with Incyte cDNA sequences and/or public cDNA sequences using the assembly process described in Example III. Alternatively, full length polynucleotide sequences were derived entirely from edited or unedited Genscan-predicted coding sequences.

# V. Assembly of Genomic Sequence Data with cDNA Sequence Data

## "Stitched" Sequences

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Partial cDNA sequences were extended with exons predicted by the Genscan gene identification program described in Example IV. Partial cDNAs assembled as described in Example III were mapped to genomic DNA and parsed into clusters containing related cDNAs and Genscan exon predictions from one or more genomic sequences. Each cluster was analyzed using an algorithm based on graph theory and dynamic programming to integrate cDNA and genomic information, generating possible splice variants that were subsequently confirmed, edited, or extended to create a full length sequence. Sequence intervals in which the entire length of the interval was present on more than one sequence in the cluster were identified, and intervals thus identified were considered to be equivalent by transitivity. For example, if an interval was present on a cDNA and two genomic sequences, then all three intervals were considered to be equivalent. This process allows unrelated but consecutive genomic sequences to be brought together, bridged by cDNA sequence. Intervals thus identified were then "stitched" together by the stitching algorithm in the order that they appear along their parent sequences to generate the longest possible sequence, as well as sequence variants. Linkages between intervals which proceed along one type of parent sequence (cDNA to cDNA or genomic sequence to genomic sequence) were given preference over linkages which change parent type (cDNA to genomic sequence). The resultant stitched sequences were translated and compared by BLAST analysis to the genpept and gbpri public databases. Incorrect exons predicted by Genscan were corrected by comparison to the top BLAST hit from genpept. Sequences were further extended with additional cDNA sequences, or by inspection of genomic DNA, when necessary.

### "Stretched" Sequences

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Partial DNA sequences were extended to full length with an algorithm based on BLAST analysis. First, partial cDNAs assembled as described in Example III were queried against public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases using the BLAST program. The nearest GenBank protein homolog was then compared by BLAST analysis to either Incyte cDNA sequences or GenScan exon predicted sequences described in Example IV. A chimeric protein was generated by using the resultant high-scoring segment pairs (HSPs) to map the translated sequences onto the GenBank protein homolog. Insertions or deletions may occur in the chimeric protein with respect to the original GenBank protein homolog. The GenBank protein homolog, the chimeric protein, or both were used as probes to search for homologous genomic sequences from the public human genome databases. Partial DNA sequences were therefore "stretched" or extended by the addition of homologous genomic sequences. The resultant stretched sequences were examined to determine whether it contained a complete gene.

## VI. Chromosomal Mapping of CYSKP Encoding Polynucleotides

The sequences which were used to assemble SEQ ID NO:35-68 were compared with sequences from the Incyte LIFESEQ database and public domain databases using BLAST and other implementations of the Smith-Waterman algorithm. Sequences from these databases that matched SEQ ID NO:35-68 were assembled into clusters of contiguous and overlapping sequences using assembly algorithms such as Phrap (Table 7). Radiation hybrid and genetic mapping data available from public resources such as the Stanford Human Genome Center (SHGC), Whitehead Institute for Genome Research (WIGR), and Généthon were used to determine if any of the clustered sequences had been previously mapped. Inclusion of a mapped sequence in a cluster resulted in the assignment of all sequences of that cluster, including its particular SEQ ID NO:, to that map location.

Map locations are represented by ranges, or intervals, of human chromosomes. The map position of an interval, in centiMorgans, is measured relative to the terminus of the chromosome's p-arm. (The centiMorgan (cM) is a unit of measurement based on recombination frequencies between chromosomal markers. On average, 1 cM is roughly equivalent to 1 megabase (Mb) of DNA in humans, although this can vary widely due to hot and cold spots of recombination.) The cM distances are based on genetic markers mapped by Généthon which provide boundaries for radiation hybrid markers whose sequences were included in each of the clusters. Human genome maps and other resources available to the public, such as the NCBI "GeneMap'99" World Wide Web site (http://www.ncbi.nlm.nih.gov/genemap/), can be employed to determine if previously identified disease genes map within or in proximity to the intervals indicated above.

In this manner, SEQ ID NO:44 was mapped to chromosome 17 within the interval from

62.90 to 64.20 centiMorgans, SEQ ID NO:49 was mapped to chromosome 14 within the interval from 73.70 to 76.40 centiMorgans, SEQ ID NO:50 was mapped to chromosome 8 within the interval from 25.80 to 40.30 centiMorgans, SEQ ID NO:54 was mapped to chromosome 1 within the interval from 117.6 to 132.4 centiMorgans, SEQ ID NO:64 was mapped to chromosome 4 within the interval from 56.7 to 60.5 centiMorgans, and SEQ ID NO:65 was mapped to chromosome 5 within the interval from 141.40 to 142.60 centiMorgans.

### VII. Analysis of Polynucleotide Expression

Northern analysis is a laboratory technique used to detect the presence of a transcript of a gene and involves the hybridization of a labeled nucleotide sequence to a membrane on which RNAs from a particular cell type or tissue have been bound. (See, e.g., Sambrook, <u>supra</u>, ch. 7; Ausubel (1995) <u>supra</u>, ch. 4 and 16.)

Analogous computer techniques applying BLAST were used to search for identical or related molecules in cDNA databases such as GenBank or LIFESEQ (Incyte Genomics). This analysis is much faster than multiple membrane-based hybridizations. In addition, the sensitivity of the computer search can be modified to determine whether any particular match is categorized as exact or similar. The basis of the search is the product score, which is defined as:

BLAST Score x Percent Identity

5 x minimum {length(Seq. 1), length(Seq. 2)}

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The product score takes into account both the degree of similarity between two sequences and the length of the sequence match. The product score is a normalized value between 0 and 100, and is calculated as follows: the BLAST score is multiplied by the percent nucleotide identity and the product is divided by (5 times the length of the shorter of the two sequences). The BLAST score is calculated by assigning a score of +5 for every base that matches in a high-scoring segment pair (HSP), and -4 for every mismatch. Two sequences may share more than one HSP (separated by gaps). If there is more than one HSP, then the pair with the highest BLAST score is used to calculate the product score. The product score represents a balance between fractional overlap and quality in a BLAST alignment. For example, a product score of 100 is produced only for 100% identity over the entire length of the shorter of the two sequences being compared. A product score of 70 is produced either by 100% identity and 70% overlap at one end, or by 88% identity and 100% overlap at the other. A product score of 50 is produced either by 100% identity and 50% overlap at one end, or 79% identity and 100% overlap.

Alternatively, polynucleotide sequences encoding CYSKP are analyzed with respect to the tissue sources from which they were derived. For example, some full length sequences are assembled,

at least in part, with overlapping Incyte cDNA sequences (see Example III). Each cDNA sequence is derived from a cDNA library constructed from a human tissue. Each human tissue is classified into one of the following organ/tissue categories: cardiovascular system; connective tissue; digestive system; embryonic structures; endocrine system; exocrine glands; genitalia, female; genitalia, male; germ cells; hemic and immune system; liver; musculoskeletal system; nervous system; pancreas; respiratory system; sense organs; skin; stomatognathic system; unclassified/mixed; or urinary tract. The number of libraries in each category is counted and divided by the total number of libraries across all categories. Similarly, each human tissue is classified into one of the following disease/condition categories: cancer, cell line, developmental, inflammation, neurological, trauma, cardiovascular, pooled, and other, and the number of libraries in each category is counted and divided by the total number of libraries across all categories. The resulting percentages reflect the tissue- and disease-specific expression of cDNA encoding CYSKP. cDNA sequences and cDNA library/tissue information are found in the LIFESEQ GOLD database (Incyte Genomics, Palo Alto CA).

### VIII. Extension of CYSKP Encoding Polynucleotides

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Full length polynucleotide sequences were also produced by extension of an appropriate fragment of the full length molecule using oligonucleotide primers designed from this fragment. One primer was synthesized to initiate 5' extension of the known fragment, and the other primer was synthesized to initiate 3' extension of the known fragment. The initial primers were designed using OLIGO 4.06 software (National Biosciences), or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the target sequence at temperatures of about 68 °C to about 72 °C. Any stretch of nucleotides which would result in hairpin structures and primer-primer dimerizations was avoided.

Selected human cDNA libraries were used to extend the sequence. If more than one extension was necessary or desired, additional or nested sets of primers were designed.

High fidelity amplification was obtained by PCR using methods well known in the art. PCR was performed in 96-well plates using the PTC-200 thermal cycler (MJ Research, Inc.). The reaction mix contained DNA template, 200 nmol of each primer, reaction buffer containing Mg<sup>2+</sup>, (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, and 2-mercaptoethanol, Taq DNA polymerase (Amersham Pharmacia Biotech), ELONGASE enzyme (Life Technologies), and Pfu DNA polymerase (Stratagene), with the following parameters for primer pair PCI A and PCI B: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C. In the alternative, the parameters for primer pair T7 and SK+ were as follows: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 57°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C.

The concentration of DNA in each well was determined by dispensing 100  $\mu$ l PICOGREEN quantitation reagent (0.25% (v/v) PICOGREEN; Molecular Probes, Eugene OR) dissolved in 1X TE and 0.5  $\mu$ l of undiluted PCR product into each well of an opaque fluorimeter plate (Corning Costar, Acton MA), allowing the DNA to bind to the reagent. The plate was scanned in a Fluoroskan II (Labsystems Oy, Helsinki, Finland) to measure the fluorescence of the sample and to quantify the concentration of DNA. A 5  $\mu$ l to 10  $\mu$ l aliquot of the reaction mixture was analyzed by electrophoresis on a 1% agarose gel to determine which reactions were successful in extending the sequence.

The extended nucleotides were desalted and concentrated, transferred to 384-well plates, digested with CviJI cholera virus endonuclease (Molecular Biology Research, Madison WI), and sonicated or sheared prior to religation into pUC 18 vector (Amersham Pharmacia Biotech). For shotgun sequencing, the digested nucleotides were separated on low concentration (0.6 to 0.8%) agarose gels, fragments were excised, and agar digested with Agar ACE (Promega). Extended clones were religated using T4 ligase (New England Biolabs, Beverly MA) into pUC 18 vector (Amersham Pharmacia Biotech), treated with Pfu DNA polymerase (Stratagene) to fill-in restriction site overhangs, and transfected into competent <u>E. coli</u> cells. Transformed cells were selected on antibiotic-containing media, and individual colonies were picked and cultured overnight at 37°C in 384-well plates in LB/2x carb liquid media.

The cells were lysed, and DNA was amplified by PCR using Taq DNA polymerase (Amersham Pharmacia Biotech) and Pfu DNA polymerase (Stratagene) with the following parameters: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 72°C, 2 min; Step 5: steps 2, 3, and 4 repeated 29 times; Step 6: 72°C, 5 min; Step 7: storage at 4°C. DNA was quantified by PICOGREEN reagent (Molecular Probes) as described above. Samples with low DNA recoveries were reamplified using the same conditions as described above. Samples were diluted with 20% dimethysulfoxide (1:2, v/v), and sequenced using DYENAMIC energy transfer sequencing primers and the DYENAMIC DIRECT kit (Amersham Pharmacia Biotech) or the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems).

In like manner, full length polynucleotide sequences are verified using the above procedure or are used to obtain 5' regulatory sequences using the above procedure along with oligonucleotides designed for such extension, and an appropriate genomic library.

### IX. Labeling and Use of Individual Hybridization Probes

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Hybridization probes derived from SEQ ID NO:35-68 are employed to screen cDNAs, genomic DNAs, or mRNAs. Although the labeling of oligonucleotides, consisting of about 20 base pairs, is specifically described, essentially the same procedure is used with larger nucleotide fragments. Oligonucleotides are designed using state-of-the-art software such as OLIGO 4.06 software (National

Biosciences) and labeled by combining 50 pmol of each oligomer, 250  $\mu$ Ci of [ $\gamma$ - $^{32}$ P] adenosine triphosphate (Amersham Pharmacia Biotech), and T4 polynucleotide kinase (DuPont NEN, Boston MA). The labeled oligonucleotides are substantially purified using a SEPHADEX G-25 superfine size exclusion dextran bead column (Amersham Pharmacia Biotech). An aliquot containing  $10^7$  counts per minute of the labeled probe is used in a typical membrane-based hybridization analysis of human genomic DNA digested with one of the following endonucleases: Ase I, Bgl II, Eco RI, Pst I, Xba I, or Pvu II (DuPont NEN).

The DNA from each digest is fractionated on a 0.7% agarose gel and transferred to nylon membranes (Nytran Plus, Schleicher & Schuell, Durham NH). Hybridization is carried out for 16 hours at 40°C. To remove nonspecific signals, blots are sequentially washed at room temperature under conditions of up to, for example, 0.1 x saline sodium citrate and 0.5% sodium dodecyl sulfate. Hybridization patterns are visualized using autoradiography or an alternative imaging means and compared.

### X. Microarrays

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The linkage or synthesis of array elements upon a microarray can be achieved utilizing photolithography, piezoelectric printing (ink-jet printing, See, e.g., Baldeschweiler, <u>supra.</u>), mechanical microspotting technologies, and derivatives thereof. The substrate in each of the aforementioned technologies should be uniform and solid with a non-porous surface (Schena (1999), <u>supra</u>). Suggested substrates include silicon, silica, glass slides, glass chips, and silicon wafers. Alternatively, a procedure analogous to a dot or slot blot may also be used to arrange and link elements to the surface of a substrate using thermal, UV, chemical, or mechanical bonding procedures. A typical array may be produced using available methods and machines well known to those of ordinary skill in the art and may contain any appropriate number of elements. (See, e.g., Schena, M. et al. (1995) Science 270:467-470; Shalon, D. et al. (1996) Genome Res. 6:639-645; Marshall, A. and J. Hodgson (1998) Nat. Biotechnol. 16:27-31.)

Full length cDNAs, Expressed Sequence Tags (ESTs), or fragments or oligomers thereof may comprise the elements of the microarray. Fragments or oligomers suitable for hybridization can be selected using software well known in the art such as LASERGENE software (DNASTAR). The array elements are hybridized with polynucleotides in a biological sample. The polynucleotides in the biological sample are conjugated to a fluorescent label or other molecular tag for ease of detection. After hybridization, nonhybridized nucleotides from the biological sample are removed, and a fluorescence scanner is used to detect hybridization at each array element. Alternatively, laser desorbtion and mass spectrometry may be used for detection of hybridization. The degree of complementarity and the relative abundance of each polynucleotide which hybridizes to an element on

the microarray may be assessed. In one embodiment, microarray preparation and usage is described in detail below.

### <u>Tissue or Cell Sample Preparation</u>

Total RNA is isolated from tissue samples using the guanidinium thiocyanate method and poly(A)<sup>+</sup> RNA is purified using the oligo-(dT) cellulose method. Each poly(A)<sup>+</sup> RNA sample is reverse transcribed using MMLV reverse-transcriptase, 0.05 pg/µl oligo-(dT) primer (21mer), 1X first strand buffer, 0.03 units/µl RNase inhibitor, 500 µM dATP, 500 µM dGTP, 500 µM dTTP, 40 µM dCTP, 40 µM dCTP-Cy3 (BDS) or dCTP-Cy5 (Amersham Pharmacia Biotech). The reverse transcription reaction is performed in a 25 ml volume containing 200 ng poly(A)<sup>+</sup> RNA with GEMBRIGHT kits (Incyte). Specific control poly(A)<sup>+</sup> RNAs are synthesized by in vitro transcription from non-coding yeast genomic DNA. After incubation at 37° C for 2 hr, each reaction sample (one with Cy3 and another with Cy5 labeling) is treated with 2.5 ml of 0.5M sodium hydroxide and incubated for 20 minutes at 85° C to the stop the reaction and degrade the RNA. Samples are purified using two successive CHROMA SPIN 30 gel filtration spin columns (CLONTECH Laboratories, Inc. (CLONTECH), Palo Alto CA) and after combining, both reaction samples are ethanol precipitated using 1 ml of glycogen (1 mg/ml), 60 ml sodium acetate, and 300 ml of 100% ethanol. The sample is then dried to completion using a SpeedVAC (Savant Instruments Inc., Holbrook NY) and resuspended in 14 µl 5X SSC/0.2% SDS.

### **Microarray Preparation**

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Sequences of the present invention are used to generate array elements. Each array element is amplified from bacterial cells containing vectors with cloned cDNA inserts. PCR amplification uses primers complementary to the vector sequences flanking the cDNA insert. Array elements are amplified in thirty cycles of PCR from an initial quantity of 1-2 ng to a final quantity greater than 5 µg. Amplified array elements are then purified using SEPHACRYL-400 (Amersham Pharmacia Biotech).

Purified array elements are immobilized on polymer-coated glass slides. Glass microscope slides (Corning) are cleaned by ultrasound in 0.1% SDS and acetone, with extensive distilled water washes between and after treatments. Glass slides are etched in 4% hydrofluoric acid (VWR Scientific Products Corporation (VWR), West Chester PA), washed extensively in distilled water, and coated with 0.05% aminopropyl silane (Sigma) in 95% ethanol. Coated slides are cured in a 110°C oven.

Array elements are applied to the coated glass substrate using a procedure described in US Patent No. 5,807,522, incorporated herein by reference. 1  $\mu$ l of the array element DNA, at an average concentration of 100 ng/ $\mu$ l, is loaded into the open capillary printing element by a high-speed robotic apparatus. The apparatus then deposits about 5 nl of array element sample per slide.

Microarrays are UV-crosslinked using a STRATALINKER UV-crosslinker (Stratagene). Microarrays are washed at room temperature once in 0.2% SDS and three times in distilled water. Non-specific binding sites are blocked by incubation of microarrays in 0.2% casein in phosphate buffered saline (PBS) (Tropix, Inc., Bedford MA) for 30 minutes at 60°C followed by washes in 0.2% SDS and distilled water as before.

### Hybridization

Hybridization reactions contain 9  $\mu$ l of sample mixture consisting of 0.2  $\mu$ g each of Cy3 and Cy5 labeled cDNA synthesis products in 5X SSC, 0.2% SDS hybridization buffer. The sample mixture is heated to 65°C for 5 minutes and is aliquoted onto the microarray surface and covered with an 1.8 cm² coverslip. The arrays are transferred to a waterproof chamber having a cavity just slightly larger than a microscope slide. The chamber is kept at 100% humidity internally by the addition of 140  $\mu$ l of 5X SSC in a corner of the chamber. The chamber containing the arrays is incubated for about 6.5 hours at 60°C. The arrays are washed for 10 min at 45°C in a first wash buffer (1X SSC, 0.1% SDS), three times for 10 minutes each at 45°C in a second wash buffer (0.1X SSC), and dried.

### Detection

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Reporter-labeled hybridization complexes are detected with a microscope equipped with an Innova 70 mixed gas 10 W laser (Coherent, Inc., Santa Clara CA) capable of generating spectral lines at 488 nm for excitation of Cy3 and at 632 nm for excitation of Cy5. The excitation laser light is focused on the array using a 20X microscope objective (Nikon, Inc., Melville NY). The slide containing the array is placed on a computer-controlled X-Y stage on the microscope and raster-scanned past the objective. The 1.8 cm x 1.8 cm array used in the present example is scanned with a resolution of 20 micrometers.

In two separate scans, a mixed gas multiline laser excites the two fluorophores sequentially. Emitted light is split, based on wavelength, into two photomultiplier tube detectors (PMT R1477, Hamamatsu Photonics Systems, Bridgewater NJ) corresponding to the two fluorophores. Appropriate filters positioned between the array and the photomultiplier tubes are used to filter the signals. The emission maxima of the fluorophores used are 565 nm for Cy3 and 650 nm for Cy5. Each array is typically scanned twice, one scan per fluorophore using the appropriate filters at the laser source, although the apparatus is capable of recording the spectra from both fluorophores simultaneously.

The sensitivity of the scans is typically calibrated using the signal intensity generated by a cDNA control species added to the sample mixture at a known concentration. A specific location on the array contains a complementary DNA sequence, allowing the intensity of the signal at that location to be correlated with a weight ratio of hybridizing species of 1:100,000. When two samples from different sources (e.g., representing test and control cells), each labeled with a different fluorophore, are hybridized to a single array for the purpose of identifying genes that are differentially

expressed, the calibration is done by labeling samples of the calibrating cDNA with the two fluorophores and adding identical amounts of each to the hybridization mixture.

The output of the photomultiplier tube is digitized using a 12-bit RTI-835H analog-to-digital (A/D) conversion board (Analog Devices, Inc., Norwood MA) installed in an IBM-compatible PC computer. The digitized data are displayed as an image where the signal intensity is mapped using a linear 20-color transformation to a pseudocolor scale ranging from blue (low signal) to red (high signal). The data is also analyzed quantitatively. Where two different fluorophores are excited and measured simultaneously, the data are first corrected for optical crosstalk (due to overlapping emission spectra) between the fluorophores using each fluorophore's emission spectrum.

A grid is superimposed over the fluorescence signal image such that the signal from each spot is centered in each element of the grid. The fluorescence signal within each element is then integrated to obtain a numerical value corresponding to the average intensity of the signal. The software used for signal analysis is the GEMTOOLS gene expression analysis program (Incyte).

### XI. Complementary Polynucleotides

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Sequences complementary to the CYSKP-encoding sequences, or any parts thereof, are used to detect, decrease, or inhibit expression of naturally occurring CYSKP. Although use of oligonucleotides comprising from about 15 to 30 base pairs is described, essentially the same procedure is used with smaller or with larger sequence fragments. Appropriate oligonucleotides are designed using OLIGO 4.06 software (National Biosciences) and the coding sequence of CYSKP. To inhibit transcription, a complementary oligonucleotide is designed from the most unique 5' sequence and used to prevent promoter binding to the coding sequence. To inhibit translation, a complementary oligonucleotide is designed to prevent ribosomal binding to the CYSKP-encoding transcript.

### XII. Expression of CYSKP

Expression and purification of CYSKP is achieved using bacterial or virus-based expression systems. For expression of CYSKP in bacteria, cDNA is subcloned into an appropriate vector containing an antibiotic resistance gene and an inducible promoter that directs high levels of cDNA transcription. Examples of such promoters include, but are not limited to, the *trp-lac* (*tac*) hybrid promoter and the T5 or T7 bacteriophage promoter in conjunction with the *lac* operator regulatory element. Recombinant vectors are transformed into suitable bacterial hosts, e.g., BL21(DE3). Antibiotic resistant bacteria express CYSKP upon induction with isopropyl beta-D-thiogalactopyranoside (IPTG). Expression of CYSKP in eukaryotic cells is achieved by infecting insect or mammalian cell lines with recombinant <u>Autographica californica</u> nuclear polyhedrosis virus

(AcMNPV), commonly known as baculovirus. The nonessential polyhedrin gene of baculovirus is replaced with cDNA encoding CYSKP by either homologous recombination or bacterial-mediated

transposition involving transfer plasmid intermediates. Viral infectivity is maintained and the strong polyhedrin promoter drives high levels of cDNA transcription. Recombinant baculovirus is used to infect Spodoptera frugiperda (Sf9) insect cells in most cases, or human hepatocytes, in some cases. Infection of the latter requires additional genetic modifications to baculovirus. (See Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945.)

In most expression systems, CYSKP is synthesized as a fusion protein with, e.g., glutathione Stransferase (GST) or a peptide epitope tag, such as FLAG or 6-His, permitting rapid, single-step, affinity-based purification of recombinant fusion protein from crude cell lysates. GST, a 26-kilodalton enzyme from Schistosoma japonicum, enables the purification of fusion proteins on immobilized glutathione under conditions that maintain protein activity and antigenicity (Amersham Pharmacia Biotech). Following purification, the GST moiety can be proteolytically cleaved from CYSKP at specifically engineered sites. FLAG, an 8-amino acid peptide, enables immunoaffinity purification using commercially available monoclonal and polyclonal anti-FLAG antibodies (Eastman Kodak). 6-His, a stretch of six consecutive histidine residues, enables purification on metal-chelate resins (QIAGEN). Methods for protein expression and purification are discussed in Ausubel (1995, supra, ch. 10 and 16). Purified CYSKP obtained by these methods can be used directly in the assays shown in Examples XVI and XVII, where applicable.

### XIII. Functional Assays

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CYSKP function is assessed by expressing the sequences encoding CYSKP at physiologically elevated levels in mammalian cell culture systems. cDNA is subcloned into a mammalian expression vector containing a strong promoter that drives high levels of cDNA expression. Vectors of choice include PCMV SPORT (Life Technologies) and PCR3.1 (Invitrogen, Carlsbad CA), both of which contain the cytomegalovirus promoter. 5-10  $\mu$ g of recombinant vector are transiently transfected into a human cell line, for example, an endothelial or hematopoietic cell line, using either liposome formulations or electroporation. 1-2  $\mu$ g of an additional plasmid containing sequences encoding a marker protein are co-transfected. Expression of a marker protein provides a means to distinguish transfected cells from nontransfected cells and is a reliable predictor of cDNA expression from the recombinant vector. Marker proteins of choice include, e.g., Green Fluorescent Protein (GFP; Clontech), CD64, or a CD64-GFP fusion protein. Flow cytometry (FCM), an automated, laser optics-based technique, is used to identify transfected cells expressing GFP or CD64-GFP and to evaluate the apoptotic state of the cells and other cellular properties. FCM detects and quantifies the uptake of fluorescent molecules that diagnose events preceding or coincident with cell death. These events include changes in nuclear DNA content as measured by staining of DNA with propidium iodide; changes in

cell size and granularity as measured by forward light scatter and 90 degree side light scatter; down-regulation of DNA synthesis as measured by decrease in bromodeoxyuridine uptake; alterations in expression of cell surface and intracellular proteins as measured by reactivity with specific antibodies; and alterations in plasma membrane composition as measured by the binding of fluorescein-conjugated Annexin V protein to the cell surface. Methods in flow cytometry are discussed in Ormerod, M.G. (1994) Flow Cytometry, Oxford, New York NY.

The influence of CYSKP on gene expression can be assessed using highly purified populations of cells transfected with sequences encoding CYSKP and either CD64 or CD64-GFP. CD64 and CD64-GFP are expressed on the surface of transfected cells and bind to conserved regions of human immunoglobulin G (IgG). Transfected cells are efficiently separated from nontransfected cells using magnetic beads coated with either human IgG or antibody against CD64 (DYNAL, Lake Success NY). mRNA can be purified from the cells using methods well known by those of skill in the art. Expression of mRNA encoding CYSKP and other genes of interest can be analyzed by northern analysis or microarray techniques.

### 15 XIV. Production of CYSKP Specific Antibodies

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CYSKP substantially purified using polyacrylamide gel electrophoresis (PAGE; see, e.g., Harrington, M.G. (1990) Methods Enzymol. 182:488-495), or other purification techniques, is used to immunize rabbits and to produce antibodies using standard protocols.

Alternatively, the CYSKP amino acid sequence is analyzed using LASERGENE software (DNASTAR) to determine regions of high immunogenicity, and a corresponding oligopeptide is synthesized and used to raise antibodies by means known to those of skill in the art. Methods for selection of appropriate epitopes, such as those near the C-terminus or in hydrophilic regions are well described in the art. (See, e.g., Ausubel, 1995, <u>supra</u>, ch. 11.)

Typically, oligopeptides of about 15 residues in length are synthesized using an ABI 431A peptide synthesizer (Applied Biosystems) using FMOC chemistry and coupled to KLH (Sigma-Aldrich, St. Louis MO) by reaction with N-maleimidobenzoyl-N-hydroxysuccinimide ester (MBS) to increase immunogenicity. (See, e.g., Ausubel, 1995, <a href="mailto:supra.">supra.</a>) Rabbits are immunized with the oligopeptide-KLH complex in complete Freund's adjuvant. Resulting antisera are tested for antipeptide and anti-CYSKP activity by, for example, binding the peptide or CYSKP to a substrate, blocking with 1% BSA, reacting with rabbit antisera, washing, and reacting with radio-iodinated goat anti-rabbit IgG.

### XV. Purification of Naturally Occurring CYSKP Using Specific Antibodies

Naturally occurring or recombinant CYSKP is substantially purified by immunoaffinity chromatography using antibodies specific for CYSKP. An immunoaffinity column is constructed by covalently coupling anti-CYSKP antibody to an activated chromatographic resin, such as

CNBr-activated SEPHAROSE (Amersham Pharmacia Biotech). After the coupling, the resin is blocked and washed according to the manufacturer's instructions.

Media containing CYSKP are passed over the immunoaffinity column, and the column is washed under conditions that allow the preferential absorbance of CYSKP (e.g., high ionic strength buffers in the presence of detergent). The column is eluted under conditions that disrupt antibody/CYSKP binding (e.g., a buffer of pH 2 to pH 3, or a high concentration of a chaotrope, such as urea or thiocyanate ion), and CYSKP is collected.

### XVI. Identification of Molecules Which Interact with CYSKP

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CYSKP, or biologically active fragments thereof, are labeled with <sup>125</sup>I Bolton-Hunter reagent. (See, e.g., Bolton A.E. and W.M. Hunter (1973) Biochem. J. 133:529-539.) Candidate molecules previously arrayed in the wells of a multi-well plate are incubated with the labeled CYSKP, washed, and any wells with labeled CYSKP complex are assayed. Data obtained using different concentrations of CYSKP are used to calculate values for the number, affinity, and association of CYSKP with the candidate molecules.

Alternatively, molecules interacting with CYSKP are analyzed using the yeast two-hybrid system as described in Fields, S. and O. Song (1989) Nature 340:245-246, or using commercially available kits based on the two-hybrid system, such as the MATCHMAKER system (Clontech).

CYSKP may also be used in the PATHCALLING process (CuraGen Corp., New Haven CT) which employs the yeast two-hybrid system in a high-throughput manner to determine all interactions between the proteins encoded by two large libraries of genes (Nandabalan, K. et al. (2000) U.S. Patent No. 6,057,101).

### XVII. Demonstration of CYSKP Activity

A microtubule motility assay for CYSKP measures motor protein activity. In this assay, recombinant CYSKP is immobilized onto a glass slide or similar substrate. Taxol-stabilized bovine brain microtubules (commercially available) in a solution containing ATP and cytosolic extract are perfused onto the slide. Movement of microtubules as driven by CYSKP motor activity can be visualized and quantified using video-enhanced light microscopy and image analysis techniques. CYSKP activity is directly proportional to the frequency and velocity of microtubule movement.

Alternatively, an assay for CYSKP measures the formation of protein filaments <u>in vitro</u>. A solution of CYSKP at a concentration greater than the "critical concentration" for polymer assembly is applied to carbon-coated grids. Appropriate nucleation sites may be supplied in the solution. The grids are negative stained with 0.7% (w/v) aqueous uranyl acetate and examined by electron microscopy. The appearance of filaments of approximately 25 nm (microtubules), 8 nm (actin), or 10 nm (intermediate filaments) is a demonstration of protein activity.

Alternatively, an assay for CYSKP measures the binding affinity of CYSKP for actin as described by Hammell, R.L. and Hitchcock-DeGregori, S.E. (1997, J. Biol. Chem. 272:22409-22416). CYSKP and actin are prepared from in vitro recombinant cDNA expression systems and the N-terminus of CYSKP is acetylated using methods well known in the art. Binding of N-terminal acetyl-CYSKP to actin is measured by cosedimentation at 25 °C in a Beckman model TL-100 centrifuge as described. The bound and free CYSKP are determined by quantitative densitometry of SDS-polyacrylamide gels stained with Coomassie Blue. Apparent binding constants (K<sub>app</sub>) and Hill coefficients (H) are determined by using methods well known in the art to fit the data to the equation as described by Hammell and Hitchcock-DeGregori (1997, supra). The CYSKP:actin ratio, determined using densitometry, is normalized. Hammell and Hitchcock-DeGregori (1997, supra) have shown that saturation of binding corresponds to a CYSKP:actin molar ratio of 0.14, a stoichiometry of 1 CYSKP:7 actin. The binding of CYSKP to actin is proportional to the CYSKP activity.

Alternatively, CYSKP activity is measured as ability to bind to microtubules. Microtubules are purified from adult rat brain by reversible assembly (Vallee, R. B. (1982) Methods Enzymol. 134:89-104) or the taxol method (Vallee, R. B. (1982) J. Cell Biol. 92:435-442) using PEM buffer (100 mM PIPES, pH 6.6, 1mM EGTA, 1mM MgSO<sub>4</sub>). To separate the MAPs from tubulin, the pellets from twice-cycled microtubules are resuspended in PEM buffer and applied to a 0.1 M MgSO<sub>4</sub>-saturated phosphocellulose column as described by Sloboda, R. D. and Rosenbaum, J. L. ((1982) Methods Enzymol. 85:409-416). The fractions containing protein are applied to a second phosphocellulose column. In a total volume of 100 ml, 20 ml of CYSKP (250 mg/ml) is added to 80 ml of whole microtubules (450 mg/ml) or tubulin (300 mg/ml) and incubated at 37 °C for 10 minutes in the presence of 1 mM GTP and 50 mM taxol. The suspension is centrifuged, the supernatant is removed, and the microtubule pellet is resuspended to the original reaction volume in PEM buffer. To assess the partitioning of CYSKP between the supernatant and pellet fractions, equal amounts of supernatant and resuspended pellet are placed in SDS sample buffer and assayed on a 5-20% gradient SDS polyacrylamide gel stained with Coomassie Brilliant Blue. The amount of CYSKP in the pellet fraction is proportional to the binding of CYSKP to microtubules.

Alternatively, CYSKP activity is associated with its ability to form protein-protein complexes and is measured by its ability to regulate growth characteristics of NIH3T3 mouse fibroblast cells. A cDNA encoding CYSKP is subcloned into an appropriate eukaryotic expression vector. This vector is transfected into NIH3T3 cells using methods known in the art. Transfected cells are compared with non-transfected cells for the following quantifiable properties: growth in culture to high density, reduced attachment of cells to the substrate, altered cell morphology, and ability to induce tumors when injected into immunodeficient mice. The activity of CYSKP is proportional to the extent of increased growth or

frequency of altered cell morphology in NIH3T3 cells transfected with CYSKP.

claims.

Various modifications and variations of the described methods and systems of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention.

Although the invention has been described in connection with certain embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments.

Indeed, various modifications of the described modes for carrying out the invention which are obvious

to those skilled in molecular biology or related fields are intended to be within the scope of the following

### Table 1

		_	_	_					_	_	_	-	-	-	_	_	-		_	=		-		_	_	=		_	$\overline{}$	_	_	-	_	
II .	Polynucleotide LD 1889577CB1	2427982CB1	2470833CB1	2080579CB1	2156553CB1	2182855CB1	2242106CB1	2726877CB1	2738233CB1	1833116CB1	001799CB1	119814CB1	1295420CB1	1309364CB1	1315267CB1	1403289CB1	1607607CB1	1660025CB1	1796836CB1	2880670CB1	2913976CB1	3092084CB1	3882482CB1	4933451CB1	5043904CB1	5202390CB1	5526375CB1	5677408ĆB1	5982278CB1	6437362CB1	4173970CB1	2772751CB1	2793768CB1	3035248CB1
	SEQ ID NO:	36	37	38	39	40	41	42	43	<b>7</b> ተ	45	46	47	48	49	50	51	52	53	54	55	26	57	58	59	90	61	62	63	79	65	99	67	68
li .	Polypeptide ID	2427982CD1	2470833CD1	2080579CD1	2156553CD1	2182855CD1	2242106CD1	2726877CD1	2738233CD1	1833116CD1	001799CD1	119814CD1	1295420CD1	1309364CD1	1315267CD1	1403289CD1	1607607CD1	1660025CD1	1796836CD1	2880670CD1	2913976CD1	3092084CD1	3882482CD1	4933451CD1	5043904CD1	5202390CD1	5526375CD1	5677408CD1	5982278CD1	6437362CD1	4173970CD1	2772751CD1	2793768CD1	3035248CD1
ded	SEQ ID NO:	2	3	4	5	9	7	8	6	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34
II W	Project ID	2427982	2470833	2080579	2156553	2182855	2242106	2726877	2738233	1833116	001799	119814	1295420	1309364	1315267	1403289	1607607	1660025	1796836	2880670	2913976	3092084	3882482	4933451	5043904	5202390	5526375	5677408	5982278	6437362	4173970	2772751	2793768	3035248

Table 2

SEQ ID NO:		1 500	1 + L L	20x10x1x 110xx01 xx
	Lucyte Dolymentide	Genbank Th MO:	Frobabilicy	GENBANK HOMOLOG
	ID		0	
	1889577CD1	g3347848	0.00E+00	kinesin light chain 2 [Mus musculus]
2	2427982CD1	g2760161	3.00E-64	outer arm dynein light chain 2 [Anthocidaris crassispina]
3	2470833CD1	g11094032	1.00E-147	ls]
		g11036542	0	[Homo sapiens] (AF237772) gamma-parvin
7	2080579CD1	g6141549	2.50E-101	JNK/SAPK-associated protein-1 (JIP-1)
				in [Mus musc
1	1 1 2 2	1	1000	<u>. ا</u>
22	2156553CD1	g5419859	2.00E~170	hypothetical protein similar to tubulin-
				tyrosine ligase [Homo sapiens] (Lafanechere, L. et al. (1998) J. Cell Sci. 11:171-181)
9	2182855CD1	g2276319	0	axonemal dynein heavy chain [Homo sapiens]
7	2242106CD1	93834443	2,00E-13	[Drosophila melanogaster] cytoplasmic dynein
				intermediate chain isororm DiC5b
		g18156	1.20E-10	70kD dynein intermediate chain [chl.mm:domnng noimhandtii]
				s reinnarduil
∞	2726877CD1	g4778	1.30E-12	٠ ــــــــــــــــــــــــــــــــــــ
				1 1 1 1 1 1
6	2738233CD1	g4185884	7.70E-33	o) [Drosophila melanogaste
				(Strumpf, D. and T. Volk (1998) J. Cell Biol.
				143:1259-1270)
		g10880797	0	[Mus musculus] Syne-1A
10	1833116CD1	g12082089	0	[Homo sapiens] hARPX
		g12082091	0	[Gallus gallus] gARPX
11	1799CD1	g3283070	1.70E-07	p80 katanin [Xenopus laevis] (McNally, F.J.,
				. Blol. Cell
		g3005599	5.00E-09	[Homo sapiens] (AF052432) katanin p80
				رب
12	119814CD1	g3243131	4.40E-18	titin [Drosophila melanogaster] (Machado, C.
			100	(1330) U. CELL DIUL.
		95870837	1.00E-113	- 1
13	1295420CD1	g180622	5.60E-37	nic linker pr
				(7867)
14	1309364CD1	g12667401	0	[Homo sapiens] NUF2R
•		g12667403	0	[Mus musculus] NUF2R

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Polvpeptide	Incvte	GenBank	Probability	GenBank Homolog
SEQ ID NO:	Polypeptide   ID	ID NO:	score	
15	1315267CD1	g53996	8.00E-74	Tcp-10 (transmission control protein) [Mus musculus] (Davies, P. et al. (1991) Mamm. Genome 1:235-241)
16	1403289CD1	g5733814	4.60E-196	angiotensin II AT2 receptor-interacting protein (Bedecs, K. et al. (1997) Biochem. J. 325:449-454)
17	1607607CD1	g3158498	1.60E-19	Contains similarity to Pfam domain: PF00628 (PHD finger) (Aasland, R. et al. (1995) Trends Biochem. Sci. 20:56-59)
18	1660025CD1	g3253105	9.80E-20	[Caenorhabditis elegans] strong similarity to the SNF2/RAD54 family of helicases (Eisen, J. et al. (1995) Nucleic Acids Res. 23:2715- 2723)
19	1796836CD1	g414111	7.20E-14	class II INCENP protein (inner centromere protein) [Gallus gallus] (Mackay, A. et al. (1993) J. Cell Biol. 123:373-385)
20	2880670CD1	g1813638	6.90E-16	PF20 [Chlamydomonas reinhardtii] (Smith, E. and P. Lefebvre (1997) Mol. Biol. Cell 8:455-467)
21	2913976CD1	963898	3.10E-56	Zyxin [Gallus gallus] (Sadler, I. et al. (1992) J. Cell Biol. 119:1573-1587)
22	3092084CD1	g1154645	2.30E-10	head-elevated expression in 0.9 kb [Drosophila melanogaster] (Yang, M.Y. et al. (2000) Genetics 154:285-297)
23	3882482CD1	g5825592	7.60E-171	katanin p60 [Xenopus laevis]
24	4933451CD1	g684936	3.20E-30	peptide with resemblance to the actin family [Homo sapiens]
25	5043904CD1	g2832237	2.10E-06	cep250 centrosome associated protein [Homo sapiens] (Mack, G.J. et al. (1998) Arthritis Rheum. 41:551-558)
26	5202390CD1	g6572155	2.90E-21	[Homo sapiens] dJ1014D13.2 (novel protein similar to ACTN3 (actinin, alpha 3))
27	5526375CD1	g2443272	2.80E-77	motor domain of KIF12 [Mus musculus] (Nakagawa, T. et al. (1997) Proc. Natl. Acad. Sci. USA 94:9654-9659)
28	5677408CD1	g6651427	2.20E-05	dynein light intermediate chain 1 (LIC-2) [Rattus norvegicus] (Hughes, S.M. et al. (1995) J. Cell Sci. 108:17-24)
29	5982278CD1	g6006743 g6723675	0	mitotic kinesin-like protein 1 [Danio rerio] [Homo sapiens] mitotic kinase-like protein-1

Table 3

SEQ		Amino	Potential		Signature Sequences, Domains and Motifs	()
NO:	Folypeptiae ID	Acid Residues	Sit	Glycosylation Sites		Methods and Databases
9	2182855CD1	1190	7197 T35		PROTEIN DYNEIN CHAIN MOTOR MICROTUBULES ATPBINDING HEPTAD REPEAT PATTERN HEAVY PD003982:S920-V1190	BLAST-PRODOM
			S68 S97	N777	do DYNEIN; HEAVY; CILIARY; CYTOSOLIC DM04585 P39057 2948-4465:L14-V1190	BLAST-DOMO
-			Y811 Y955 T45 T98 S149 T163 T233 T350 S406 T446 S468 S524 S896 S976		G_Beta_Repeats:L130-N144	MOTIFS
7	2242106CD1	270	356 S		signal_cleavage:M1-T25	SPScan
			S189 S203 T240		omain:V29-L53	HMMER
			S77 T213		WD domain, G-beta repeat WD40:A116-S155, T207- <u>0</u> 245	HMMER-PFAM
<u>&amp;</u>	2726877CD1	647	r173 S44	N99 N120 N316 N480 N508	PROTEIN COILED COIL CHAIN MYOSIN REPEAT HEAVY ATPBINDING FILAMENT HEPTAD	BLAST-PRODOM
			S510	N644		
			T189 T408 S447 S461 S472 S510 S579 S593			
<u>o</u> _	2738233CD1	1086	S12 S32	N182 N359	Spectrin repeat:R2-E66, N69-E171, V174-	HMMER-PFAM
			T343 T404	) #	NZ30-11374, GJ97-NJ01, 1039 )836	
			E S		Spectrin repeat proteins PF00435:W155-K170	BLIMPS-PFAM
			HE			
			ı Ω			
			Ω į,			.,
			, W			
			1404 S512 S556 S559 S658 S675 S977 T987			
10	1833116CD1	396	S108 T341	N21 N101	P59-K245, Q266-D395 Actin	HMMER_PFAM
			S367 T34		T3-L37, Q73-A127, R137-R191, I289-T343, D346-D395 Actins	BLIMPS_BLOCKS
			T243		93 Actins signatures	PROFILESCAN
					AND ACTIN-RELATED PROTEINS	BLAST DOMO
					Q266-K393 ACTIN	BLAST_PRODOM

SEQ	Incyte	Amino	Potential	Potential	Signature Sequences, Domains and Motifs	()
TO NO:	Polypeptide ID	Acid Residue	Phosphorylation Sites	Glycosylation Sites		Methods and Databases
11	001799CD1	1	Y121 Y183 T33	N31 N52 N124	W172-D189 Aldehyde dehydrogenases motif	BLIMPS_BLOCKS
			S43 T58 S137		9 P_value 5.9e-07	BLAST_PRODOM
			S254 S30 S89 S176 S131 S229 S255		KATANIN P80 centrosome-binding subunit	
12	119814CD1	201	T139 Y24 S4 S60	148	M1-A23 signal_cleavage	SPSCAN
			S68 T96 T106 T144		G66-T125, S10-A28 Immunoglobulin domain	HMMER_PFAM
13	1295420CD1	547	T399T2 T58 T77		G314-F345 G436-F467 Cap_Gly	MOTIFS
			T192 T260 S491	N190	G314-S356, G436-P478 CAP-Gly domain	HMMER_PFAM
			T518 T154 T309		T117-R158, T160-S191, N197-R229 Ank repeat	HMMER_PFAM
		****	T374 T377 T386			BLIMPS_BLOCKS
			T454 T515 T518		G436-F476 MICROTUBULES CYTOSKELETON COILED COIL	BLAST_PRODOM
					E417-P492, L294-K362 CAP(cytoskeleton- associated protein)-GLY DOMAIN	BLAST_DOMO
14	1309364CD1	464	Y369 Y445 S118	N30 N215	Q177-K418 COILED COIL MYOSIN REPEAT	BLAST_PRODOM
		·	S232 S239 S340			
	-		T24 T32 T90			
			T137 S147 S232 T372 T428			
15	1315267CD1	569	S3 T68 S85 S103	46 N121 N155	Muscarinic M4 receptor s	BLIMPS PFAM
			T229 S306 S356 T408 T482 T535	N304 N406	H386-A524 TCOMPLEX Male germ-cell specific protein	BLAST_PRODOM
			S551 T246 S20			,
			179 131 1140 9167 4917 9392	***		
ا بر			T318 S385 T450 T477			
16	1403289CD1	436	S4 T17 S111	N80 N336	ıı	MOTIFS
			T167 T212 S222	•	COILED COIL MYOSIN REPEA	BLAST_PRODOM
			S421 S434 T35		QIU/-E395 TRICHUHYALIN (nair root sneatn protein)	BLAST_DOMO
			S390 T58 T76			
			T97 T139 T153			
		m-	T187 S213 S220			
			Y74			

				Taul	radio 3 (volit.)	
SEQ	Incyte		Potential	Potential	Signature Sequences, Domains and Motifs	Analytical
<u>A</u>	Polypeptide		Phosphorylation Glycosylation	Glycosylation		Methods and
NO:	ΩI			Sites		Databases
17	1607607CD1	363	S106 T206 T275	N229 N307	L6-L27 L55-L76 Leucine_Zipper	MOTIFS
			S324		Prepro orexin s	BLIMPS_PRINTS
			T341 S51 T63		S3-E199 F33E11.3 PROTEIN similar to PHD	BLAST_PRODOM
-			S212		finger	
118	1660025CD1	247	S366 S45 S69		K159-K165 Regulator of G protein signaling	BLIMPS_PFAM
			3139 8			
*****************			S161 S183 S238		092-R103 5-hydroxytriptamine 2C receptor	BLIMPS_PRINTS
			T392			
			\$224		E60-P247 TOPOISOMERASE I DNA ISOMERASE	BLAST_PRODOM
			T369 S381		REPEAT	
					A17-S246 CYLICIN II sperm head cytoskeletal	BLAST_DOMO
					protein	
13	1796836CD1	441	S45	N113 N128	Q133-K383 COILED COIL MYOSIN REPEAT	BLAST PRODOM
			3139 8		Q135-Q412 TRICHOHYALIN (hair root sheath	BLAST_DOMO
			S183		protein)	
			T392			
			Y399 S224 T369 S381			
20	2880670CD1	183	T48 S53 S68 T88 N66	N66	[A	BLAST-PRODOM
21	2913976CD1	212	S124 S143 T49		LIM domains: C22-E80; C82-A139; C142-A208	HMMER-PFAM
			¥75		LIM domain BL00478: Y43-L57	BLIMPS-BLOCKS
					LIM domain signatures:	PROFILESCAN
					E3-Y75; Y63-R206; M1-K137	
					LIM METAL-BINDING REPEAT	BLAST-DOMO
					DM00055   Q04584   464-533: F134-H203	
					LIM domain motifs:	MOTIFS
					C142-L1	
22	3092084CD1	227		N19	<u>러</u> 1	BLAST~DOMO
			5221 50 T201 Y141		T/-NLS/(P-Value = 7.0e-08)	

				·	LUIN J (VOIII)	
SEQ	Incyte	Amino		Potential	Signature Sequences, Domains and Motifs	11 0
<u>a</u>	Polypeptide	Acid	Phosphorylation	Glycosylation		Methods and
 Oal		Kesidnes	Sites	Sites	A STATE OF THE PROPERTY OF THE	Databases
23	3882482CD1	490	318 S42 S71	N271	1: G243-R433	HMMER-PFAM
			S87 S143 T150 S174 T318 S416		ily protease signatures 227: W241-A262: S274-R316	BLIMPS-BLOCKS
			440 S448		G338-K384; G414-R433	
			473 S75		KATANIN P60 SÚBUNIT PD116869: M1-P135	BLAST-PRODOM
,			252 T2		IN FAMILY D	BLAST-DOMO
			314 S32:			
			Y3//		AAA motif: V352-R370	MOTIFS
					ATP/GTP binding site (P-loop): G248-T255	MOTIFS
24	4933451CD1	133	S52 S115		Actin domain: M1-M114	HMMER-PFAM
					Actins proteins signature BL00406: T5-K39	BLIMPS-BLOCKS
					Actin signature PR00190: E24-V33	BLIMPS-PRINTS
		_	-			BLAST-PRODOM
	-				ACETYLATION MUSCLE CYTOSKELETON CYTOPLASMIC	
					ACTINLIKE PD000056: V6-L117	
					ACTINS AND ACTIN-RELATED PROTEINS	BLAST-DOMO
				- 1	DM00167 P20360 3-272: A2-M114	
(25	5043904CD1	912	162 S592 S773	N483 N742	PROTEIN COILED COIL CHAIN MYOSIN REPEAT	BLAST-PRODOM
			S54 S6		SINDING FILA	
-			135 S2		(P-value	
_			278 T2		C2-DOMAIN DM00150 P24506 157-283:	BLAST-DOMO
-,			383 85		R460-S584 (P-value = 7.2e-06)	
			620 T6			
			690 S74			
			823 288			
_			908 \$7			
			157 T19			
			243 T308			•
			334 T542			
		_	576 S584			
			671 S752			<del>***</del>
			774			
			862			****

			-		- 1	
SEQ	Incyte	Amino	Potentia i	Potential	Signature Sequences, Domains and Motits	u
i N	Folypeptide ID	Acıa Residues	rno Sit	Glycosylation Sites		Methods and Databases
26	5202390CD1	1076	S602 S903 T10		Calponin homology (CH) domain: P288-S393	HMMER-PFAM
			T176 S204	N312 N405	TININ ACTIN-BIN	BLAST-DOMO
·—			5 8393 8422		DM00325 P18091 28-252: A290-L385	
			2 S494			
			3 S575			
			S632			
			2 S748			-
			5 \$822			
			T954			
			S57 S1			
			1 T185			
			5 5253			
			S406			
			5548			
-			5 S647			
			7856			
			T898 T950 Y149			***
27	5526375CD1	542	3 8313	N188 N292	Kinesin motor domain: R31-N394	HMMER-PFAM
			S162 S		1 BL00411:	BLIMPS-BLOCKS
			3 T158		P25-E39; G100-G121; V157-L175 G216-L240;	
			S218 S231 S272			
			5 T366		Kinesin motor domain signature: A247-A297	PROFILESCAN
					Kinesin heavy chain signatures PR00380:	BLIMPS-PRINTS
						BLAST-PRODOM
			_		MICROTUBULES KINESINLIKE KINESIN MITOSIS	
		_			KINESIN MOTOR DOMAIN DM00198 P46871 3-343:	BLAST-DOMO
					E23-Q370	
*==					ATP/GTP binding site (P-loop): G109-T116	MOTIFS
					motif: G262-E273	MOTIFS
28	5677408CD1	351	T5 S22 T77	N20	binding site (P-loop): G38-T45	MOTIFS
			S92 S136 S186			
			\$286			
			8196			
			534.			

SEQ	Incyte				Signature Sequences, Domains and Motifs	
(I	Polypeptide	_ <u>`</u>				Methods and
NO:	A	Resi	Sites	Sites		Databases
29	5982278CD1	856	318 S807	N48 N49 N345	Kinesin motor domain: R31-E466	HMMER-PFAM
			T84 S125	N361 N565	dome	BLIMPS-BLOCKS
			S221 T266 S334 T347	N800	P25-D39; K69-Q85; G103-G124 G130-F140; V016-F034. G283-T307 F333-F374. M385-P415	-
			T369 T580 S710			PROFILESCAN
			S763		ignatures F	BI,TMPS-PRTNTS
			S3 T			
·			S18		PROTEIN MOTOR MICROTUBULES ATPBINDING	BLAST-PRODOM
			8341		COILED COIL KINESINLIKE SIMILAR MITOTIC	
			T382		PROTEIN1 PD013891: E664-L841	
71124			T622 T663 T679 T793 S802		KINESIN MOTOR DOMAIN DM00198   Q02241   4-443: 84-1446	BLAST-DOMO
					ATP/GTP binding site (P-loop): G112-T119	MOTIFS
					motor domain motif: S33	MOTIFS
30	6437362CD1	1056	S256 S377	N272 N275	986-S1049	HMMER-PFAM
			S979 S38	N475 N609	Calbonin family repeat BL01052: F44-I69	BLIMPS-BLOCKS
			3173	-	LIM domain: F1008-L1022	BLIMPS-BLOCKS
			S32		LIM domain signature: K964-S1047	PROFILESCAN
			T45		i i	BLIMPS-PRINTS
			2 0	•	C55-L72	
			S951 S972 S1027		CALPONIN FAMILY REPEAT DM01491   P51911   6- 147: P4-S116	BLAST-DOMO
•			3109		ਾਨ	MOTITION
			S16		Comatii mocti:	
			T537			
			S647			
×			S710 S753 S889 S974 S981 Y179 Y294			
31	4173970CD1	1569			: K347-N379;	HMMER-PFAM
					T144-V1/6; S1//-G209; D212-E244; N240-K2/8; T279-V311; S314-K346	
32	2772751CD1	089	S212 S340	N233	signal_cleavage: M1-A18	SPSCAN
			S357 T104	1 N56 N66	signal_peptide: M1-A18	HMMER
			T159 T276 T328	N89	transmembrane domain:	HMMER
			T342	- 10	ı,	
					AC133 ANTIGEN PROMININ HOMOLOG: M1-R333	BLAST_PRODOM

SEQ	Incyte	Amino		Potential	Signature Sequences, Domains and Motifs	Analytical
El I	Polypeptide Acid	Acid	hor	ylation Glycosylation		Methods and
NO:	ID	Residues Sites		Sites		Databases
33	2793768CD1	590	S3 S373 S432	N101 N166	signal_cleavage: M1-S56	SPSCAN
		. —	S461 S495 S499	N233	Ankyrin repeat: R40-R72, Q73-T102	HMMER_PFAM
			S548 S56 S561		SIMILAR TO ANKYRIN REPEAT REGION OF FOWLPOX BLAST PRODOM	BLAST_PRODOM
			S586 S83 T199		VIRUS BAMHIORF7 PROTEIN:	
			T228 T234 T255		W75-H284, G348-Y469	
÷			T286 T331 T354			
_			T357 T433 T445			
			T454 T534 T547			
			T588			
34	3035248CD1	315	S151 S291 T115		signal_cleavage: M1-G26	SPSCAN
			T207 T273		signal_peptide: M1-S24	HMMER
					transmembrane domain: P4-F20	HMMR
					INTERMEDIATE FILAMENT ASSOCIATED PROTEIN	BLAST_PRODOM
<del></del>					K147~T218	
					TROPOMYOSIN ALTERNATIVE SPLICING SIGNAL	BLAST_PRODOM
					PRECURSOR CHAIN: E39-S213	
					INTERMEDIATE FILAMENT: K121-T218	BLAST_DOMO

Table 4

10.1.m.1.0.1	1,000		0.10.21	E (2000)	2 7 5	10:4:200	77
SEQ ID NO:	Polynucleotide ID	Length	Fragment(s)	בניקבייניי בי ממייניייי בי ממייניייייי	2		
35	1889577CB1	2345	902-952	-	(BRAXNOT01)	1	284
35	1889577CB1	2345		$\sim$	PROSTUT04)	777	1278
35	1889577CB1	2345		_	(PROSTUT12)	102	646
35	1889577CB1	2345			(ADRENON04)	6641	2066
35	1889577CB1	2345		1 1	BSTMNON02)	1608	1840
35	1889577CB1	2345			(BRAINOT10)	1873	2345
35	1889577CB1	2345		$\sim$	OT01)	1516	1769
35	1889577CB1	2345			(DRGLNOT01)	1016	1563
35	1889577CB1	2345		1649402F6 (PROS	(PROSTUT09)	384	968
36	2427982CB1	709		71263527V1		624	709
36	2427982CB1	709		71247061V1		T	683
37	2470833CB1	1569	1-721	-	PROSNOT15)	Ţ	497
37	2470833CB1	1569		868966R6 (LUNGAST01	ST01)	842	1407
37	2470833CB1	1569		ı	(BRONNOT01)	086	1569
37	2470833CB1	1569			(SPLNNOT04)	213	1197
37	2470833CB1	1569			NOT02)	493	730
37	2470833CB1	1569			(UCMCNOT02)	207	652
38	2080579CB1	1172	1-148, 686-854	868135H1 (BRAITUT03	UT03)	368	632
38	2080579CB1	1172			(293TF1T01)	1	433
38	2080579CB1	1172			(UTRSNOT08)	583	1143
38	2080579CB1	1172			(LUNGFET05)	939	1172
38	2080579CB1	1172		5174845H1 (EPIB	(EPIBTXT01)	442	637
39	2156553CB1	2380	1-360, 2126-2380, 1121-1655	ľ	(FIBPFEN06)	П	260
39	2156553CB1	2380		2916949T6 (THYMFET03	FET03)	1769	2380
39	2156553CB1	2380		DI	(BRAITUT03)		937
39	2156553CB1	2380		2916949F6 (THYM	(THYMFET03)	1	1886
39	2156553CB1	2380		_	(BRAINOT09)		1224
39 .	2156553CB1	2380		1 —	(BLADNOT06)		1582
39	2156553CB1	2380		, –	(OVARDIT01)		531
39	2156553CB1	2380		1758833H1 (PITU	(PITUNOT03)		485
40	2182855CB1	4396	2141-2667, 1- 1505, 1737- 1890, 3803-	1	(BRSTNOT14)	3897	4396
40	2182855CB1	4396	0	2967273F6 (SCOR	(SCORNOTO4)	580	1116

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SEQ ID NO:	Folynucleotide ID	Length	Fragment(s)	מפלתפוונים דרמאוווניונים		
40	2182855CB1	4396		1611084F6 (COLNTUT06)	3554	3992
40	2182855CB1	4396		1785722H1 (BRAINOT10)	2610	2857
07	2182855CB1	4396		2182855F6 (SININOT01)	3389	3914
07	2182855CB1	4396		1484284F6 (CORPNOT02)	-1	607
40	2182855CB1	4396		g2276318	61	3799
40	2182855CB1	4396		2321435H1 (OVARNOT02)	2609	2794
40	2182855CB1	4396		1578313H1 (DUODNOT01)	1452	1546
40	2182855CB1	4396			2925	3464
40	2182855CB1	4396		õ	1811	2391
41	2242106CB1	1831	1~509, 626- 1018	965728R1 (BRSTNOT05)	1283	1831
41	2242106CB1	1831		1650350F6 (PROSTUT09)	322	1031
41	2242106CB1	1831		1396324T1 (THYRNOT03)	948	1660
41	2242106CB1	1831		6843794H1 (KIDNTMN03)	191	985
41	2242106CB1	1831		956964T1 (KIDNNOT05)	1019	1667
41	2242106CB1	1831		70846228V1	1	216
42	2726877CB1	3249	1979-2045, 2854-2873, 1857-1919, 2543-2608	3728286F6 (SMCCNON03)	1073	1504
42	2726877CB1	3249		3645568F6 (LUNGNOT34)	383	933
42	2726877CB1	3249		4969912H1 (KIDEUNC10)	151	425
42	2726877CB1	3249		2500944T6 (ADRETUTOS)	1254	1839
42	2726877CB1	3249		2726877F6 (OVARTUT05)	1603	2060
42	2726877CB1	3249		4195125T6 (COLITUT02)	2652	3249
42	2726877CB1	3249		)	901	1191
42	2726877CB1	3249		4195125F6 (COLITUT02)	2257	2810
42	2726877CB1	3249		5492146H1 (DRGTNON04)	2128	2380
42	2726877CB1	3249		3894803H1 (TLYMNOT05)	1	296
42	2726877CB1	3249		3728286T6 (SMCCNON03)	1682	2279
43	2738233CB1	4133	1-194, 4026-4133, 1607-2971, 3066-3186	1649466F6 (PROSTUT09)	2737	3394
43	2738233CB1	4133		2267313R6 (UTRSNOT02)	2211	2730
43	2738233CB1	4133		g3882312_CD	126	3516
43	2738233CB1	4133		- 1	2710	3006
4.3	Z/38233CBL	4133		2242201F6 (PANCTUT02)	1538	2077

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SEQ ID NO:	Polynucleotide ID	Length	Fragment(s)			
43	2738233CB1	4133		653470R6 (EOSINOT03)	685	1068
43	2738233CB1	4133		2267313T6 (UTRSNOT02)	3481	4109
43	2738233CB1	4133		,  – ,	7	277
43	2738233CB1	4133		J~	1948	2420
43	2738233CB1	4133		1 1	1394	1704
43	2738233CB1	4133		1	217	469
43	2738233CB1	4133		1	3959	4133
43	2738233CB1	4133		1 1	3370	4017
43	2738233CB1	4133		1486351H1 (CORPNOT02)	1236	1463
43	2738233CB1	4133		3765643F6 (BRSTNOT24)	277	758
44	1833116CB1	1754	1700-1754	2852676F6 (BRSTTUT13)	25	529
44	1833116CB1	1754			539.	839
44	1833116CB1	1754			632	886
44	1833116CB1	1754		J	979	1615
44	1833116CB1	1754		1442616R1 (THYRNOT03)	1247	1754
44	1833116CB1	1754		$\sim$	1	284
44	1833116CB1	1754		5172858H1 (EPIBTXT01)	331	588
44	1833116CB1	1754		1920612R6 (BRSTTUT01)	998	1402
45	001799CB1	2713	1-27, 1464-2008	5994129H1 (FTUBTUT02)	587	912
45	001799CB1	2713		4245126H1 (BRABDIT01)	1982	2239
45	001799CB1	2713			1	549
45	001799CB1	2713		5054327H1 (COLATMT01)	1456	1726
45	001799CB1	2713		6739739H1 (BRAFDIT02)	2153	2713
45	001799CB1	2713		71336820V1	276	868
45	001799CB1	2713		$\rightarrow$	884	1198
45	001799CB1	2713			1575	1981
45	001799CB1	2713		3515211H1 (LUNGNOT33)	1940	2217
45	001799CB1	2713		$\overline{}$	907	1471
45	001799CB1	2713			1401	1685
45	001799CB1	2713		4771110H1 (BRATNOT02)	1715	1990
46	119814CB1	1768	1-688	2637776F6 (BONTNOT01)	896	1494
46	119814CB1	1768		119814R1 (MUSCNOT01)	739	1406
46	119814CB1	1768		2638913F6 (BONTNOT01)	1410	1768
46	119814CB1	1768		$\vdash$	656	940
46	119814CB1	1768		g2184959	349	846
46	119814CB1	1768		g1218792	т-1	579
46	119814CB1	1768		2395927T6 (THP1AZT01)	20	182

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SEQ ID NO:	Polynucleotide ID	Length	Fragment(s)	מסווטוולס אל ססווטוולס אולי		
47	1295420CB1	3287	914-1321,	148434H1 (FIBRNGT01)	812	1028
47	1295420CB1	3287		2697808H1 (UTRSNOT12)	675	877
47	1295420CB1	3287		5778293H1 (BRAXNOT03)	1185	1447
47	1295420CB1	3287		2149151F6 (BRAINOT09)	1661	2176
47	1295420CB1	3287		2883729F6 (SINJNOT02)	1	515
47	1295420CB1	3287		1888639H1 (BLADTUT07)	898	1147
47	1295420CB1	3287		l	2720	3278
47	1295420CB1	3287			1334	2009
47	1295420CB1	3287		998245R1 (KIDNTUT01)	2118	2637
47	1295420CB1	3287		1	323	816
47	1295420CB1	3287		4027885H1 (BRAINOT23)	666	1261
47	1295420CB1	3287		835343R1 (PROSNOT07)	2247	2814
47	1295420CB1	3287		1367731R1 (SCORNON02)	2856	3287
48	1309364CB1	1748	1-49, 1037-1135		1162	1416
48	1309364CB1	1748		4904644F6 (TLYMNOTO8)	799	1399
48	1309364CB1	1748		2914466F6 (THYMFET03)	₽	536
48	1309364CB1	1748		-	678	933
48	1309364CB1	1748		1309364F6 (COLNFET02)	1227	1748
48	1309364CB1	1748		3727909H1 (SMCCNON03)	513	816
49	1315267CB1	2163	705-799	898915H1 (BRSTTUT03)	1839	2163
49	1315267CB1	2163		465550R6 (LATRNOT01)	673	1257
49	1315267CB1	2163			1029	1630
49	1315267CB1	2163		5191222F6 (OVARDIT06)	1	538
49	1315267CB1	2163			426	1041
49	1315267CB1	2163		1315267F6 (BLADTUT02)	1434	2022
50	1403289CB1	1615	1119-1170	101	489	1121
50	1403289CB1	1615		059048R6 (MUSCNOT01)	1080	1615
50	1403289CB1	1615		3502723H1 (ADRENOT11)	932	1225
50	1403289CB1	1615		1403289F6 (LATRTUT02)	1	601
51	1607607CB1	1356	1–157, 1263–1356	7262994H1 (UTRETMC01)	252	929
51	1607607CB1	1356			350	1048
51	1607607CB1	1356		-	1	275
51	1607607CB1	1356		1607607F6 (LUNGNOT15)	872	1356
52	1660025CB1	1268	1-88, 424- 459, 775-836	2580277F6 (KIDNTUT13)	213	932
			222			

בייל בייל בייל בייל בייל בייל בייל בייל	ׅׅ֓֝֝֝֡֜֜֝֜֜֜֝֜֜֜֜֜֜֜֜֜֜֜֜֜֜֜֜֜֜֜֜֜֜֜֜֜֜	1				
Polynucleotide ID	Length	Fragment(s)		ין לתוניה בין היים היים היים היים היים היים היים הי		110-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1-1
1660025CB1	1268		2172241H1	(ENDCNOT03)		240
1660025CB1	1268		1988094R6	ľ	750	1268
.660025CB1	1268		1756804R6	$  \sim  $	248	996
1796836CB1	2554	1177-1245, 1-61	2280307T6	(PROSNON01)	687	1302
796836CB1	2554		4971676H1	(HELATXT02)		227
796836CB1	2554		2582430T6	(KIDNTUT13)	1198	1545
796836CB1	2554		2497103T6	(ADRETUTO5)	1949	2533
1796836CB1	2554		2534742H1	(BRAINOT18)	1527	1758
796836CB1	2554		2553754T6	(THYMNOT03)	1817	2528
796836CB1	2554			(OVARTUTO5)	1630	1863
796836CB1	2554		$\sim$	TESTNOT03)	1311	1568
796836CB1	2554		2938533H1	(THYMFET02)	1056	1326
796836CB1	2554			(PITUDIR01)	47	684
796836CB1	2554			(BRSTNOT05)	2162	2554
796836CB1	2554			(HEARFET02)	319	877
880670CB1	1216	605-636	2889280T7	(LUNGFET04)	489	1192
880670CB1	1216		1358092F1	(LUNGNOT09)	246	920
880670CB1	1216		$\sim$	OVARTUT01)	657	1216
880670CB1	1216		2529604H1	(GBLANOT02)		357
913976CB1	1457	1-446, 1406-1457	3736188F6	(SMCCNOS01)	1173	1428
2913976CB1	1457	•	2913976F6	(KIDNTUT15)		520
913976CB1	1457		4645636H1	(PROSTUT20)	1187	1449
913976CB1	1457		4331439H1	(KIDNNOT32)	461	712
913976CB1	1457		4643722H1	(PROSTMT03)	1108	1325
913976CB1	1457		1312116F1	(COLNFET02)	583	1127
3092084CB1	1636	857-1636	1709866F6	(PROSNOT16)	1097	1634
092084CB1	1636		2807436F6	(BLADTUTO8)	1277	1636
092084CB1	1636		6906626H1	(MUSLTDR02)	1	610
092084CB1	1636		SBMA03169F	1	597	1181
092084CB1	1636		3092084F6	(BRSTNOT19)	543	1147
882482CB1	1742	1-82, 923-994	2286328X19	F1 (BRAINON01)	742	1251
882482CB1	1742		- 1	(PENCNOT01)	299	926
882482CB1	1742			LVENNOT03)	1088	1742
882482CB1	1742		3175528F6	(UTRSTUT04)	1	317
882482CB1	1742		1232503F6	(LUNGFET03)	096	1553
	1660025CB1 1660025CB1 1796836CB1 1796836CB1 1796836CB1 1796836CB1 1796836CB1 1796836CB1 1796836CB1 1796836CB1 1796836CB1 1796836CB1 1796836CB1 1796836CB1 1796836CB1 1796836CB1 2880670CB1 2880670CB1 2880670CB1 2880670CB1 2913976CB1 2913976CB1 2913976CB1 2913976CB1 2913976CB1 3882482CB1 3882482CB1 3882482CB1 3882482CB1 3882482CB1 3882482CB1		1268 1268 1268 1268 1268 1268 1268 1674 1674 1636 1636 1636 1636 1636 1636 1636 1636 1636 1636 1636 1636 1636 1636 1636 1636 1636 1636 1636 1636 1636 1636 1636 1636 1742 1742 1742	1268 1268 1268 1268 1268 1264 1-61 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2554 2605-636 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1216 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 1217 12	1268	1268

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Polynucleotide	Incyte	Sequence	Selected	Sequence Fragments	5' Position	3' Position
SEQ ID NO:	Polymucleotide   ID	Length	Fragment(s)			
57	3882482CB1	1742		2286328X16F1 (BRAINON01)	402	696
58	4933451CB1	602	1-401	4931591H1 (BRSTTUT20)	341	602
58	4933451CB1	602		2502933F6 (CONUTUTO1)	1	479
59	5043904CB1	3237	1964-2007,	2153280F6 (BRAINOT09)	335	888
			2911~2969, 993~1057, 1328~1589			
59	5043904CB1	3237		5043904R6 (PLACFER01)		583
59	5043904CB1	3237		075538H1 (THP1PEB01)	1604	1818
59	5043904CB1	3237		3189755X301D1 (THYMNONO4)	1131	1674
59	5043904CB1	3237		_ ;	614	1177
59	5043904CB1	3237			2026	2510
59	5043904CB1	3237		$  \cdot  $	2686	3237
59	5043904CB1	3237		1365052R6 (SCORNON02)	2440	2957
59	5043904CB1	3237		3250182H1 (SEMVNOT03)	1831	2128
59	5043904CB1	3237			2266	2513
59	5043904CB1	3237		3804331H1 (BLADTUT03)	1777	2089
0.9	5202390CB1	3640	1412-1489,	2544502H2 (UTRSNOT11)	2656	2927
			2294-2508,			
			3244-3640	:		
09	5202390CB1	3640		2321656R6 (OVARNOT02)	2039	2448
09	5202390CB1	3640		2557486F6 (THYMNOT03)		629
60	5202390CB1	3640		1441193F6 (THYRNOT03)	3317	3640
90	5202390CB1	3640		2844888H1 (DRGLNOT01)	864	1009
09	5202390CB1	3640		_	3047	3352
90	5202390CB1	3640		2122377F6 (BRSTNOT07)	2844	3297
60	5202390CB1	3640			2322	2662
60	5202390CB1	3640		5092148H1 (UTRSTMR01)	685	961
90	5202390CB1	3640			525	3562
60	5202390CB1	3640		Н	1590	1851
60	5202390CB1	3640		- 1	1999	2279
90	5202390CB1	3640		듸	411	738
60	5202390CB1	3640		<u>, -i</u> ]	2600	2796
60	5202390CB1	3640		(BRAINOT03	1041	1567
60	5202390CB1	3640			1431	1694
09	5202390CB1	3640		2842892F6 (DRGLNOT01)	1604	2220

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Polymucleotide	Incyte	Sequence	Selected	Sequence Fra	Fragments	5' Position	3' Position
SEQ ID NO:	Polynucleotide	Length	Fragment(s)				
61	5526375CB1	2111	1-50, 1540-2111	l	(BRSTNOT16)	1004	1278
61	5526375CB1	2111		ı	(LIVRTUT04)	1600	2111
61	5526375CB1	2111		[ -	(KIDNNOT25)	1263	1529
61	5526375CB1	2111		ı	(KIDNFET02)	729	1224
61	5526375CB1	2111		ſ <sup></sup>	(PROSTUS19)	1385	1626
61	5526375CB1	2111			(IDNFET02)	1242	1479
61	5526375CB1	2111			(KIDNNOT25)	518	788
61	5526375CB1	2111		2580307F6 (F	(KIDNTUT13)		519
61	5526375CB1	2111		2070882F6 (J	(ISLTNOT01)	225	715
62	5677408CB1	1389	1-177	535789R1 (AI	(ADRENOTO3)	1079	1389
62	5677408CB1	1389		881149T6 (TE	HYRNOT02)	793	1378
62	5677408CB1	1389		6023544H1 (1	(TESTNOT11)	736	1017
62	5677408CB1	1389			(THYRNOT02)	1	772
63	5982278CB1	3331	809-1149, 1755-1989	_	(CONDTUT01)	2203	2470
63	5982278CB1	3331		1390622H1 (F	EOSINOT01)	2029	2257
63	5982278CB1	3331		<u> </u>	EPIMNOT01)	1740	1993
63	5982278CB1	3331		$\overline{}$	293TF4T01)	2462	2796
63	5982278CB1	3331		3405843H1 (E	(ESOGNOT03)	919	1174
63	5982278CB1	3331			(CONDIUTOI)	2611	2862
63	5982278CB1	3331		_	UTRSNOT06)	1619	1733
63	5982278CB1	3331		g1521431		2559	3331
63	5982278CB1	3331		3591491H1 (2	(293TF5T01)	1674	1981
63	5982278CB1	3331		$\sim$	TLYMNOT05)	1464	1726
63	5982278CB1	3331			(MEGBUNT01)	2280	2536
63	5982278CB1	3331		$\sim$	TBLYNOT01)	1370	1562
63	5982278CB1	3331			(THYMNOTOS)	24	624
63	5982278CB1	3331		4983673H1 (F	HELATXT05)	1865	2134
63	5982278CB1	3331		g34671_CD		130	3166
63	5982278CB1	3331		3449505X304I	3449505X304D1 (UTRSNON03)	942	1450
63	5982278CB1	3331		2640427T6 (I	(LUNGIUT08)	2646	3309
63	5982278CB1	3331			(SPLNFET02)	1	514
64	6437362CB1	3558	1-428,	720069R6 (SY	(SYNOOAT01)	3159	3558
			923-1583				
64	6437362CB1	3558			(BRSTNOT13)	2462	2730
64	6437362CB1	3558		1568793H1 (U	(UTRSNOT05)	1264	1467

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Polynucleotide	Incyte	Sequence	Selected	Sequence Fragments	5' Position	3' Position
SEQ ID NO:	Polynucleotide   ID	Length	Fragment(s)			-
64	6437362CB1	3558		987366H1 (LVENNOT03)	763	1088
64	6437362CB1	3558		g5689540_CD	678	3298
64	6437362CB1	3558		-	1	500
64	6437362CB1	3558		1988667R6 (LUNGAST01)	2160	2709
64	6437362CB1	3558			981	1248
64	6437362CB1	3558		6437362H1 (LUNGNON07)	432	1052
₹9	6437362CB1	3558		_	1760	2328
64	6437362CB1	3558		3208061H1 (PENCNOT03)	2706	2948
64	6437362CB1	3558		1 -	1433	2050
64	6437362CB1	3558		865171R1 (BRAITUTO3)	2929	3483
65	4173970CB1	5373	3418-5373,	829704R1 (PROSTUT04)	1356	1950
			1-186, 1641-2444, 857-1058			
65	4173970CB1	5373		5604442H1 (MONOTXN03)	2038	2310
65	4173970CB1	5373		1708630F6 (PROSNOT16)	3923	4539
65	4173970CB1	5373		4561514F6 (KERATXT01)	4397	5197
65	4173970CB1	5373			3691	4247
65	4173970CB1	5373		1433309R1 (BEPINON01)	3666	4214
65	4173970CB1	5373		$\sim$	1	494
65	4173970CB1	5373		_	5043	5373
65	4173970CB1	5373		$\overline{}$	2520	3017
65	4173970CB1	5373		4173970F6 (SINTNOT21)	2115	2677
65	4173970CB1	5373		2277402R6 (PROSNON01)	099	1229
65	4173970CB1	5373		1708630T6 (PROSNOT16)	4639	5347
65	4173970CB1	5373		5944958H1 (COLADITO5)	1710	2018
65	4173970CB1	5373			1133	1687
65	4173970CB1	5373		g2737563	1885	2171
65	4173970CB1	5373		209752R1 (SPLNNOT02)	3040	3690
65	4173970CB1	5373			210	996
65	4173970CB1	5373		T	4284	4560
65	4173970CB1	5373		516280R6 (MMLR1DT01)	2996	3438
99	2772751CB1	4333	2456-3205,	70475866V1	507	1008
			1-23, 1459-1957, 3695-4333			
66	2772751CB1	4333		70472414V1	2045	2690

		!	Table + (cont	COIII.)	i	!
Polynucleotide	Incyte	Sequence	Selected	Sequence Fragments	5' Position	3' Position
SEQ ID NO:	Polynucleotide ID	Length	Fragment(s)			
99	2772751CB1	4333		3402774H1 (ESOGNOT03)		256
99	2772751CB1	4333		70475304V1	1481	2093
99	2772751CB1	4333		70475403V1	976	1554
99	2772751CB1	4333		70470913V1	1545	2109
99	2772751CB1	4333		70747026V1	3561	4054
99	2772751CB1	4333		g3923880	319	678
99	2772751CB1	4333		1298255T6 (BRSTNOT07)	3001	3690
99	2772751CB1	4333		70472159V1	1049	1556
99	2772751CB1	4333		70472656V1	2219	2925
99	2772751CB1	4333		g5340324	24	467
99	2772751CB1	4333 .		6849173H1 (KIDNTMN03)	3801	4333
99	2772751CB1	4333		6221536U1	2737	3452
19	2793768CB1	2213	2186-2213, 1066-1156	70843048V1	1646	2213
29	2793768CB1	2213		2026465R6 (KERANOT02)	709	1268
67	2793768CB1	2213		7712268H1 (TESTTUE02)	1621	2213
29	2793768CB1	2213		g1958420	-	421
29	2793768CB1	2213		6584157H1 (ESOGTMC01)	984	1576
29	2793768CB1	2213		2793768F6 (COLNTUT16)	131	777
67	2793768CB1	2213		71279716V1	1510	2183
89	3035248CB1	1142	1-55, 555-605	71515027V1	585	1140
89	3035248CB1	1142		71486327V1	402	787
89	3035248CB1	1142		71514455V1	⊣	576

Table 5

	COTON	
Polynucleotide	Incyte Droicet ID	Representative Library
י אין עד אין טיי	1000577071	DDAC INTIM 1 2
35	18895//CB1	PROSTUTI Z
36	2427982CB1	DRGCNOT01
37	2470833CB1	LUNGAST01
38	2080579CB1	UTRSNOT08
39	2156553CB1	THYMFET03
40	2182855CB1	SCORNOT04
41	2242106CB1	COLNPOT01
42	2726877CB1	LUNGNOT34
43	2738233CB1	MENITUT03
44	1833116CB1	THYRNOT03
45	001799CB1	BRSTTUT02
46	119814CB1	MUSCNOT01
47	1295420CB1	BRAITUT12
48	1309364CB1	TL YMUNTO1
49	1315267CB1	BLADTUT02
50	1403289CB1	LATRIUT02
51	1607607CB1	BRAIDIT01
52	1660025CB1	BRAWNOT01
53	1796836CB1	BRSTNOT05
54	2880670CB1	OVARTUT01
55	2913976CB1	ENDCNOT04
56	3092084CB1	HEAANOT01
57	3882482CB1	SPLMNOT11
58	4933451CB1	BRSTTUT20
53	5043904CB1	PLACFER01
0.9	5202390CB1	TESTITUT02
61	5526375CB1	KIDNFET02
62	5677408CB1	ADRENOT03
63	5982278CB1	SPLNFET02
79	6437362CB1	BRAINOT23
65	4173970CB1	BRSTNOT07
99	2772751CB1	BRSTNOT07
67	2793768CB1	UTRSNOT12
68	3035248CB1	TLYMNOT05

## Table (

Library	Vector	Library Description
ADRENOT03	PSPORT1	Library was constructed using RNA isolated from the adrenal tissue of a 17-year-old Caucasian male, who died from cerebral anoxia.
BLADTUT02	PINCY	ted using emale duri vasive tra
BRAIDIT01	pincy	Library was constructed using RNA isolated from diseased brain tissue. Patient history included multiple sclerosis, type II lesion.
BRAINOT23	PINCY	RNA ng a stic s con nvuls inclu acqui
BRAITUT12	pincy	Library was constructed using RNA isolated from brain tumor tissue removed from the left frontal lobe of a 40-year-old Caucasian female during excision of a cerebral meningeal lesion. Pathology indicated grade 4 gemistocytic astrocytoma.
BRSTNOT01	PINCY	

Library	Vector	Library Description
BRSTNOT07	AINCA	constructed using RNA isolated from c
		43-year-old Caucasian female during a unilateral extended simple mastectomy. Pathology
		/ proliferative fib
		á.
		invasive grade 4, nuclear grade 3 mammary adenocarcinoma with extensive comedo necrosis.
TOMOTAMO GG	TMCW	anatemeted maine DNA included from discass, and type in diabetes.
DESCRIPTION	plinci	constructed using MNA isolated irom diseased breast tissue remove
		43-year-old Caucasian female during a unilateral extended simple mastectomy. Pathology
		/ proliferative fik
		itosis, and duct ectasia. Pathology for the associated tumor tissue inc
		4, nuclear grade 3 mammary adenocarcinoma with
BRSTTUT02	PSPORT1	Library was constructed using RNA isolated from breast tumor tissue removed from a 54-
		ř١
		indicated residual invasive grade 3 mammary ductal adenocarcinoma. The
		breast parenchyma exhibited proliferative fibrocystic changes without
All		10 axillary lymph nodes had metastatic
		Patient history included kidney infection and condyloma acuminatum. Family history
		included benign hypertension, hyperlipidemia, and a malignant colon neoplasm.
BRSTTUT20	DINCY	
07		66-year-old Black female during a unilateral extended simple mastectomy and fine needle
		. Pathology indicated invasive grade 4, nuclear grade 3 adenocarcinoma
		involved, including the deep surgical margin. Extensive angiolymphatic invasion was
		identified, including superficial dermal lymphatics. Metastatic grade 4 adenocarcinoma
		nodes tissue were positive for metastatic mammary carcinoma. Left chest wall biopsy
		netasta
		metastatic grade 4, nuclear grade 3, metastatic mammary carcinoma. The patient presented
		laise and
-		condary malignant neoplasm of the brain/spine, deficiency anemia, type II
		chronic renal failure, and normal delivery. Patient medications i
		yclophosphamide/epirubicin and 5-Fl
		history included benign hypertension, type II diabetes, hyperlipidemia, and depressive disorder in the mother.
COLMPOT01	DINCY	۱.,
		ucasian female during a total colectomy. Pathology indicated an inflam
		pseudopolvo; this tissue was associated with a focally invasive grade 2 adenocarcinoma
		ıltiple tubuvillous adenomas. Patient history included a benign nec

## Table 6 (cont.)

Library	Vector	Library Description
DRGCNOT01	pincy	constructed using RNA isolated from dorsal root ganglion tissue remove spine of a 32-year-old Caucasian male who died from acute pulmonary pneumonia, bilateral pleural and pericardial effusions, and malignant atural killer cell type). Patient history included probable cytomegal sepatic congestion and steatosis, splenomegaly, hemorrhagic cystitis, and Bell's palsy. Surgeries included colonoscopy, large intestine bilectomy, and nasopharyngeal endoscopy and biopsy; treatment included lerapy.
ENDCNOT04	pincy	Library was constructed using RNA isolated from coronary artery endothelial cell tissue removed from a 3-year-old Caucasian male.
HEAANOT01	pincy	was con artery clusion artery rattery tricula
KIDNFET02	pincy	const who w
LATRTUT02	pINCY	Library was constructed using RNA isolated from a myxoma removed from the left atrium of a 43-year-old Caucasian male during annuloplasty. Pathology indicated atrial myxoma. Patient history included pulmonary insufficiency, acute myocardial infarction, atherosclerotic coronary artery disease, hyperlipidemia, and tobacco use. Family history included benign hypertension, acute myocardial infarction, atherosclerotic coronary artery disease, and type II diabetes.
LUNGAST01	PSPORT1	Library was constructed using RNA isolated from the lung tissue of a 17-year-old Caucasian male, who died from head trauma. Patient history included asthma.
LUNGNOT34	pincy	ted using RNA isolated from lung tissue removed
MENITUT03	pINCY	Library was constructed using RNA isolated from brain meningioma tissue removed from a 35-year-old Caucasian female during excision of a cerebral meningeal lesion. Pathology indicated a benign neoplasm in the right cerebellopontine angle of the brain. Patient history included hypothyroidism. Family history included myocardial infarction and breast cancer.
MUSCNOT01	PBLUESCRIPT	Library was constructed at Stratagene (STR937209), using RNA isolated from the skeletal muscle tissue of a patient with malignant hyperthermia.

# Table 6 (cont.)

	Library Ovarmment	Vector	
			Describrion
	701011110	PSPORTI	constructed using RNA isolated from ovarian tumor tissue removed frocasian female during removal of the fallopian tubes and ovaries. Pa
			indicated grade 2 mucinous cystadenocarcinoma involving the entire left ovary. Patient history included mitral valve disorder, pneumonia, and viral hepatitis. Family history
			included atherosclerotic coronary artery disease, pancreatic cancer, stress reaction,
	PLACFER01	DINCY	The library was constructed using RNA isolated from placental tissue removed from a
		ı	etus, who died after 16 weeks' gestation from fetal demise and hyc
			Patient history included umbilical cord wrapped around the head (3 times) and the shoulders (1 time). Serology was positive for anti-CMV. Family history included multiple
<u></u>	PROSTUT12	PINCY	using F
			ucasian male during a radical prostatectomy. Pathology indicated an
[			٠,
	SCOPNOTIONAL	TMCV	partent presented with elevated prostate specific antigen (FSA).
-	# 0 T O T O T O T O T O T O T O T O T O T	Princi	constructed asting take isolated from acute bulmonary edema and bronchoppenmon'
			eural and pericardial effusions, and malignant lymphoma (
			hepatic congest
1(	_		d hemorrhage
)9			-
			eal endoscopy and biopsy; treatment included radiation then
	SPLNFET02	pINCY	s constructed using RNA
1			who died at 23 weeks' gestation.
	SPLNNOT11	pincy	constructed using RNA isolated from diseased spleen tissue
-			14-year-old Asian male during a total splenectomy. Pathology indicated changes
1			consistent with idiopathic thrombocytopenic purpura. The patient presented with bruising. Patient medications included Vincristing.
<u> </u>	TESTTUT02	pINCY	cted using RNA isolated from testicular tumor remo
			ian male during unil
<del>-</del>	THYMFET03	pINCY	Library was constructed using RNA isolated from thymus tissue removed from a Caucasian male fetus.
<u> </u>	THYRNOT03	pINCY	rs O
	,,		a 28-year-old Caucasian female during a complete
			lidicated a small nodule of adenomatous nyperplasia present in the left thyrold. Dathology for the associated tumor tissue indicated dominant follicular adenoma forming
			tor the associated randi trassue indicated commission fortificated adenough, appulated mass in the left thyroid.
	TLYMNOT05	pincy	Library was constructed RNA isolated from nonactivated Th2 cells. These cells were differentiated from umbilical cord CD4 T cells with IL-4 in the presence of anti-IL-12
_]			antibodies and B7-transfected COS cells.

## Table 6 (cont.)

Library	Vector	Library Description
TLYMUNT01 DINCY	PINCY	Library was constructed using RNA isolated from restingallogenic T-lymphocyte tissue removed from an adult (40-50-year-old) Caucasian male.
UTRSNOT08	pINCY	Library was constructed using RNA isolated from uterine tissue removed from a 35-year-old Caucasian female during a vaginal hysterectomy with dilation and curettage.
·		Pathology indicated that the endometrium was secretory phase with a benign endometrial polyp 1 cm in diameter. The cervix showed mild chronic cervicitis. Family history
		included atherosclerotic coronary artery disease and type 11 diabetes.
UTRSNOT12 PINCY	pincy	Library was constructed using RNA isolated from uterine myometrial tissue removed from a 41-year-old Caucasian female during a vaginal hysterectomy with dilation and curettage.
		The endometrium was secretory and contained fragments of endometrial polyps. Benign
_		associated tumor tissue indicated uterine leiomyoma. Patient history included ventral
		hernia and a benign ovarian neoplasm.

#### Table 7

Program	Description	Reference	Parameter Threshold
ABI FACTURA	A program that removes vector sequences and masks ambiguous bases in nucleic acid sequences.	Applied Biosystems, Foster City, CA.	
ABI/PARACEL FDF	A Fast Data Finder useful in comparing and annotating amino acid or nucleic acid sequences.	Applied Biosystems, Foster City, CA; Paracel Inc., Pasadena, CA.	Mismatch <50%
ABI AutoAssembler	A program that assembles nucleic acid sequences.	Applied Biosystems, Foster City, CA.	
 BLAST	A Basic Local Alignment Search Tool useful in sequence similarity search for amino acid and nucleic acid sequences. BLAST includes five functions: blastp, blastn, blastx, tblastn, and tblastx.	Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410; Altschul, S.F. et al. (1997) Nucleic Acids Res. 25:3389-3402.	ESTs: Probability value= 1.0E-8 or less Full Length sequences: Probability value= 1.0E-10 or less
 FASTA	A Pearson and Lipman algorithm that searches for similarity between a query sequence and a group of sequences of the same type. FASTA comprises as least five functions: fasta, fasta, fastx, tfastx, and ssearch.	Pearson, W.R. and D.J. Lipman (1988) Proc. Natl. Acad Sci. USA 85:2444-2448; Pearson, W.R. (1990) Methods Enzymol. 183:63-98; and Smith, T.F. and M.S. Waterman (1981) Adv. Appl. Math. 2:482-489.	ESTs: fasta E value=1.06E-6 Assembled ESTs: fasta Identity= 95% or greater and Match length=200 bases or greater; fastx E value=1.0E-8 or less Full Length sequences: fastx score=100 or greater
BLIMPS	A BLocks IMProved Searcher that matches a sequence against those in BLOCKS, PRINTS, DOMO, PRODOM, and PFAM databases to search for gene families, sequence homology, and structural fingerprint regions.	Henikoff, S. and J.G. Henikoff (1991) Nucleic Acids Res. 19:6565-6572; Henikoff, J.G. and S. Henikoff (1996) Methods Enzymol. 266:88-105; and Attwood, T.K. et al. (1997) J. Chem. Inf. Comput. Sci. 37:417-424.	Probability value= 1.0E-3 or less
HMMER	An algorithm for searching a query sequence against hidden Markov model (HMM)-based databases of protein family consensus sequences, such as PFAM.	Krogh, A. et al. (1994) J. Mol. Biol. 235:1501-1531; Sonnhammer, E.L.L. et al. (1988) Nucleic Acids Res. 26:320-322; Durbin, R. et al. (1998) Our World View, in a Nutshell, Cambridge Univ. Press, pp. 1-350.	PFAM hits: Probability value=1.0E-3 or less Signal peptide hits: Score=0 or greater

Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221; Wisconsin Package Program Manual, version 9, page M51-59, Genetics Computer Group, Madison, WI.

A program that searches amino acid sequences for patterns

Motifs

that matched those defined in Prosite.

### Table 7 (cont.)

	1 auto 1	Table / (colle.)	
Program	Description	Reference	Parameter Threshold
ProfileScan	An algorithm that searches for structural and sequence motifs in protein sequences that match sequence patterns defined in Prosite.	Gribskov, M. et al. (1988) CABIOS 4:61-66; Gribskov, M. et al. (1989) Methods Enzymol. 183:146-159; Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221.	Normalized quality score GCG-specified "HIGH" value for that particular Prosite motif. Generally, score=1.4-2.1.
Phred	A base-calling algorithm that examines automated sequencer traces with high sensitivity and probability.	Ewing, B. et al. (1998) Genome Res. 8:175-185; Ewing, B. and P. Green (1998) Genome Res. 8:186-194.	
Phrap	A Phils Revised Assembly Program including SWAT and CrossMatch, programs based on efficient implementation of the Smith-Waterman algorithm, useful in searching sequence homology and assembling DNA sequences.	Smith, T.F. and M.S. Waterman (1981) Adv. Appl. Math. 2:482-489; Smith, T.F. and M.S. Waterman (1981) J. Mol. Biol. 147:195-197; and Green, P., University of Washington, Seattle, WA.	Score= 120 or greater; Match length= 56 or greater
Consed	A graphical tool for viewing and editing Phrap assemblies.	Gordon, D. et al. (1998) Genome Res. 8:195-202.	2.
SPScan	A weight matrix analysis program that scans protein sequences for the presence of secretory signal peptides.	Nielson, H. et al. (1997) Protein Engineering 10:1-6; Claverie, J.M. and S. Audiç (1997) CABIOS 12:431-439.	Score=3.5 or greater
TMAP	A program that uses weight matrices to delineate transmembrane segments on protein sequences and determine orientation.	Persson, B. and P. Argos (1994) J. Mol. Biol. 237:182-192; Persson, B. and P. Argos (1996) Protein Sci. 5:363-371.	
TMHMMER	A program that uses a hidden Markov model (HMM) to delineate transmembrane segments on protein sequences and determine orientation.	Sonnhammer, E.L. et al. (1998) Proc. Sixth Intl. Conf. on Intelligent Systems for Mol. Biol., Glasgow et al., eds., The Am. Assoc. for Artificial Intelligence Press, Menlo Park, CA, pp. 175-182.	ial 2.
		,	

What is claimed is:

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1. An isolated polypeptide selected from the group consisting of:

- a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-34.
  - b) a naturally occurring polypeptide comprising an amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-34,
  - c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEO ID NO:1-34, and
- d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34.
- 2. An isolated polypeptide of claim 1 selected from the group consisting of SEQ ID NO:1-34.

3. An isolated polynucleotide encoding a polypeptide of claim 1.

- 4. An isolated polynucleotide encoding a polypeptide of claim 2.
- 5. An isolated polynucleotide of claim 4 selected from the group consisting of SEQ ID NO:35-68.
  - 6. A recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide of claim 3.
    - 7. A cell transformed with a recombinant polynucleotide of claim 6.
    - 8. A transgenic organism comprising a recombinant polynucleotide of claim 6.
    - 9. A method for producing a polypeptide of claim 1, the method comprising:
  - a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said
     cell is transformed with a recombinant polynucleotide, and said recombinant polynucleotide
     comprises a promoter sequence operably linked to a polynucleotide encoding the polypeptide of claim
     1, and
    - b) recovering the polypeptide so expressed.

- 10. An isolated antibody which specifically binds to a polypeptide of claim 1.
- 11. An isolated polynucleotide selected from the group consisting of:
- a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting
   of SEQ ID NO:35-68,
  - b) a naturally occurring polynucleotide comprising a polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:35-68,
    - c) a polynucleotide complementary to a polynucleotide of a),
    - d) a polynucleotide complementary to a polynucleotide of b), and
    - e) an RNA equivalent of a)-d).
  - 12. An isolated polynucleotide comprising at least 60 contiguous nucleotides of a polynucleotide of claim 11.
- 13. A method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 11, the method comprising:
  - a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and
  - b) detecting the presence or absence of said hybridization complex, and, optionally, if present, the amount thereof.
    - 14. A method of claim 13, wherein the probe comprises at least 60 contiguous nucleotides.
  - 15. A method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 11, the method comprising:
  - a) amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and
  - b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.
  - 16. A composition comprising a polypeptide of claim 1 and a pharmaceutically acceptable excipient.

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17. A composition of claim 16, wherein the polypeptide has an amino acid sequence selected from the group consisting of SEQ ID NO:1-34.

- 18. A method for treating a disease or condition associated with decreased expression of functional CYSKP, comprising administering to a patient in need of such treatment the composition of claim 16.
  - 19. A method for screening a compound for effectiveness as an agonist of a polypeptide of claim 1, the method comprising:
    - a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
    - b) detecting agonist activity in the sample.

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- 20. A composition comprising an agonist compound identified by a method of claim 19 and a pharmaceutically acceptable excipient.
- 21. A method for treating a disease or condition associated with decreased expression of functional CYSKP, comprising administering to a patient in need of such treatment a composition of claim 20.
- 22. A method for screening a compound for effectiveness as an antagonist of a polypeptide of claim 1, the method comprising:
  - a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
  - b) detecting antagonist activity in the sample.
- 23. A composition comprising an antagonist compound identified by a method of claim 22 and a pharmaceutically acceptable excipient.
  - 24. A method for treating a disease or condition associated with overexpression of functional CYSKP, comprising administering to a patient in need of such treatment a composition of claim 23.
  - 25. A method of screening for a compound that specifically binds to the polypeptide of claim 1, said method comprising the steps of:
  - a) combining the polypeptide of claim 1 with at least one test compound under suitable conditions, and
- b) detecting binding of the polypeptide of claim 1 to the test compound, thereby identifying a

compound that specifically binds to the polypeptide of claim 1.

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26. A method of screening for a compound that modulates the activity of the polypeptide of claim 1, said method comprising:

- a) combining the polypeptide of claim 1 with at least one test compound under conditions permissive for the activity of the polypeptide of claim 1,
  - b) assessing the activity of the polypeptide of claim 1 in the presence of the test compound, and
- c) comparing the activity of the polypeptide of claim 1 in the presence of the test compound with the activity of the polypeptide of claim 1 in the absence of the test compound, wherein a change in the activity of the polypeptide of claim 1 in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide of claim 1.
- 27. A method for screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a sequence of claim 5, the method comprising:
- a) exposing a sample comprising the target polynucleotide to a compound, under conditions suitable for the expression of the target polynucleotide,
  - b) detecting altered expression of the target polynucleotide, and
- c) comparing the expression of the target polynucleotide in the presence of varying amounts of
   the compound and in the absence of the compound.
  - 28. A method for assessing toxicity of a test compound, said method comprising:
  - a) treating a biological sample containing nucleic acids with the test compound;
- b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at
  least 20 contiguous nucleotides of a polynucleotide of claim 11 under conditions whereby a specific
  hybridization complex is formed between said probe and a target polynucleotide in the biological
  sample, said target polynucleotide comprising a polynucleotide sequence of a polynucleotide of claim
  11 or fragment thereof;
  - c) quantifying the amount of hybridization complex; and
- d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.

29. A diagnostic test for a condition or disease associated with the expression of CYSKP in a biological sample comprising the steps of:

- a) combining the biological sample with an antibody of claim 10, under conditions suitable for the antibody to bind the polypeptide and form an antibody:polypeptide complex; and
- 5 b) detecting the complex, wherein the presence of the complex correlates with the presence of the polypeptide in the biological sample.
  - 30. The antibody of claim 10, wherein the antibody is:
  - a) a chimeric antibody,
  - b) a single chain antibody,
  - c) a Fab fragment,
  - d) a F(ab')<sub>2</sub> fragment, or
  - e) a humanized antibody.
- 15 31. A composition comprising an antibody of claim 10 and an acceptable excipient.
  - 32. A method of diagnosing a condition or disease associated with the expression of CYSKP in a subject, comprising administering to said subject an effective amount of the composition of claim 31.

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- 33. A composition of claim 31, wherein the antibody is labeled.
- 34. A method of diagnosing a condition or disease associated with the expression of CYSKP in a subject, comprising administering to said subject an effective amount of the composition of claim
   33.
  - 35. A method of preparing a polyclonal antibody with the specificity of the antibody of claim 10 comprising:
- a) immunizing an animal with a polypeptide having an amino acid sequence selected from
   the group consisting of SEQ ID NO:1-34, or an immunogenic fragment thereof, under conditions to elicit an antibody response;
  - b) isolating antibodies from said animal; and
  - c) screening the isolated antibodies with the polypeptide, thereby identifying a polyclonal antibody which binds specifically to a polypeptide having an amino acid sequence selected from the

group consisting of SEQ ID NO:1-34.

- 36. An antibody produced by a method of claim 35.
- 5 37. A composition comprising the antibody of claim 36 and a suitable carrier.
  - 38. A method of making a monoclonal antibody with the specificity of the antibody of claim 10 comprising:
  - a) immunizing an animal with a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34, or an immunogenic fragment thereof, under conditions to elicit an antibody response;
    - b) isolating antibody producing cells from the animal;
    - c) fusing the antibody producing cells with immortalized cells to form monoclonal antibodyproducing hybridoma cells;
      - d) culturing the hybridoma cells; and
    - e) isolating from the culture monoclonal antibody which binds specifically to a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34.
      - 39. A monoclonal antibody produced by a method of claim 38.

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- 40. A composition comprising the antibody of claim 39 and a suitable carrier.
- 41. The antibody of claim 10, wherein the antibody is produced by screening a Fab expression library.

- 42. The antibody of claim 10, wherein the antibody is produced by screening a recombinant immunoglobulin library.
- 43. A method for detecting a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34 in a sample, comprising the steps of:
  - a) incubating the antibody of claim 10 with a sample under conditions to allow specific binding of the antibody and the polypeptide; and
  - b) detecting specific binding, wherein specific binding indicates the presence of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34 in

the sample.

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44. A method of purifying a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34 from a sample, the method comprising:

- a) incubating the antibody of claim 10 with a sample under conditions to allow specific binding of the antibody and the polypeptide; and
- b) separating the antibody from the sample and obtaining the purified polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-34.
- 45. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:1.
  - 46. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:2.
  - 47. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:3.
  - 48. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:4.
  - 49. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:5.
- 20 50. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:6.
  - 51. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:7.
  - 52. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:8.
  - 53. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:9.
  - 54. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:10.
- 30 55. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:11.
  - 56. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:12.
  - 57. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:13.

58. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:14. 59. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:15. 5 60. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:16. 61. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:17. 10 62. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:18. 63. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:19. 64. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:20. 15 65. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:21. 66. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:22. 20 67. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:23. 68. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:24. 69. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:25. 25 70. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:26. 71. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:27. 30 72. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:28. 73. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:29, 74. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:30.

75. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:31.

- 76. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:32.
- 77. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:33.
- 78. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:34.
- 10 79. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:35.

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- 80. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:36.
- 81. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:37.
- 82. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ IDNO:38.
  - 83. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:39.
- 84. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:40.
  - 85. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:41.
  - 86. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:42.
    - 87. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID

NO:43.

88. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:44.

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- 89. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:45.
- 90. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:46.
  - 91. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:47.
- 15 92. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:48.
  - 93. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:49.

- 94. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:50.
- 95. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:51.
  - 96. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:52.
- 97. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:53.
  - 98. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:54.

	99. A polynucleotide of claim 11.	comprising the polynucleotide sequence of SEQ ID
NO:55		

- 5 100. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:56.
  - 101. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:57.
  - 102. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:58.

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- 103. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ IDNO:59.
  - 104. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:60.
- 20 105. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:61.
  - 106. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:62.
  - 107. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:63.
- 108. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID30 NO:64.
  - 109. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:65.

 $110.\,$  A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:66.

- 111. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ IDNO:67.
  - 112. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:68.

```
<110> INCYTE GENOMICS, INC.
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Asp Leu Phe Pro Asn Glu Asp Glu Gln Ser Pro Ala Pro Ser Pro
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                365
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                                                          600
Leu Ser Asp Ser Arg Thr Leu Ser Ser Ser Ser Met Asp Leu Ser
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Arg Arg Ser Ser Leu Val Gly
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<sup>&</sup>lt;212> PRT

<sup>&</sup>lt;213> Homo sapiens

<sup>&</sup>lt;220>

<sup>&</sup>lt;221> misc\_feature

<sup>&</sup>lt;223> Incyte ID No: 2427982CD1

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Met Leu Ala Asn Cys Glu Lys Leu Ser Leu Ser Thr Asn Cys Ile
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Glu Lys Ile Ala Asn Leu Asn Gly Leu Lys Asn Leu Arg Ile Leu
                                      70
                 65
                                                           75
Ser Leu Gly Arg Asn Asn Ile Lys Asn Leu Asn Gly Leu Glu Ala
                 80
                                      85
Val Gly Asp Thr Leu Glu Glu Leu Trp Ile Ser Tyr Asn Phe Ile
                 95
                                     100
Glu Lys Leu Lys Gly Ile His Ile Met Lys Lys Leu Lys Ile Leu
                110
                                     115
                                                          120
Tyr Met Ser Asn Asn Leu Val Lys Asp Trp Ala Glu Phe Val
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                                     130
                                                          135
Leu Ala Glu Leu Pro Cys Leu Glu Asp Leu Val Phe Val Gly Asn
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                                     145
                                                          150
Pro Leu Glu Glu Lys His Ser Ala Glu Asn Asn Trp Ile Glu Glu
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Ala Thr Lys Arg Val Pro Lys Leu Lys Leu Asp Gly Thr Pro
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Val Ile Lys Gly Asp Glu Glu Glu Asp Asn
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                                      25
Lys Tyr Leu Pro Pro Thr Ser Arg Lys Asp Pro Lys Phe Glu Glu
                 35
                                      40
Leu Gln Lys Val Leu Met Glu Trp Ile Asn Ala Thr Leu Leu Pro
                 50
                                      55
                                                           60
Glu His Ile Val Val Arg Ser Leu Glu Glu Asp Met Phe Asp Gly
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                                      70
Leu Ile Leu His His Leu Phe Gln Arg Leu Ala Ala Leu Lys Leu
                 80
                                      85
                                                           90
Glu Ala Glu Asp Ile Ala Leu Thr Ala Thr Ser Gln Lys His Lys
                 95
                                     100
Leu Thr Val Val Leu Glu Ala Val Asn Arg Ser Leu Gln Leu Glu
                110
                                     115
                                                          120
Glu Trp Gln Ala Lys Trp Ser Val Glu Ser Ile Phe Asn Lys Asp
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                                     130
                                                          135
Leu Leu Ser Thr Leu His Leu Leu Val Ala Leu Ala Lys Arg Phe
                140
                                     145
                                                          150
Gln Pro Asp Leu Ser Leu Pro Thr Asn Val Gln Val Glu Val Ile
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                                     160
Thr Ile Glu Ser Thr Lys Ser Gly Leu Lys Ser Glu Lys Leu Val
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                                     175
                                                          180
Glu Gln Leu Thr Glu Tyr Ser Thr Asp Lys Asp Glu Pro Pro
                                                          Lys
                                     190
                185
                                                          195
Asp Val Phe Asp Glu Leu Phe Lys Leu Ala Pro Glu Lys Val Asn
                200
                                     205
Ala Val Lys Glu Ala Ile Val Asn Phe Val Asn Gln Lys Leu Asp
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220
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Arg Leu Gly Leu Ser Val Gln Asn Leu Asp Thr Gln Phe Ala Asp
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                                    235
                                                         240
Gly Val Ile Leu Leu Leu Ile Gly Gln Leu Glu Gly Phe Phe
                245
                                    250
                                                         255
Leu His Leu Lys Glu Phe Tyr Leu Thr Pro Asn Ser Pro Ala Glu
                260
                                     265
Met Leu His Asn Val Thr Leu Ala Leu Glu Leu Leu Lys Asp Glu
                275
                                     280
Gly Leu Leu Ser Cys Pro Val Ser Pro Glu Asp Ile Val Asn Lys
                290
                                    295
                                                         300
Asp Ala Lys Ser Thr Leu Arg Val Leu Tyr Gly Leu Phe Cys
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His Thr Gln Lys Ala His Arg Asp Arg Thr Pro His Gly Ala Pro
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Asn
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Gly Leu Ala Gly Ser Ile Tyr Arg Glu Phe Glu Arg Leu Ile His
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                                     40
Cys Tyr Asp Glu Glu Val Val Lys Glu Leu Met Pro Leu Val Val
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Asn Val Leu Glu Asn Leu Asp Ser Val Leu Ser Glu Asn Gln Glu
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                 65
His Glu Val Glu Leu Glu Leu Arg Glu Asp Asn Glu Gln Leu
                 80
                                     85
Leu Thr Gln Tyr Glu Arg Glu Lys Ala Leu Arg Arg Gln Ala Glu
                 95
                                     100
                                                         105
Glu Lys Phe Ile Glu Phe Glu Asp Ala Leu Glu Gln Glu Lys Lys
                110
                                     115
Glu Leu Gln Ile Gln Val Glu His Tyr Glu Phe Gln Thr Arg Gln
                                     130
                125
Leu Glu Leu Lys Ala Lys Asn Tyr Ala Asp Gln Ile Ser Arg Leu
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Glu Glu Arg Glu Ser Glu Met Lys Lys Glu Tyr Asn Ala Leu His
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                                     160
Gln Arg His Thr Glu Met Ile Gln Thr Tyr Val Glu His Ile Glu
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                                     175
                                                         180
Arg Ser Lys Met Gln Gln Val Gly Gly Asn Ser Gln Thr Glu Ser
                                     190
                185
Ser Leu Pro Gly Arg Arg Tyr Ala Gly Arg Gly Val Glu Val
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Arg Gly Ala Arg Arg Gly Gly Gly Thr Gln Asp Ala Ala His Ala
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Ala Met Cys His Arg Arg Met Gly Val Arg Pro Ala Val Pro Leu Leu Thr Gln Arg Gly Ser Gly Glu Gly Lys Asp Ser Gly Ile Pro Thr His Arg Ser Ala Ser Arg Lys Gly Thr Gly Ala Arg Ser Leu Gly His Ser Glu Lys Pro Val Ser Thr Ala Thr Thr Ser Ala Pro Gly Lys Gly Lys Lys Gly Lys Ala Lys Arg Ala Thr Ala Leu Val Cys Pro Asn Leu Trp Glu Trp Asp Ala Pro Ser Thr Arg Met Gly Cys Ile Phe Thr Met Thr Phe Ser Ser Gly Asp Arg Gln Pro His His Leu Asn Arg Leu Pro Leu Ser Pro Lys Asn Pro Gln Ala Leu Gly Lys Thr Ile Pro Pro Lys His Pro Ser Val Pro Arg Arg Phe Ile Pro Ala Leu Gln Ala Pro Pro Asn His Leu Asp Gln Pro Pro 5/69

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Ala Thr Ala Glu Lys Ile Lys Cys Gln Gln Glu Ala Asp Ala Thr
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Asn Arg Val Ile Leu Leu Ala Asn Arg Leu Val Gly Gly Leu Ala
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                                      55
Ser Glu Asn Ile Arg Trp Ala Glu Ser Val Glu Asn Phe Arg Ser
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                                      70
Gln Gly Val Thr Leu Cys Gly Asp Val Leu Leu Ile Ser Ala Phe
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                                      85
Val Ser Tyr Val Gly Tyr Phe Thr Lys Lys Tyr Arg Asn Glu Leu
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                                                          105
Met Glu Lys Phe Trp Ile Pro Tyr Ile His Asn Leu Lys Val Pro
                110
                                     115
Ile Pro Ile Thr Asn Gly Leu Asp Pro Leu Ser Leu Leu Thr Asp
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                125
Asp Ala Asp Val Ala Thr Trp Asn Asn Gln Gly Leu Pro Ser Asp
                140
                                     145
                                                          150
Arg Met Ser Thr Glu Asn Ala Thr Ile Leu Gly Asn Thr Glu Arg
                155
                                     160
Trp Pro Leu Ile Val Asp Ala Gln Leu Gln Gly Ile Lys Trp Ile
                170
                                     175
Lys Asn Lys Tyr Arg Ser Glu Leu Lys Ala Ile Arg Leu Gly Gln
                185
                                     190
Lys Ser Tyr Leu Asp Val Ile Glu Gln Ala Ile Ser Glu Gly Asp
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                                     205
                                                          210
Thr Leu Leu Ile Glu Asn Ile Gly Glu Thr Val Asp Pro Val
                                                          Leu
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                                     220
                                                          225
Asp Pro Leu Leu Gly Arg Asn Thr Ile Lys Lys Gly Lys Tyr
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                                     235
Lys Ile Gly Asp Lys Glu Val Glu Tyr His Pro Lys Phe Arg Leu
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Ile Leu His Thr Lys Tyr Phe Asn Pro His Tyr Lys Pro Glu Met
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                                     265
Gln Ala Gln Cys Thr Leu Ile Asn Phe Leu Val Thr Arg Asp Gly
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                                     280
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Leu Glu Asp Gln Leu Leu Ala Ala Val Val Ala Lys Glu Arg Pro
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Asp Leu Glu Gln Leu Lys Ala Asn Leu Thr Lys Ser Gln Asn Glu
                305
                                     310
                                                          315
Phe Lys Ile Val Leu Lys Glu Leu Glu Asp Ser Leu Leu Ala Arg
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Leu Ser Ala Ala Ser Gly Asn Phe Leu Gly Asp Thr Ala Leu Val
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                                     340
Glu Asn Leu Glu Thr Thr Lys His Thr Ala Ser Glu Ile Glu Glu
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                350
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Lys Val Val Glu Ala Lys Ile Thr Glu Val Lys Ile Asn Glu Ala
                365
                                     370
                                                          375
Arg Glu Asn Tyr Arg Pro Ala Ala Glu Arg Ala Ser Leu Leu Tyr
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                380
Phe Ile Leu Asn Asp Leu Asn Lys Ile Asn Pro Val Tyr Gln Phe
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				395					400					405
Ser	Leu	Lys	Ala		Asn	Val	Val	Phe		Lys	Ala	Ile	Gln	
Thr	Thr	Pro	Ala	Asn 425	Glu	Val	Lys	Gln	Arg 430	Val	Ile	Asn	Leu	Thr 435
Asp	Glu	Ile	Thr		Ser	Val	Tyr	Met		Thr	Ala	Arg	Gly	
Ph∈	e Glu	Arg	Asp		Leu	Ile	Phe	Leu		Gln	Val	Thr	Phe	
Val	. Leu	ser	Met		Lys	Glu	Leu	Asn		Val	Glu	Leu	Asp	
Leu	l Leu	Arg	Phe	Pro	Phe	ГЛЗ	Ala	Gly	Val	Val	Ser	Pro	Val	Asp
Ph∈	Leu	Gln	His	485 Gln 500	Gly	Trp	Gly	Gly	490 Ile 505	Lys	Ala	Leu	Ser	
Met	: Asp	Glu	Phe		Asn	Leu	Asp	Ser		Ile	Glu	Gly	Ser	
Lys	Arg	Trp	Lys		Leu	Val	Glu	Ser		Ala	Pro	Glu	Lys	
Ile	Phe	Pro	Lys		Trp	Lуs	Asn	Lys	Thr	Ala	Leu	Gln	Lys	
Суз	Met	Val	Arg	Cys	Leu	Arg	Pro	Asp		Met	Thr	Tyr	Ala	
Lys	Asn	Phe	Va1		Glu	Lys	Met	Gly		Lys	Phe	Val	Glu	
Arg	ser ser	Val	Glu		Ser	Lys	Ser	Tyr		Glu	Ser	Ser	Pro	
Thr	Ser	Ile	Phe		Ile	Leu	ser	Pro		Val	Asp	Pro	Leu	
Asp	Val	Glu	Ala		Gly	Lys	Lys	Leu		Phe	Thr	Ile	qzA	
Gly	Lys	Leu	His		Val	Ser	Leu	Gly		Gly	Gln	Glu	Val	
Ala	Glu	Asn	Ala		Asp	Val	Ala	Ala		Lys	Gly	His	Trp	
Ile	Leu	Gln	Asn		His	Leu	Val	Ala		Trp	Leu	Gly	Thr	
Asp	Lys	Lys	Leu		Arg	Tyr	Ser	Thr		Ser	His	Glu	Asp	
Arg	yal	Phe	Ile	680 Ser 695	Ala	Glu	Pro	Ala		Ser	Pro	Glu	Thr	
Ile	lle	Pro	Gln	Gly	Ile	Leu	Glu	Asn	700 Ala 715	Ile	Lys	Ile	Thr	
Glu	Pro	Pro	Thr		Met	Tyr	Ala	Asn	Leu	His	Lys	Ala	Leu	
Leu	ı Phe	Thr	Gln		Thr	Leu	Glu	Met		Thr	Lys	Glu	Met	
Phe	Lys	Cys	Met	740 Leu 755	Phe	Ala	Leu	Суз	745 Tyr 760	Phe	His	Ala	Val	
Ala	Glu	Arg	Arg		Phe	Gly	Ala	Gln		Trp	Asn	Arg	Ser	765 Tyr 780
Pro	Phe	Asn	Asn		Asp	Leu	Thr	Ile	Ser	Ile	Asn	Val	Leu	Tyr
Asn	Tyr	Leu	Glu	Ala	Asn	Pro	Lys	Val		Trp	Asp	Asp	Leu	
Tyr	Leu	Phe	Gly		Ile	Met	Tyr	Gly		His	Ile	Thr	Asp	
Trp	Asp	Arg	Arg		Cys	Arg	Thr	Tyr		Ala	Glu	Tyr	Ile	
Thr	Glu	Met	Leu		Gly	Asp	Val	Leu		Ala	Pro	G1y	Phe	
Ile	Pro	Pro	Asn		Asp	Tyr	Lys	Gly		His	Glu	Tyr	Ile	
Glu	. Asn	Leu	Pro		Glu	Ser	Pro	Tyr		Tyr	Gly	Leu	His	
Asr	Ala	Glu	Ile		Phe	Leu	Thr	Val		Ser	Glu	Lys	Leu	
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Arg Thr Val Leu Glu Met Gln Pro Lys Glu Thr Asp Ser Gly Ala
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                                     910
Gly Thr Gly Val Ser Arg Glu Glu Lys Val Lys Ala Val Leu Asp
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                                     925
                                                          930
Asp Ile Leu Glu Lys Ile Pro Glu Thr Phe Asn Met Ala Glu Ile
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                                     940
Met Ala Lys Ala Ala Glu Lys Thr Pro Tyr Val Val Val Ala Phe
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                                     955
Gln Glu Cys Glu Arg Met Asn Ile Leu Thr Asn Glu Met Arg Arg
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                                     970
Ser Leu Lys Glu Leu Asn Leu Gly Leu Lys Gly Glu Leu Thr
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                980
                                     985
                                                          990
Thr Thr Asp Val Glu Asp Leu Ser Thr Ala Leu Phe Tyr Asp Thr
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                                    1000
                                                         1005
Val Pro Asp Thr Trp Val Ala Arg Ala Tyr Pro Ser Met Met Gly
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                                    1015
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Leu Ala Ala Trp Tyr Ala Asp Leu Leu Leu Arg Ile Arg Glu Leu
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Glu Ala Trp Thr Thr Asp Phe Ala Leu Pro Thr Thr Val Trp Leu
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                                    1045
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Ala Gly Phe Phe Asn Pro Gln Ser Phe Leu Thr Ala Ile Met Gln
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                                    1060
                                                         1065
Ser Met Ala Arg Lys Asn Glu Trp Pro Leu Asp Lys Met Cys Leu
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                                    1075
                                                         1080
Ser Val Glu Val Thr Lys Lys Asn Arg Glu Asp Met Thr Ala Pro
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                                    1090
Pro Arg Glu Gly Ser Tyr Val Tyr Gly Leu Phe Met Glu Gly Ala
                                    1105
               1100
Arg Trp Asp Thr Gln Thr Gly Val Ile Ala Glu Ala Arg Leu Lys
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                                    1120
                                                         1125
Glu Leu Thr Pro Ala Met Pro Val Ile Phe Ile Lys Ala Ile Pro
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                                    1135
                                                         1140
Val Asp Arg Met Glu Thr Lys Asn Ile Tyr Glu Cys Pro Val Tyr
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                                    1150
                                                         1155
Lys Thr Arg Ile Arg Gly Pro Thr Tyr Val Trp Thr Phe Asn Leu
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Leu Trp Gln Gly Ile Gly Val Gly Gln Leu Gln Leu Thr Glu Gly
                 35
                                      40
                                                           45
Phe Ala Leu Val Met Gln Gln Leu Pro Arg Ser Thr Lys Leu Lys
                 50
                                      55
Lys His Pro Arg Gly Glu Thr Glu Val Gly Ala Thr Ala Val Ala
                 65
                                      70
                                                           75
Phe Ser Ser Phe Asp Pro Arg Leu Phe Ile Leu Gly Thr Glu Gly
                 80
                                      85
Gly Phe Pro Leu Lys Cys Ser Leu Ala Ala Gly Glu Ala Ala Leu
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                                     1.00
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Thr Arg Met Pro Ser Ser Val Pro Leu Arg Ala Pro Ala Gln Phe
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                                     115
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Thr Phe Ser Pro His Gly Gly Pro Ile Tyr Ser Val Ser Cys Ser
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                                     130
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Pro Phe His Arg Asn Leu Phe Leu Ser Ala Gly Thr Asp Gly His
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Val His Leu Tyr Ser Met Leu Gln Ala Pro Pro Leu Thr Ser
                                                         Len
                155
                                     160
                                                          165
Gln Leu Ser Leu Lys Tyr Leu Phe Ala Val Arg Trp Ser Pro Val
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                                     175
Arg Pro Leu Val Phe Ala Ala Ala Ser Gly Lys Gly Asp Val Gln
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                                     190
Leu Phe Asp Leu Gln Lys Ser Ser Gln Lys Pro Thr Val Leu
                                                          Ile
                200
                                     205
                                                          210
Lys Gln Thr Gln Asp Glu Ser Pro Val Tyr Cys Leu Glu Phe Asn
                215
                                     220
                                                          225
Ser Gln Gln Thr Gln Leu Leu Ala Ala Gly Asp Ala Gln Gly
                                                          Thr
                                     235
                230
                                                          240
Val Lys Val Trp Gln Leu Ser Thr Glu Phe Thr Glu Gln Gly
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Asn Gly Lys Asp Pro Asp Ser Ser Ser Lys Val Leu Glu Leu Leu
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Leu Ala Phe Cys Ser Val Thr Gln Leu Arg His Met Leu Thr Gln
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Met Met Phe Glu Gln Ser Pro Pro Gly Ser Ala Thr Leu Gly Ser
                 65
                                      70
His Thr Lys Cys Leu Glu Pro Thr Val Ala Leu Leu Arg Trp Leu
                 80
                                      85
Ser Gln Pro Leu Asp Gly Ser Glu Asn Cys Ser Val Leu Ala Leu
                 95
                                     100
                                                          105
Glu Leu Phe Lys Glu Ile Phe Glu Asp Val Ile Asp Ala Ala Asn
                110
                                     115
Cys Ser Ser Ala Asp Arg Phe Val Thr Leu Leu Pro Thr Ile
                125
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Leu Asp Gln Leu Gln Phe Thr Glu Gln Asn Leu Asp Glu Ala Leu
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Thr Arg Gln Lys Cys Glu Arg Ile Ala Lys Ala Phe Glu Val Leu
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Leu Thr Leu Cys Gly Asp Asp Thr Leu Lys Met His Ile Ala Lys
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Ile Leu Thr Thr Val Lys Cys Thr Thr Leu Ile Glu Gln Gln Phe
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Thr Tyr Gly Lys Ile Asp Leu Gly Phe Gly Thr Lys Val Ala Asp
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                                     205
Ser Glu Leu Cys Lys Leu Ala Ala Asp Val Ile Leu Lys Thr Leu
                215
                                     220
                                                          225
Asp Leu Ile Asn Lys Leu Lys Pro Leu Val Pro Gly Met Glu Val
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                                     235
                                                          240
Ser Phe Tyr Lys Ile Leu Gln Asp Pro Arg Leu Ile Thr Pro Leu
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Ala Phe Ala Leu Thr Ser Asp Asn Arg Glu Gln Val Gln Ser Gly
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Leu Arg Ile Leu Leu Glu Ala Ala Pro Leu Pro Asp Phe Pro Ala

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275
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Leu Val Leu Gly Glu Ser Ile Ala Ala Asn Asn Ala Tyr Arg Gln
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Gln Glu Thr Glu His Ile Pro Arg Lys Met Pro Trp Gln Ser Ser
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                                     310
                                                          315
Asn His Ser Phe Pro Thr Ser Ile Lys Cys Leu Thr Pro His Leu
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                320
Lys Asp Gly Val Pro Gly Leu Asn Ile Glu Glu Leu Ile Glu Lys
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                                     340
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Leu Gln Ser Gly Met Val Val Lys Asp Gln Ile Cys Asp Val Arg
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Ile Ser Asp Ile Met Asp Val Tyr Glu Met Lys Leu Ser Thr
                                                         Leu
                365
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Ala Ser Lys Glu Ser Arg Leu Gln Asp Leu Leu Glu Thr Lys Ala
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Leu Ala Leu Ala Gln Ala Asp Arg Leu Ile Ala Gln His Arg Cys
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Gln Arg Thr Gln Ala Glu Thr Glu Ala Arg Thr Leu Ala Ser Met
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Leu Arg Glu Val Glu Arg Lys Asn Glu Glu Leu Ser Val Leu Leu
                425
                                     430
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Lys Ala Gln Gln Val Glu Ser Glu Arg Ala Gln Ser Asp Ile Glu
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His Leu Phe Gln His Asn Arg Lys Leu Glu Ser Val Ala Glu Glu
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His Glu Ile Leu Thr Lys Ser Tyr Met Glu Leu Leu Gln Arg Asn
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Glu Ser Thr Glu Lys Lys Asn Lys Asp Leu Gln Ile Thr Cys Asp
                485
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Ser Leu Asn Lys Gln Ile Glu Thr Val Lys Lys Leu Asn Glu Ser
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Leu Lys Glu Gln Asn Glu Lys Ser Ile Ala Gln Leu Ile Glu Lys
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Glu Glu Gln Arg Lys Glu Val Gln Asn Gln Leu Val Asp Arg Glu
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His Lys Leu Ala Asn Leu His Gln Lys Thr Lys Val Gln Glu Glu
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Lys Ile Lys Thr Leu Gln Lys Glu Arg Glu Asp Lys Glu Glu Thr
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Ile Asp Ile Leu Arg Lys Glu Leu Ser Arg Thr Glu Gln Ile Arg
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                                     580
Lys Glu Leu Ser Ile Lys Ala Ser Ser Leu Glu Val Gln Lys Ala
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                                     595
Gln Leu Glu Gly Arg Leu Glu Glu Lys Glu Ser Leu Val Lys Leu
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                                     610
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Gln Gln Glu Glu Leu Asn Lys His Ser His Met Ile Ala Met Ile
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His Ser Leu Ser Gly Gly Lys Ile Asn Pro Glu Thr Val Asn Leu
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Ile	Ser	Ala	Asp	20 Asp 35	Ile	Ser	Ile	Leu	25 Asn 40	Glu	Arg	Val	Glu	30 Leu 45
Leu	Gln	Arg	Gln	Trp 50	Glu	Glu	Leu	Cys	His 55	Gln	Leu	Ser	Leu	Arg 60
Arg	Gln	Gln	Ile	Gly 65	Glu	Arg	Leu	Asn	Glu 70	Trp	Ala	Val	Phe	
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Lys	Val	Ser	Gln	Asn 95	Gly	Asp	Ile	Leu		Glu	Glu	Met	Ile	
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Glu	Lys	His	Ser	Thr 230	Gly	Val	Ala	Ser		Leu	Asn	Leu	Cys	
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Glu	Asp	Trp	Leu	Lys 305	Ser	Ser	Glu	Arg		Ala	Ala	Phe	Pro	
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Gln	Arg	Trp	Asp	Asn 380	Leu	Gln	Lys	Arg		Thr	Ser	Ile	Leu	
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Lys	Ile	Glu	Gln	Ile 455	Ile	Ala	Gln	Gly		Gln	Leu	Ile	Glu	
Ser	Glu	Pro	Leu	Asp	Ala	Ala	Ile	Ile	Glu	Glu	Glu	Leu	Asp	Glu
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Pro	Leu	Glu	Trp	Asp 575	His	Asp	Tyr	Asp		Ser	Arg	Asp	Leu	
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Cys	Gln	Gly	Phe	His 845	Glu	Met	Ser	His	Gly 850	Leu	Leu	Leu	Met	Leu 855
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Lys Lys Glu Ala Ile Ile Arg Ile Asn Val Gly Gly Lys Leu Leu
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Gln Leu Ala Ala Tyr
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Lys Lys Val Leu Glu Gly Asp Ser Val Lys Leu Glu Cys Gln Ile
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Tyr Leu Ala Leu Asn Gly Lys Gly Leu Asn Val Lys Gln Ala Phe
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Ala Asp Val Thr Leu Arg Ser Arg Trp Thr Asn Met Asn Ala Leu
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His Tyr Ala Ala Tyr Phe Asp Val Pro Asp Leu Val Arg Val Leu
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Leu	Glu	Pro	Leu	Asn 365	Phe	Pro	Asp	Pro		Tyr	Lys	Glu	Glu	
Glu	Asp	Gln	Asp	Ile 380	Gln	Gly	Glu	Ile		His	Pro	Asp	Gly	
Va1	Glu	Lys	Val	Tyr 395	Lys	Asn	Gly	Cys		Val	Ile	Leu	Phe	
Asn	Gly	Thr	Arg	Lys 410	Glu	Val	Ser	Ala		Gly	Lys	Thr	Ile	
Va1	Thr	Phe	Phe	Asn 425	Gly	Asp	Val	Lys		Val	Met	Pro	Asp	
Arg	Val	Ile	Tyr	Tyr 440	Tyr	Ala	Ala	Ala		Thr	Thr	His	Thr	
Tyr	Pro	Glu	Gly	Leu 455	Glu	Val	Leu	His		Ser	Ser	Gly	Gln	
Glu	Lys	His	Tyr	Pro 470	Asp	Gly	Arg	Lys		Ile	Thr	Phe	Pro	
Gln	Thr	Val	Lys	Asn 485	Leu	Phe	Pro	Asp		Gln	Glu	Glu	Ser	
Phe	Pro	Asp	Gly	Thr	Ile	Val	Arg	Val		Arg	Asp	Gly	Asn	

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Leu Ile Glu Phe Asn Asn Gly Gln Arg Glu Leu His Thr Ala Gln
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Phe Lys Arg Arg Glu Tyr Pro Asp Gly Thr Val Lys Thr Val
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Ala Asn Gly His Gln Glu Thr Lys Tyr Arg Ser Gly Arg Ile Arg
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Phe Arg Arg Ser Thr Val Val Phe His Thr Val Glu Lys Ser Arg
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Gln Lys Asn Pro Arg Ser Leu Cys Ile Gln Pro Gln Thr Ala Pro
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Asp Ala Leu Pro Pro Glu Lys Thr Leu Glu Leu Thr Gln Tyr Lys
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Thr Lys Cys Glu Asn Gln Ser Gly Phe Ile Leu Gln Leu Lys Gln
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Leu Leu Ala Cys Gly Asn Thr Lys Phe Glu Ala Leu Thr Val Val
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                                     100
Ile Gln His Leu Leu Ser Glu Arg Glu Glu Ala Leu Lys Gln His
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                                     115
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Lys Thr Leu Ser Gln Glu Leu Val Asn Leu Arg Gly Glu Leu Val
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Thr Ala Ser Thr Thr Cys Glu Lys Leu Glu Lys Ala Arg Asn Glu
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Leu Gln Thr Val Tyr Glu Ala Phe Val Gln Gln His Gln Ala Glu
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Lys Thr Glu Arg Glu Asn Arg Leu Lys Glu Phe Tyr Thr Arg Glu
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Tyr Glu Lys Leu Arg Asp Thr Tyr Ile Glu Glu Ala Glu Lys
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Lys Met Gln Leu Gln Glu Gln Phe Asp Asn Leu Asn Ala Ala His
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Glu Thr Ser Lys Leu Glu Ile Glu Ala Ser His Ser Glu Lys Leu
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Glu Leu Leu Lys Lys Ala Tyr Glu Ala Ser Leu Ser Glu Ile Lys
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Lys Gly His Glu Ile Glu Lys Lys Ser Leu Glu Asp Leu Leu Ser
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Glu Lys Gln Glu Ser Leu Glu Lys Gln Ile Asn Asp Leu Lys Ser
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                                                          270
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Glu Asn Asp Ala Leu Asn Glu Lys Leu Lys Ser Glu Glu Gln Lys
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Arg Arg Ala Arg Glu Lys Ala Asn Leu Lys Asn Pro Gln Ile Met
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                                     295
Tyr Leu Glu Glu Leu Glu Ser Leu Lys Ala Val Leu Glu Ile
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                                     310
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Lys Asn Glu Lys Leu His Gln Gln Asp Ile Lys Leu Met Lys Met
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Glu Lys Leu Val Asp Asn Asn Thr Ala Leu Val Asp Lys Leu Lys
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                                     340
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Arg Phe Gln Glu Asn Glu Glu Leu Lys Ala Arg Met Asp Lys
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His Met Ala Ile Ser Arg Gln Leu Ser Thr Glu Gln Ala Val Leu
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Gln Glu Ser Leu Glu Lys Glu Ser Lys Val Asn Lys Arg Leu Ser
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Met Glu Asn Glu Glu Leu Leu Trp Lys Leu His Asn Gly Asp Leu
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Cys Ser Pro Lys Arg Ser Pro Thr Ser Ser Ala Ile Pro Leu Gln
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Val Val Asp Glu Gln Ala Asn Ser Ala Ala Leu Lys Glu Gln Leu
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Lys Met Lys Asp Gln Ser Leu Arg Lys Leu Gln Gln Glu Met Asp
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Ser Leu Thr Phe Arg Asn Leu Gln Leu Ala Lys Arg Val Glu Leu
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Leu Gln Asp Glu Leu Ala Leu Ser Glu Pro Arg Gly Lys Lys Asn
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Lys Lys Ser Gly Glu Ser Ser Gln Leu Ser Gln Glu Gln Lys
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Ser Val Phe Asp Glu Asp Leu Gln Lys Lys Ile Glu Glu Asn Glu
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Arg Leu His Ile Gln Phe Phe Glu Ala Asp Glu Gln His Lys His
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Val Glu Ala Glu Leu Arg Ser Arg Leu Ala Thr Leu Glu Thr Glu
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Ala Ala Gln His Gln Ala Val Val Asp Gly Leu Thr Arg Lys
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Met Glu Thr Ile Glu Lys Leu Gln Asn Asp Lys Ala Lys Leu Glu
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Val Lys Ser Gln Thr Leu Glu Lys Glu Ala Lys Glu Cys Arg Leu
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Arg Thr Glu Glu Cys Gln Leu Gln Leu Lys Thr Leu His Glu Asp
                200
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Leu Ser Gly Arg Leu Glu Glu Ser Leu Ser Ile Ile Asn Glu Lys
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Val Pro Phe Asn Asp Thr Lys Tyr Ser Gln Tyr Asn Ala Leu Asn
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Val Pro Leu His Asn Arg Arg His Gln Leu Lys Met Arg Asp Ile
                245
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Ala Gly Gln Ala Leu Ala Phe Val Gln Asp Leu Val Thr Ala Leu
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Leu Asn Phe His Thr Tyr Thr Glu Gln Arg Ile Gln Ile Phe Pro
                275
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Val Asp Ser Ala Ile Asp Thr Ile Ser Pro Leu Asn Gln Lys Phe
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Ser Gln Tyr Leu His Glu Asn Ala Ser Tyr Val Arg Pro Leu Glu
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Glu Gly Met Leu His Leu Phe Glu Ser Ile Thr Glu Asp Thr Val
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320
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Thr Val Leu Glu Thr Thr Val Lys Leu Lys Thr Phe Ser Glu His
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Leu Thr Ser Tyr
                Ile Cys Phe Leu Arg Lys Ile Leu Pro Tyr Gln
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Leu Lys Arg
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Gln Asp Tyr Leu Asn Arg Pro Arg Pro Thr Trp Glu Glu Val
                                                         Lys
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Glu Gln Leu Glu Lys Lys Lys Gly Ser Lys Ala Leu Ala Glu
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Phe Glu Glu Lys Met Asn Glu Asn Trp Lys Lys Glu Leu Glu Lys
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                 65
                                      70
His Arg Glu Lys Leu Leu Ser Gly Ser Glu Ser Ser Lys Lys
                 80
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Arg Gln Arg Lys Lys Glu Lys Lys Lys Ser Gly Arg Tyr
                                                         Ser
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                                     100
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Ser Ser Ser Ser Ser Ser Asp Ser Ser Ser Ser Ser Ser Asp
                110
                                     115
                                                         120
Ser Glu Asp Glu Asp Lys Lys Gln Gly Lys Arg Arg Lys Lys
                125
                                     130
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Lys Asn Arg Ser His Lys Ser Ser Glu Ser Ser Met Ser Glu
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Glu Ser Asp Ser Lys Asp Ser Leu Lys Lys Lys Lys Ser
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Asp Gly Thr Glu Lys Glu Lys Asp Ile Lys Gly Leu Ser Lys
                                                         Lys
                170
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Arg Lys Met Tyr Ser Glu Asp Lys Pro Leu Ser Ser Glu Ser Leu
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Ser Glu Ser Glu Tyr Ile Glu Glu Val Arg Ala Lys Lys
                                                         Lуs
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Ser Ser Glu Glu Arg Glu Lys Ala Thr Glu Lys Thr Lys Lys
                                                         Lys
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Lys Lys His Lys Lys His Ser Lys Lys Lys Lys Lys Ala Ala
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Ser Ser Ser Pro Asp Ser Pro
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Trp Ala Ala Phe Pro Ala Val Ser Gly Val His Leu Ser Pro Ser
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Ser Pro Glu Ile Val Leu Asp Arg Asp His Ser Ser Ser Ile Gly
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Cys Leu Ser Ser Asp Ala Ile Ile Ser Ser Pro Glu Asn Thr His
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                                      70
Ala Ala Asn Ser Ile Val Ser Gln Thr Ile Pro Lys Ala Gln Ile
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Gln Gln Ser Thr His Thr His Leu Asp Ile Ser Leu Phe Pro Leu
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Gly Leu Thr Asp Glu Lys Ser Asn Gly Thr Ile Ala Leu Val Asp
                110
                                     115
Asp Ser Glu Asp Pro Gly Ala Asn Val Ser Asn Ile Gln Leu Gln
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                                                          135
Gln Lys Ile Ser Ser Leu Glu Ile Lys Leu Lys Val Ser Glu Glu
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Glu Lys Gln Arg Ile Lys Gln Asp Val Glu Ser Leu Met Glu Lys
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                                     160
His Asn Val Leu Glu Lys Gly Phe Leu Lys Glu Lys Glu Gln Glu
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                 170
Ala Ile Ser Phe Gln Asp Arg Tyr Lys Glu Leu Gln Glu Lys His
                185
                                     190
Lys Gln Glu Leu Glu Asp Met Arg Lys Ala Gly His Glu Ala Leu
                200
                                     205
                                                          210
Ser Ile Ile Val Asp Glu Tyr Lys Ala Leu Leu Gln Ser Ser Val
                215
                                     220
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Lys Gln Gln Val Glu Ala Ile Glu Lys Gln Tyr Ile Ser Ala Ile
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Glu Lys Gln Ala His Lys Cys Glu Glu Leu Leu Asn Ala Gln His
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Gln Arg Leu Leu Glu Met Leu Asp Thr Glu Lys Glu Leu Leu Lys
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Glu Lys Ile Lys Glu Ala Leu Ile Gln Gln Ser Gln Glu Gln Lys
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                                     280
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Glu Ile Leu Glu Lys Cys Leu Glu Glu Glu Arg Gln Arg Asn Lys
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                                     295
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Glu Ala Leu Val Ser Ala Ala Lys Leu Glu Lys Glu Ala Met Lys
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Asp Ala Val Leu Lys Val Val Glu Glu Glu Arg Lys Asn Leu Glu
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                                     325
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Lys Ala His Ala Glu Glu Arg Glu Leu Trp Lys Thr Glu His Ala
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Lys Asp Gln Glu Lys Val Ser Gln Glu Ile Gln Lys Ala Ile Gln
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Glu Gln Arg Lys Ile Ser Gln Glu Thr Val Lys Ala Ala Ile Ile
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Glu Glu Gln Lys Arg Ser Glu Lys Ala Val Glu Glu Ala Val Lys
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Arg Thr Arg Asp Glu Leu Ile Glu Tyr Ile Lys Glu Gln Lys Arg
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                                     400
Leu Asp Gln Val Ile Arg Gln Arg Ser Leu Ser Ser Leu Glu Leu
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Asp Thr Ala Asp Ala Val Ala Ala Glu Gly Ala Tyr Tyr Leu Glu
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Gln Val Thr Ile Thr Glu Ala Ser Glu Asp Asp Tyr Glu Tyr Glu
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Glu Ile Pro Asp Asp Asn Phe Ser Ile Pro Glu Gly Glu Glu Asp
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Leu Ala Lys Ala Ile Gln Met Ala Gln Glu Gln Ala Thr Asp Thr
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                                      85
Glu Ile Leu Glu Arg Lys Thr Val Leu Pro Ser Lys His Ala Val
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                                     100
                                                          105
Pro Glu Val Ile Glu Asp Phe Leu Cys Asn Phe Leu Ile Lys Met
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                                     115
Gly Met Thr Arg Thr Leu Asp Cys Phe Gln Ser Glu Trp Tyr Glu
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Leu Ile Gln Lys Gly Val Thr Glu Leu Arg Thr Val Gly Asn Val
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                                     145
Pro Asp Val Tyr Thr Gln Ile Met Leu Leu Glu Asn Glu Asn Lys
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Asn Leu Lys Lys Asp Leu Lys His Tyr Lys Gln Ala Ala Glu Tyr
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Gln Cys Phe Thr Cys Arg Thr Cys Arg Arg Gln Leu Ala Gly Gln
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Ser Phe Tyr Gln Lys Asp Gly Arg Pro Leu Cys Glu Pro Cys Tyr
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Gln Asp Thr Leu Glu Arg Cys Gly Lys Cys Gly Glu Val Val Arg
                 8.0
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Asp His Ile Ile Arg Ala Leu Gly Gln Ala Phe His Pro Ser Cys
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Phe Thr Cys Val Thr Cys Ala Arg Cys Ile Gly Asp Glu Ser
                                                         Phe
                110
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Ala Leu Gly Ser Gln Asn Glu Val Tyr Cys Leu Asp Asp Phe
                                                         Tyr
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Arg Lys Phe Ala Pro Val Cys Ser Ile Cys Glu Asn Pro Ile Ile
Pro Arg Asp Gly Lys Asp Ala Phe Lys Ile Glu Cys Met Gly Arg
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Asn Phe His Glu Asn Cys Tyr Arg Cys Glu Asp Cys Arg Ile Leu
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Leu Ser Val Glu Pro Thr Asp Gln Gly Cys Tyr Pro Leu Asn Asn
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His Leu Phe Cys Lys Pro Cys His Val Lys Arg Ser Ala Ala Gly
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Cys Cys

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Asn Val Ile Asp Arg Met Lys Glu Ser Ser Pro Ser Gly Ser Lys
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Ser Gln Arg Tyr Ser Gly Ala Tyr Gly Ala Ser Val Ser Asp Glu
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Glu Leu Lys Arg Arg Val Ala Glu Glu Leu Ala Leu Glu Gln Ala
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Lys Lys Glu Ser Glu Asp Gln Lys Arg Leu Lys Gln Ala Lys Glu
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Leu Asp Arg Glu Arg Ala Ala Asn Glu Gln Leu Thr Arg Ala
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Ile Leu Arg Glu Arg Ile Cys Ser Glu Glu Glu Arg Ala Lys Ala
                110
                                     115
Lys His Leu Ala Arg Gln Leu Glu Glu Lys Asp Arg Val Leu Lys
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Lys Gln Asp Ala Phe Tyr Lys Glu Gln Leu Ala Arg Leu Glu Glu
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                                     145
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Arg Ser Ser Glu Phe Tyr Arg Val Thr Thr Glu Gln Tyr Gln Lys
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Ala Ala Glu Glu Val Glu Ala Lys Phe Lys Arg Tyr Glu Ser His
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                                     175
                                                          180
Pro Val Cys Ala Asp Leu Gln Ala Lys Ile Leu Gln Cys Tyr Arg
                185
                                     190
                                                          195
Glu Asn Thr His Gln Thr Leu Lys Cys Ser Ala Leu Ala Thr Gln
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Tyr Met His Cys Val Asn His Ala Lys Gln Ser Met Leu Glu Lys
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Tyr Ala Leu Leu Gly Asn Tyr Asp Ser Ser Met Val Tyr Tyr Gln
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Gly Val Met Gln Gln Ile Gln Arg His Cys Gln Ser Val Arg Asp
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Pro Ala Ile Lys Gly Lys Trp Gln Gln Val Arg Gln Glu Leu Leu
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Glu Glu Tyr Glu Gln Val Lys Ser Ile Val Ser Thr Leu Glu Ser
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Phe Lys Ile Asp Lys Pro Pro Asp Phe Pro Val Ser Cys Gln Asp
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Glu Pro Phe Arg Asp Pro Ala Val Trp Pro Pro Pro Val Pro Ala
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Glu His Arg Ala Pro Pro Gln Ile Arg Arg Pro Asn Arg Glu Val
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Arg Pro Leu Arg Lys Glu Met Ala Gly Val Gly Ala Arg Gly Pro
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Val Gly Arg Ala His Pro Ile Ser Lys Ser Glu Lys Pro Ser
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Ser Arg Asp Lys Asp Tyr Arg Ala Arg Gly Arg Asp Asp Lys Gly
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Arg Lys Asn Met Gln Asp Gly Ala Ser Asn Gly Glu Met Pro Lys
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Phe Asp Gly Ala Gly Tyr Asp Lys Asp Leu Val Glu Ala Leu Glu
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Arg Asp Ile Val Ser Arg Asn Pro Ser Ile His Trp Asp Asp Ile
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Ala Asp Leu Glu Glu Ala Lys Lys Leu Leu Arg Glu Ala Val Val
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Leu Pro Met Trp Met Pro Asp Phe Phe Lys Gly Ile Arg Arg Pro
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Trp Lys Gly Val Leu Met Val Gly Pro Pro Gly Thr Gly Lys Thr
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Met Leu Ala Lys Ala Val Ala Thr Glu Cys Gly Thr Thr Phe Phe
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Asn Val Ser Ser Ser Thr Leu Thr Ser Lys Tyr Arg Gly Glu Ser
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Glu Lys Leu Val Arg Leu Leu Phe Glu Met Ala Arg Phe Tyr
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Pro Thr Thr Ile Phe Ile Asp Glu Ile Asp Ser Ile Cys Ser Arg
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Arg Gly Thr Ser Asp Glu His Glu Ala Ser Arg Arg Val Lys Ser
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Glu Leu Leu Ile Gln Met Asp Gly Val Gly Gly Ala Leu Glu Asn
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Asp Asp Pro Ser Lys Met Val Met Val Leu Ala Ala Thr Asn Phe
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Pro Trp Asp Ile Asp Glu Ala Leu Arg Arg Arg Leu Glu Lys Arg
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Ile Tyr Ile Pro Leu Pro Thr Ala Lys Gly Arg Ala Glu Leu Leu
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Lys Ile Asn Leu Arg Glu Val Glu Leu Asp Pro Asp Ile Gln Leu
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Glu Asp Ile Ala Glu Lys Ile Glu Gly Tyr Ser Gly Ala Asp Ile
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Thr Asn Val Cys Arg Asp Ala Ser Leu Met Ala Met Arg Arg
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Ile Asn Gly Leu Ser Pro Glu Glu Ile Arg Ala Leu Ser Lys Glu
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Glu Leu Gln Met Pro Val Thr Lys Gly Asp Phe Glu Leu Ala Leu
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Lys Lys Ile Ala Lys Ser Val Ser Ala Ala Asp Leu Glu Lys Tyr
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Glu Lys Trp Met Val Glu Phe Gly Ser Ala
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Lys Ala Gly Thr Ala Gly Trp Asn Glu Pro Gln Met Val Phe Pro
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Tyr Ala Arg Lys Arg Val Ser Leu Gly Ile Asp Ile Cys His Pro
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Asp Thr Phe Ser Tyr
                    Pro Ile Glu Arg Gly Arg Ile Leu Asn
                                                          Trp
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Glu Gly Val Gln Tyr
                    Leu Trp Ser Phe Val Leu Glu Asn His Arg
                 80
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Arg Glu Gln Glu Val Pro Pro Val Ile Ile Thr Glu Thr Pro Leu
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Arg Glu Pro Ala Asp Arg Lys Lys Met Ser Ser Leu Glu Thr Leu
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Gln Gly Thr Val Phe Pro Gly Trp Pro Ile Ile Gly Val
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Val Leu Arg Glu Thr Lys Ala Ile Lys Lys Ala Ile Thr Cys Gly
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Glu Lys Glu Lys Gln Asp Leu Ile Lys Ser Leu Ala Met Leu Lys
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Asp Gly Phe Cys Thr Asp Arg Gly Ser His Ser Asp Leu Trp Ser
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Ser Ser Ser Leu Glu Ser Ser Phe Pro Leu Pro Lys
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Tyr Leu Asp Val Ser Ser Gln Thr Asp Ile Ser Gly Ser Phe Gly
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Ile Asn Ser Asn Asn Gln Leu Ala Glu Lys Val Arg Leu Arg Leu
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Arg Tyr Glu Glu Ala Lys Arg Arg Ile Ala Asn Leu Lys Ile Gln
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Leu Ala Lys Leu Asp Ser Glu Ala Trp Pro Gly Val Leu Asp Ser
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Glu Arg Asp Arg Leu Ile Leu Ile Asn Glu Lys Glu Glu Leu Leu
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Lys Glu Met Arg Phe Ile Ser Pro Arg Lys Trp Thr Gln Gly Glu
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Val Glu Gln Leu Glu Met Ala Arg Lys Arg Leu Glu Lys Asp Leu
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Gln Ala Ala Arg Asp Thr Gln Ser Lys Ala Leu Thr Glu Arg Leu
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Lys Leu Asn Ser Lys Arg Asn Gln Leu Val Arg Glu Leu Glu Glu
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Ala Thr Arg Gln Val Ala Thr Leu His Ser Gln Leu Lys Ser
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Ser Ser Ser Met Gln Ser Leu Ser Ser Gly Ser Ser Pro Gly
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                230
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Leu Thr Ser Ser Arg Gly Ser Leu Val Ala Ser Ser Leu Asp
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Ser Thr Ser Ala Ser Phe Thr Asp Leu Tyr
                                         Tyr Asp Pro Phe Glu
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Gln Leu Asp Ser Glu Leu Gln Ser Lys Val Glu Phe Leu Leu
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Glu Gly Ala Thr Gly Phe Arg Pro Ser Gly Cys Ile Thr Thr
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His Glu Asp Glu Val Ala Lys Thr Gln Lys Ala Glu Gly Gly Gly
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Thr	Ser	Leu	Ser		Arg	Ser	Ser	Leu		Ser	Pro	Ser	Pro	Pro 345
Cys	Ser	Pro	Leu		Ala	Asp	Pro	Leu		Ala	Gly	Asp	Ala	
Leu	Asn	Ser	Leu		Phe	Glu	Asp	Pro		Leu	Ser	Ala	Thr	Leu 375
Cys	Glu	Leu	Ser		Gly	Asn	Ser	Ala		Glu	Arg	Tyr	Arg	
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Thr	Ala	Gln	Gly		Gly	Leu	Lys	Val		Cys	Val	ser	Ala	
Val	Ser	Asp	Glu	Ser 425	Val	Ala	Gly	Asp		Gly	Val	Tyr	Glu	
Ser	Val	Gln	Arg		Gly	Ala	Ser	Glu		Ala	Ala	Phe	Asp	
Asp	Glu	Ser	Glu	Ala 455	Val	Gly	Ala	Thr		Ile	Gln	Ile	Ala	
Lys	Tyr	Asp	Glu	Lys 470	Asn	Lys	Gln	Phe	Ala 475	Ile	Leu	Ile	Ile	Gln 480
Leu	Ser	Asn	Leu	Ser 485	Ala	Leu	Leu	Gln	Gln 490	Gln	Asp	Gln	Lys	
Asn	Ile	Arg	Val	Ala 500	Val	Leu	Pro	Cys	Ser 505	Glu	Ser	Thr	Thr	Cys 510
Leu	Phe	Arg	Thr	Arg 515	Pro	Leu	Asp	Ala	Ser 520	Asp	Thr	Leu	Val	Phe 525
Asn	Glu	Val	Phe	Trp 530	Val	Ser	Met	Ser	Tyr 535	Pro	Ala	Leu	His	Gln 540
Lys	Thr	Leu	Arg	Val 545	Asp	Val	Cys	Thr	Thr 550	Asp	Arg	Ser	His	Leu 555
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Arg	Ser	Gly	Glu	Arg 575	Ser	Thr	Arg	Trp	Tyr 580	Asn	Leu	Leu	Ser	Tyr 585
Lys	Tyr	Leu	Lys	Lys 590	Gln	Ser	Arg	Glu	Leu 595	Lys	Pro	Val	Gly	Val 600
Met	Ala	Pro	Ala	Ser 605	Gly	Pro	Ala	Ser	Thr 610	Asp	Ala	Val	Ser	Ala 615
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Gln	Tyr	Val	Cys	Arg 740	Leu	Asn	Arg	Ser	Asp 745	Ser	Asp	Ser	Ser	Thr 750
Leu	Ser	Lys	Lys	Pro 755	Pro	Phe	Va1	Arg	Asn 760	Ser	Leu	Glu	Arg	Arg 765
Ser	Val	Arg	Met	Lys 770	Arg	Pro	Ser	Ser	Val 775	Lys	Ser	Leu	Arg	Ser 780
Glu	Arg	Leu	Ile	Arg 785	Thr	Ser	Leu	Asp	Leu 790	Glu	Leu	Asp	Leu	
Ala	Thr	Arg	Thr		His	Ser	Gln	Leu		Gln	Glu	Ile	Ser	
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Glu Lys Glu Leu Pro Gln Trp Leu Arg Glu Asp Glu Arg Phe Arg
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Lys Asp Val His Arg Leu Arg Gly Gln Ser Cys Lys Glu Pro Pro
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Asp Asp Glu Asn Arg Val Asn Gln Glu Glu Lys Ala Ala Lys Ile
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Asn Pro Phe Asp Glu Pro Glu Ala Phe Val Thr Ile Lys Asp Ser
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Ser Lys Tyr Leu Tyr Ala Asp Ser Ser Lys Thr Glu Glu Glu
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Pro Asn Asn Leu Val Asn Pro Val Gln Glu Leu Glu Thr Glu Arg
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			Ser	470					475			_		480
			Thr	485			_	_	490	_		_		495
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			Asp	545					550			_		555
			Ile	560				_	565		_			570
	_		Gly	575				_	580	_				585
			Glu	590			_		595				_	600
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			Leu	620					625					630
•			Pro	635					640					645
			Lys	650					655					660
			Glu	665					670					675
			Leu	680					685					690
			Cys Gln	695	_				700			_		705
				710					715					720
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			Asn	740					745				Pro	750
			Val	755					760					765
				770					775					780
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	_	_	Ala	800					805					810
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                                    1045
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Phe Thr Phe Gly Gln Thr Gly Ser Gly Lys Thr Tyr Thr Leu Thr
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Glu Leu Leu Gln Thr Gly Leu Ser Arg Arg Arg Asn Ser Ala His
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Tyr Ile Ser Arg Gln Thr Ala Gln Gln Met Pro Ser Val Asp Pro
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Pro Phe Arg Asp Ser Lys Leu Thr Lys Leu Leu Ala Asp Ser Leu
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Ala Gln Cys Leu Pro Glu Thr Leu Ser Thr Leu Arg Tyr Ala Ser
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                                     430
Arg Leu Arg Lys Glu Lys Ser Gln Leu Gln Asn Ser Arg Glu Leu
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Glu Arg Arg Leu Leu Ser Ala Cys Tyr His His Gln Gln Gly Pro
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Cys Pro Leu Cys Arg Val Pro Leu Ala His Trp Gly Cys Leu Pro
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Thr Leu Ala Leu Glu Tyr Thr Tyr Gly Arg Arg Ala Lys Gly His
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Asn Thr Pro Lys Asp Ile Ala His Phe Trp Glu Leu Gly Gly
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Leu Arg Thr Phe Ser Leu Val Leu Val Leu Asp Leu Ser Lys Pro
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Ser His Val Asp Lys Val Ile Met Lys Leu Gly Lys Thr Asn Ala
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Lys Ala Val Ser Glu Met Arg Gln Lys Ile Trp Asn Asn Met Pro
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Lys Asp His Pro Asp His Glu Leu Ile Asp Pro Phe Pro Val Pro
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Leu Val Ile Ile Gly Ser Lys Tyr Asp Val Phe Gln Asp Phe Glu
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Ser Glu Lys Arg Lys Val Ile Cys Lys Thr Leu Arg Phe Val Ala
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His Tyr Tyr Gly Ala Ser Leu Met Phe Thr Ser Lys Ser Glu Ala
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Arg Val Arg Pro Leu Gly Phe Pro Asp Gln Glu Cys Cys Ile Glu
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Val Ile Asn Asn Thr Thr Val Gln Leu His Thr Pro Glu Gly
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Lys Gln Val Phe Gly Thr His Thr Thr Gln Lys Glu Leu Phe Asp
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Tyr	Val	Phe	Lys	Ser 155	Asn	Asp	Arg	Asn		Met	Asp	Ile	Gln	
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Asn	Pro	Lys	Thr		Ser	Ser	Lys	Arg	Gln 190	Val	Asp	Pro	Glu	
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Asp	Glu	Asp	Ser		Tyr	Gly	Val	Phe		Ser	Tyr	Ile	Glu	
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Lys	Asn	His	Asn	Met 260	Tyr	Val	Ala	Gly	Cys 265	Thr	Glu	Val	Glu	
Lys	Ser	Thr	Glu	Glu 275	Ala	Phe	Glu	Val	Phe 280	Trp	Arg	Gly	Gln	Lys 285
Lys	Arg	Arg	Ile	Ala 290	Asn	Thr	His	Leu	Asn 295	Arg	Glu	Ser	Ser	Arg 300
Ser	His	Ser	Val	Phe 305	Asn	Ile	Lys	Leu		Gln	Ala	Pro	Leu	
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Asn	Gln	Ser	Leu	Met 365	Thr	Leu	Arg	Thr		Met	Asp	Val	Leu	
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Ser	Lys	Leu	Thr	His 395	Leu	Phe	Lys	Asn	Tyr 400	Phe	Asp	Gly	Glu	Gly 405
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Pro	Ser	Суз	Glu		Leu	Asp	Ile	Asn		Glu	Gln	Thr	Leu	
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Met	Ile	Asp	Glu		Asn	Lys	Gln	Ser		Ala	Phe	Lys	Ala	
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Lys	Ile	Glu	Ile		Glu	Lys	Thr	Thr		Ile	Tyr	Glu	Glu	
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Val Lys Asp Glu Lys Leu Lys Gln Leu Lys Ala Ile Val Thr Glu
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Pro Lys Thr Glu Lys Pro Glu Arg Pro Ser Arg Glu Arg Asp Arg
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His Ala Ile Thr Val Ser Val Ala Asn Glu Lys Ala Leu Ala Lys
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Cys Glu Lys Tyr Met Leu Thr His Gln Glu Leu Ala Ser Asp Gly
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Glu Ile Glu Thr Lys Leu Ile Lys Gly Asp Ile Tyr Lys Thr Arg
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Ala Ala Asn Ser Cys Thr Ser Tyr Ser Gly Thr Thr Leu Asn Leu
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Ser	Arg	Ser	Arg	Gln	Thr	Pro	Ser	Pro	Asp	Val	Val	Leu	Arg	Gly	Ser
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Glu Glu Lys Glu Arg Glu Cys Pro Thr Val Ala Pro Ala His Ser 575	Val	Asp	Gly	Lys	Val	Ser	Val	Asn	Gly	Glu	Thr	Val	His	Arg	Glu
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Ala	Arg	Gly	Ala	Asn 170	Lys	Glu	His	Arg	Asn 175	Val	Ser	Asp	Tyr	Thr 180
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Cys	Glu	Leu	Leu	Ile 335	His	Arg	Gly	Ala	His 340	Ile	Asp	Val	Arg	
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Ile	Asn	Gly	Glu		Arg	Gly	Gly	Gly	Ala	Gly	Gly	Asn	Ser	
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Asn	Met	Ser	Trp	Ile 170	Arg	Gly	Arg	Arg		Phe	Ile	Phe	Lys	
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Leu	Thr	Leu	Asp	Leu 215	Met	Lys	Pro	Lys	Ser 220	Arg	Glu	Val	Glu	Arg 225
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			Val	275				_	280				His	285
			Glu	290	_	_	-		295	_	-			Leu 300
			Leu	305					310		_			315
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_			Ile	365		Ile			370					375 Ala
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			Val	395	_	_			400				Leu	405
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